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PII: S0956-5663(20)30712-0

DOI: <https://doi.org/10.1016/j.bios.2020.112724>

Reference: BIOS 112724



To appear in: *Biosensors and Bioelectronics*

Received Date: 29 April 2020

Revised Date: 7 October 2020

Accepted Date: 12 October 2020

Please cite this article as: Masurkar, N., Thangavel, N.K., Yurgelevic, S., Varma, S., Auner, G.W., Reddy Arava, L.M., Reliable and highly sensitive biosensor from suspended MoS₂ atomic layer on nano-gap electrodes, *Biosensors and Bioelectronics* (2020), doi: <https://doi.org/10.1016/j.bios.2020.112724>.

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Author contributions

NIRUL MASURKAR, GREGORY W AUNER, and LEELA MOHANA REDDY ARAVA: conceived and designed the experiments; **NIRUL MASURKAR** and **SUNDEEP VARMA:** fabricated and characterized materials and devices, **NIRUL MASURKAR** and **SUNDEEP VARMA:** electrical characterization; **NIRUL MASURKAR, SALLY YURGELEVIC,** and **NARESH KUMAR THANGAVEL:** performed the biological experiments; **NIRUL MASURKAR, SUNDEEP VARMA, GREGORY W AUNER,** and **LEELA MOHANA REDDY ARAVA:** analyzed the data; **NIRUL MASURKAR, NARESH KUMAR THANGAVEL , SALLY YURGELEVIC:** wrote the paper.

Reliable and highly sensitive biosensor from suspended MoS₂ atomic layer on nano-gap electrodes

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Abstract

The uneven morphology and the trapped charges at the surface of the traditionally used supporting substrate-based 2D biosensors produces a scattering phenomenon, which leads to irregular signal from individually fabricated device. Though suspended 2D channel material has the potential to overcome scattering effects from the substrates but achieving reliability and selectivity, have been limiting the biosensor technology. Here, we have demonstrated nanogap electrodes fabrication by using the self-assembly technique, which provides suspension to the 2D-MoS₂. These nano-spacing electrodes not only give suspension but also provide robust strength to the atomic layer, which remains freestanding after coating of the Hafnium oxide (HfO₂) as well as linkers and antibodies. For evaluating the electrical characteristics of suspended MoS₂ FET, gating potential was applied through electrolyte on suspended MoS₂ transistor, which achieved lower subthreshold swing 70 mV/dec and ON/OFF ratio 10⁷. Afterward, pH detection has demonstrated at room temperature, which shows an impressive sensitivity of ~880 by changing 1 unit of pH. Besides, we have successfully shown Escherichia coli (*E. coli*) bacteria sensing from suspended MoS₂ transistor by functionalizing dielectric layer with *E. coli* antibodies. The reported biosensor has shown the ~9% of conductance changes with a lower concentration of *E. coli* (10 CFU/mL; colony-forming unit per mL) as well as maintain the constant sensitivity in three fabricated devices. This type of the architecture has a potential to detect range of biomolecules such as COVID-19 viruses also by altering the oxide surface.

Keywords: Biosensor, Field Effect transistors, Escherichia coli, MoS₂, Nanogap

1.Introduction

Due to their potential of providing continuous and real-time information, biosensors are prominent devices in the medical field and garnering substantial interest in other domains such as forensic industries, national security, and ecological monitoring (Mehrotra and research 2016). Specifically, electrochemical field-effect transistor (FET)-based biosensors have been recognized as one of the promising candidates because of their compatibility with electronic devices, low power consumption, label-free detection of specific biomolecules, and low-cost mass production (Kim et al. 2016; Kuriyama and Kimura 1991; Suvarnaphaet and Pechprasarn 2017). In FET based biosensor, the charged biomolecules generate the electrostatic effect and bring the variation in the conductivity of the channel, which can easily measure by transistor characteristics such as source to drain current (Kaisti 2017; Sarkar et al. 2014a). To obtain better electrostatic effects from charged biomolecules, many semiconducting materials have been employed (Holzinger et al. 2014; Li et al. 2017; Zhu et al. 2012). Among the various materials, two-dimensional (2D) semiconducting materials are attracting much attention because of their tunable bandgap, a high surface to volume ratio that provides higher sensitivity for detection.

In the 2D domain, transition metal dichalcogenides (TMDs) like MoS₂ has gained much attention in the field of FET-biosensor due to its direct bandgap, biocompatibility, and high mobility (Kalantar-zadeh and Ou 2015b). Despite these merits, the performance and consistency of such atomic layer crystals are easily affected by supporting substrate interaction (Chen et al. 2018). The interaction of supporting substrate with atomic layers of MoS₂ degrade the transport properties by scattering, which implies the interface control is vital for the performance and reliability of biosensor devices (Du et al. 2008). For instance, the surface of silicon dioxide (SiO₂) substrate is highly disordered as well as chemically active due to the trapped atmospheric gases,

chemical adsorbates, and unknown functional groups (Zhang et al. 2009). Therefore, transferring another layer of MoS₂ on the top of SiO₂ or any other insulating substrate cannot contribute to carrier charge transport distinctly, which leads to the unreliable output of every single device. In recent years, many efforts have been employed to enhance the quality of the substrate, such as active layer interface by using Poly(methyl methacrylate) (PMMA) and polymer electrolytes [9]. These layers elude chemical bonding or surface roughness and improve carrier transport properties, but they cannot be employed in biosensor applications due to the fabrication and reproducibility issues. The other approach is creating suspension of an atomic layer in between electrodes to enhance the carrier transport and remove scattering effect by wet etching of the SiO₂ layer underneath the monolayer MoS₂. Freestanding MoS₂ has shown better performance than the supporting on the SiO₂ substrate in terms of back gating electronic conduction (Jin et al. 2013). However, the existing SiO₂ requires hazardous chemical etchants such as hydrofluoric acid (HF), which is difficult to handle and affects the 2D film structure and purity (Zhang et al. 2017). Secondly, freestanding MoS₂ sags between the two electrodes because of the large spacing ($\sim 2 \mu\text{m}$), which makes it impossible to coat dielectric layers such as hafnium oxide (HfO₂) and antibodies. Therefore, this structure impedes making top gate FET biosensors, which allows only back gating. However, the back-gate FET requires more power (input gate voltage) to turn ON the device than the top gate, which hinders making a low power and highly sensitive biosensor. In FET based biosensor, the sensitivity is inversely proportional to the subthreshold swing (SS), which is defined as a change in the current with respective dielectric surface potential ($SS = dV_{gate}/d \log_{10} I_{drain}$; dV_{gate} and I_{drain} are a change in gate potential and drain current respectively). Thus, in back gate requires higher input voltage to turn on the device than the top gate, which reduces the sensitivity of detection (Sarkar et al. 2014a; Sarkar et al. 2014b).

Given the importance of reliability and sensitivity of FET based biosensor, in this work, chemical vapor deposition (CVD) grown MoS_2 is transferred by using novel dry stamping method on self-assembled photolithographically patterned nano-gaps to achieve suspension. Fabricated nanogap electrodes have 70 to 90 nm spacing, which provides mechanical strength to the MoS_2 monolayer and does not allow to sag between the two electrodes. This robustness in the monolayer film permits the coating of another thin dielectric layer as well as linkers/antibodies without interacting, supporting the substrate. HfO_2 as a dielectric material was considered, which is easily functionalized by a linker (3-Aminopropyl)triethoxysilane (APTES), glutaraldehyde, and *E. coli* antibodies to bring variation to the suspended 2D MoS_2 by targeting a charged *E. coli* bacteria. The suspended FET architecture lowered down the subthreshold swing (SS) to 70 mV/dec by terminating the scattering phenomenon and enhance the sensitivity of the pH as well as *E. coli* detection (Liao et al. 2018). Freestanding nature of 2D channel material i.e., MoS_2 , retains its intrinsic properties and provides reliability and consistency in every single device.

2.Experimental section

2.1. CVD synthesis. Experimental procedures of CVD (In-house built) grown MoS_2 are as follows: quartz boat consists of MoO_3 powder, and SiO_2/Si pieces (top of the powder) were positioned in the middle of furnace whereas Sulphur boat placed at the upstream of lower temperature zone. In the typical process flow of CVD grown MoS_2 , Ar gas was passed for 20 mins at 250 sccm to wash out all environmental gases and then ramp up the temperature till 750°C in 15 mins. Afterward, the temperature was maintained at 750°C for 20 mins. In this condition, Sulphur starts evaporating and reduced MoO_3 to convert into metallic MoO_2 . More sulphurization of MoO_2 leads towards the triangular shape of MoS_2 on the SiO_2/Si pieces.

2.2. Photolithography of nanogaps. In this process, the self-assembly technique was adapted to achieve nanogaps between the metal electrodes. Here, few parameters were optimized while fabrication such as lift-off resists thickness, rpm rate of spinning photoresist, baking, developing the photoresist, avoiding the residues during fabrication, and most important optimization thickness of metal electrode layer. The optical photolithography technique of patterning the substrate was done by coating the first positive photoresist (MICROPOSIT S1800) and expose (SUSS MicroTec Mask Aligner MA6) it for 80 mJ/cm^2 (flood exposure). Then another layer of photoresist 1811 is coated on the top and pattern it with design chrome mask followed by evaporation of the Ti/Au/Cr (thickness is 10 nm/ 50 nm/ 80 nm). After patterning three metal layers Ti/Au/Cr, wafer kept for 12 Hrs. to control thermal oxidation of the chromium to achieve nanogap. Nano-spacing between electrodes rely on the rate of oxidation of the Cr layer, where temperature controls the rate of the oxidation. Therefore, it is essential to maintain the temperature as well as environmental gases of the wafer constant throughout the oxidation process. To achieve the proper pattern of the second layer, the thickness of the Ti/Au (second layer, whose thickness is 10 nm/ 50 nm) should not exceed more than 50% of chromium thickness. Liftoff the chromium as well as the second Au electrode is processed in chromium etchant solution (Chromium etchant, Sigma Aldrich).

2.3. Electrical Characterization. Nano-gaps cannot be seen via microscope; therefore, before transferring the 2D material, it was characterized by the current-voltage method. I-V curve between two electrodes was measured by the source meter (Keysight B2912A).

2.4. Transferring of 2D material and coating of the HfO_2 . After achieving the optimum nanogap electrodes, triangular shape MoS_2 was transferred on the top of the spacing by using a novel dry stamping method. This method provides proper placement of CVD grown MoS_2 in the

desired area. Atomic layer deposition (ALD) from the University of Michigan, Ann Arbor (Veeco Fiji ALD), was considered to deposit 25 nm of the high dielectric layer.

2.5. Raman and AFM Characterization. As prepared samples of MoS₂ are characterized by Raman Spectroscopy using 514 nm laser, whereas AFM (XE NSOM) mapping was performed by using a customized tip from App Nano.

2.6. Functionalization of the HfO₂. 3-Aminopropyltriethoxysilane (99%) (Sigma Aldrich), glutaraldehyde solution (50 wt.% in H₂O, Bio Basic), *E. coli* antibodies (Rabbit polyclonal to *E. coli*, Abcam) was considered for functionalization of the HfO₂. The suspended device was absorbed into APTES for 2 Hrs in the mixture of ethanol/water followed by the cleaning of the chip by ethanol and blow-dried. Then, APTES functionalized chip was immersed in a glutaraldehyde solution for approximately 1 Hr. Afterward, *E. coli* antibodies were incubated to the suspended chip in 0.05 M PBS solution for 2 Hrs. at room temperature.

2.7. Bacteria growth. Bacterial specimens used in the study were prepared from culture plates. An isolated singer colony was added to 5 ml of media in a 14 ml culture tube. The culture tube was placed on a shaker in a 37°C incubator and incubated overnight (18 hours). The overnight culture was centrifuged at room temperature for 5 minutes at 3500 rpm. The supernatant was removed, and the bacteria pellet was re-suspended in 5 ml of filtered sterilized tap water. The bacteria were centrifuged, and the washing process repeated once. After the final wash, filtered tap water was added to the bacteria pellet until the optical density (measured at a wavelength of 600nm using a Spectro Max PLUS spectrophotometer) of the solution reached the desired value. After the optical density at 600nm was measured for the stock solution, serial dilutions were employed.

3.Results and Discussions:

3.1. Suspended Biosensor Fabrications

Our approach for patterning the nanogap electrode uses a self-assembly photolithography technique, as shown in Fig. 1 (a). The metal lift-off method has been used for engraving the first electrode gold (Ti/Au) and chromium (Cr), the thickness of the Au kept less than the Cr because it acts as a sacrificial layer as represented in Fig. 1(a) step I. After achieving the first electrode, the wafer kept at ambient temperature for an overnight where Cr gets oxidized and forms a thin layer of Cr_xO_y . Due to the formation of Cr_xO_y , the overall volume of the Cr increases by a few nanometers and overhangs on the edges of the bottom electrode, as demonstrated in Fig. 1(a) step II. It is crucial to keep the wafer in a controlled environment to achieve uniform expansion of the Cr to avoid unevenness in the nanogap and connection between two electrodes (Fursina et al. 2008). In the next step, Au is deposited and patterned for the second electrode. However, enlargement of the Cr layer in a few nanometers protects the second electrode from meeting the first metal layer and creates the nanometer spacing, as shown in Fig. 1 (a) step III. At IV step, Cr was stripped out left behind two Au electrodes, which are separated by few nanometers. In this self-assembly technique, the lateral resolution of the gap between two gold electrodes achieved nearly 70 to 90 nm at room temperature. The nanogap spacing between two electrodes is reproducible, constant throughout the wafer, and the feasible for bulk fabrication due to the involvement of the wafer-scale lithography. To examine the fabricated electrode, which is not electrically connected, the voltage sweep has been performed, and the resulting current monitored. This electrical characteristic demonstrated that the open circuit or short circuit between two nanogap electrodes (Supplementary Fig. 1). Throughout the wafer scale fabrication process of the nanogaps, there are several factors involves for achieving high yield such as

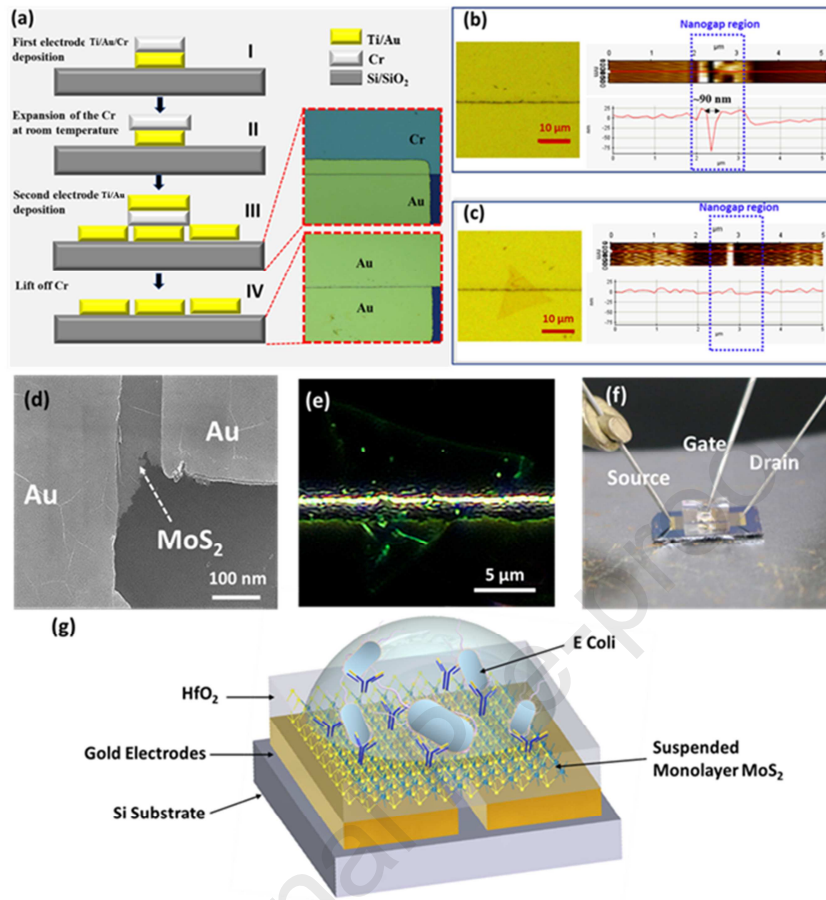


Fig. 1| Fabrication of suspended MoS₂ FET: (a) Schematic representation of the process flow to achieve nanogap electrodes. Optical image, AFM mapping, and line diagram of nanogaps (b) without MoS₂ (c) with MoS₂. (d) high magnification SEM image of MoS₂ on the top of nanogaps (e) Optical dark field image of MoS₂ triangle on the top nanogaps. (f) Image of biosensor device with Source, Drain, and Gate (Ag/AgCl electrode) connection (g) Schematic of Biosensor with *E. coli* and antibodies.

chromium thickness, thermal oxidation, and thickness of the second layer Ti/Au. In our optimized process flow, the yield of the constant spacing was 70% to 80% (± 4 nm), which are mostly found in the center of the Silicon wafer.

For channel material in FET, CVD grown MoS₂ crystal used in this work, which was characterized by using Raman spectroscopy and atomic force microscopy (AFM) (Supplementary Fig. 2). After achieving nanogap electrodes, a single atomic layer of MoS₂ flake

was transferred on the top of nano spaced electrode by using a novel dry stamping method (Supplementary Fig. 3). The PMMA acts a sacrificial layer to transfer 2D material onto the nano gap electrode. After the transfer is done the PMMA is removed by placing the device in acetone bath, this will dissolve the PMMA without affecting 2D material transferred onto the nano gaps. This dry stamping method allows transferring a single crystalline MoS₂ triangle precisely on the top of nanogaps by using a micromanipulator. Nano-spacing between the electrode (90 nm) does not allow the MoS₂ membrane (0.65 nm) to touch the supporting substrate, and it is suspended in between the two electrodes. To confirm the freestanding nature of MoS₂, AFM imaging was performed on the nanogap region with and without MoS₂ to elucidate the suspension of the atomic layer on the electrodes. Fig. 1 (b) shows the optical image and AFM mapping with the line profile of the nanogap area without MoS₂ flake. AFM line profile of nanogap electrodes demonstrates the step height of 75 nm in between the two Au electrodes, which confirms the two metal layers are separated by nanometer spacing. Fig. 1(c) represents the optical image and AFM mapping of transferred MoS₂. AFM line profile of transferred MoS₂ on the top of nanogap indicates the flat line and no step height, which verify that the atomic membrane is suspended and remarkably stiff as well as robust. This stiffness of the atomic layer confirms that it is not interacted with supporting substrates and eliminates the scattering phenomena of the interface between the 2D material and the holder substrate. This mechanical strength provided by nano spacing does not allow the monolayer sags between the electrodes, which permits depositing another layer, i.e., HfO₂ and biological receptor for selective sensing. To further validate the rigid suspension of the MoS₂ monolayer, a scanning electron microscope (SEM) and dark field image of the MoS₂ triangle on the top of the nanogaps were performed, as shown in Fig. 1 (d) &

(e). These images clearly illustrate that the MoS₂ triangle does not sag in between the nanogap spacing; it is suspended and flat on the top of two Au electrodes.

After confirming the rigidity of the MoS₂, it is desirable to eliminate the density of defects by covering the bare channel material with insulating material and then functionalizing linkers and antibodies on the top of the insulator (Kalantar-zadeh and Ou 2015a; Sarkar et al. 2014a). If biomolecule directly functionalized on the semiconductor layer without insulation, it might attach to the metal electrodes and change the work function as well as the contact resistance, which leads to the reliability issue and bring the electrical variation in the sensor output. Therefore, the dielectric layer is essential to passivate the metal electrodes and eliminate direct contact with electrolyte or biomolecules. Therefore, a 25 nm HfO₂ dielectric layer is deposited on the top of the suspended device using atomic layer deposition (ALD). Afterward, macro fluidic storage is fabricated using Polydimethylsiloxane (PDMS) for encompassing electrolyte droplets, which is placed precisely on the top MoS₂ triangle covered with HfO₂. A reference electrode (Ag/AgCl) is used to apply bias through the electrolyte for stabilizing the process and better control of the operation under the biosensor regime, as shown in the optical image of Fig. 1 (f) (Sarkar and Banerjee 2012). Fig. 1 (g) represents the schematic structure of the suspended MoS₂ FET biosensor by selective detection of the *E. coli* bacteria.

3.2. Electrical Characterization:

The electrostatic effect of MoS₂ FET investigated to compare the electrical performance of the supported and suspended devices. Fig. 2 represents the current-voltage (I-V) characteristics by applying different gate voltages (V_{BG}) of fabricated devices in a dry and wet environment. Fig. 2 (a) and (b) illustrates the I_D - V_{BG} curve of supported and free-standing MoS₂ at 100 mV bias voltage (V_{DS}), where supported channel length is 2 μ m and suspended is ~90 nm.

The suspended device represents excellent ON/OFF ratio 10^7 and threshold voltage (V_T) 3.9 V as compared to the supported one, which is equal to 10^6 and -9 V at room temperature. The improvement of the V_T , i.e., switching of the device at low voltage, is due to the elimination of the supporting substrate, which confines the electrons mean free path by scattering as well as from trapped charges on the surface (Masurkar et al. 2020). Fig. 2 (a') and (b') represents the I_D - V_{DS} characteristics at different V_{BG} (-10V to 30V). In both cases, the V_{DS} varies from the 0 mV to 100 mV, and the output current shows the linearity with the input voltage, which confirms that MoS₂ has ohmic contact with the Au/Ti contacts. Suspended FET displays better V_T and an ohmic contact with metal in back gate voltage characteristics. Still, due to the high power consumption, it is not feasible to use in the biosensor domain. Therefore, we have used a thin dielectric layer of the HfO₂ on the top of the channel and studied the ionic gating effect on the suspended MoS₂ channel, as shown in Fig. 2 (c) and (c'). Phosphate buffer solution as an electrolyte has been used for the ionic gating measurements by transporting a drop in macro fluidic storage. I_D - V_{LG} analysis of suspended ionic gating FET displayed impressive Subthreshold swing (SS) 70 mV/dec, which is near to ideal at room temperature as well as the mobility (13.5 cm² V⁻¹ s⁻¹) ON/OFF ratio (10^7) and V_T (-0.29 V). The mobility of all three types of the devices are calculated and shown in the supplementary Table S1 (Supplementary Fig S4). Low SS, threshold voltage, high mobility and elimination of substrate scattering make these types of the FET structure more sensitive as well as consistency with better gate controllability in a wet environment.

3.3. pH sensing

After achieving excellent performance in the ionic gating effect, the suspended device investigated by changing the pH of the electrolyte solution. The pH sensing strongly depends

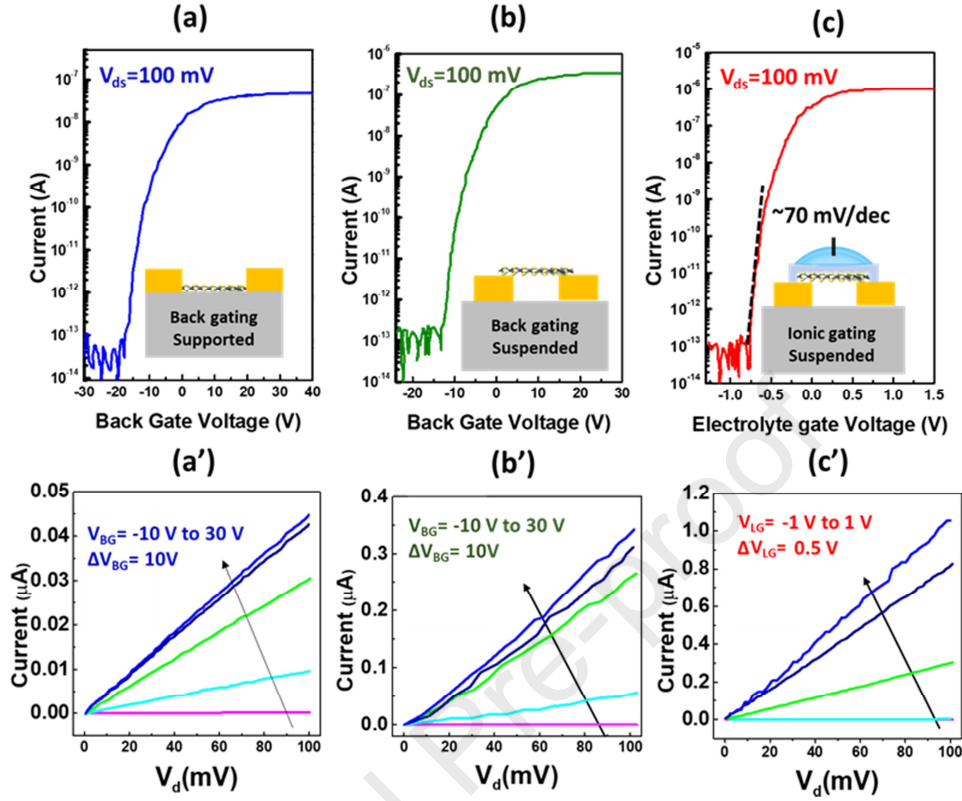


Fig. 2| Electrical characterization of suspended MoS₂ FET: Current-gate voltage curve at the constant bias 100 mV (a) back gating of supported MoS₂ (b) back gating of suspended MoS₂ with 90 nm channel length (c) Ionic gating of suspended MoS₂. Current-drain voltage characterization (a') Supported MoS₂ with back gating voltage from -10 V to 30 V (b') Suspended MoS₂ with back gating from -10 V to 30 V. (c') Suspended MoS₂ with ionic liquid gating from -1 V to 1V.

upon the proton (H^+) present in the electrolyte solution react with the OH group on the HfO₂ surface, which causes the protonation and deprotonation, as shown in Fig. 3 (a) (Shadman et al. 2016). Depending upon the pH value of the electrolyte solution, the dielectric surface charge of the suspended FET varies, a lower pH value protonates the surface by generating OH_2^+ on the HfO₂ whereas the higher pH value deprotonates the surface by extracting H^+ (Ameri et al. 2015; Dinar et al. 2019). Fig. 3 (b) represents the drain current (I_D) (100 mV bias voltage) at different pH values (3, 5, 7, and 9) by changing the electrolyte gate voltage (V_{LG}). It is found that the V_T of I_D - V_{LG} curves shifts to the positive side from low to high pH due to the negative charge

developed on the dielectric surface. The change in ΔV_T found to be 59.1 mV/pH, which is satisfied the Nernst limit at room temperature, i.e., 59.3 mV/pH and agree with early studies of ion-sensitive field-effect transistor (ISFET) characterization on HfO_2 (Van Hal et al. 1995; Zafar et al. 2011). Therefore, HfO_2 does not require any functionalization for pH sensing as compared to the SiO_2 , whose change in threshold voltage found to be 30-40 mV/pH (Bergveld and Physical 1996; Gao et al. 2009). Fig. 3 (c) represents the sensitivity of the pH at three different regions (Subthreshold, saturation, and linear), which is defined by the equation (1), where S_n is the sensitivity of pH, I_{pH1} and I_{pH2} are the transistor current measured from two different pH values (where $pH1 > pH2$).

$$S_n = \frac{I_{pH1} - I_{pH2}}{I_{pH2}} \times 100 \quad (1)$$

The sensitivity at the subthreshold region found to be much higher because the drain current is an exponential function of the gate voltage. In contrast, the saturation and linear parts are quadratic and linear with the change in gate potential. In a suspended device, the sensitivity at the subthreshold region was found to be ~876 from pH 5 to 6 and ~880 from pH 7 to 8 respectively, which is far better than the previous reports of MoS_2 FET biosensor on the supporting substrate (Sarkar et al. 2014b). The suspended MoS_2 FET possesses higher sensitivity than supported one because of the lower SS, where SS defined as the $SS = dV_{LG}/d \log_{10} I_D$. This equation elucidates that the change in subthreshold current by one decade is a function of the applied gate voltage, whereas the consistency in the sensitivity of the suspended device at two different ΔpH ranges is due to the elimination of the external scattering from the supporting substrate.

3.4. Bacteria sensing from suspended FET

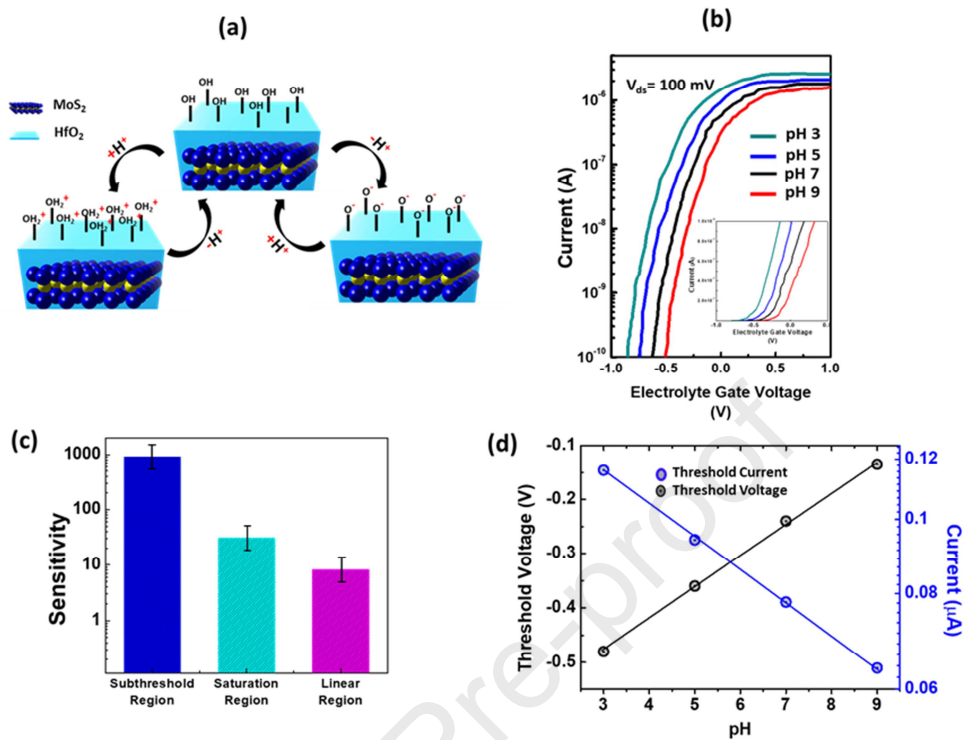


Fig. 3| pH detection of suspended MoS₂ FET: (a) Schematic of protonation and deprotonation on HfO₂ substrate. (b) I_D - V_{LG} curve of different pH concentration on the suspended MoS₂ (subset: Linear graph of I_D - V_{LG}). (c) Sensitivity of pH in subthreshold, saturation, and linear regions from pH 5 to pH 7. (d) experimental threshold voltage of suspended MoS₂ FET from pH 3 to pH 9 (Left-axis); experimental current corresponds to threshold voltage (Right-axis).

One more time, suspended FETs are fabricated as described in the fabrication section. To specifically detect the *E. coli* bacteria from the buffer solution (pH=7.4), linkers were immobilized on the top of the HfO₂. For evaluating the bonding of the linkers with oxide surface, di-thiol and (3-Aminopropyl)triethoxysilane (APTES) have used, followed by glutaraldehyde treatment and antibodies immobilization (Gunda et al. 2014; Lee et al. 2006). From these two molecules, APTES has shown promising I_D - V_{LG} curve shifts while detecting bacteria as compared to the di-thiol (Supplementary Fig. 5). It is interesting to see that after the functionalization of the APTES/glutaraldehyde linkers on the surface of the HfO₂, a small shift

of the I_D - V_{LG} curve illustrated (Supplementary Fig. 6). The overall process flow of the linkers binding, antibodies immobilization on suspended MoS_2 FET through macro storage fluidic channel to detect *E. coli* as shown in the schematic of Fig. 4 (a) (Gunda et al. 2014; Wu et al. 2013). The I_D - V_{LG} transfer curve of the linkers, antibodies, PBS buffer solution, and the 100 CFU/mL of *E. coli* bacteria was performed, as demonstrated in Fig. 4 (b). It is essential to introduce a buffer solution again after antibodies immobilization to confirm there is no change of conductance (Cyan curve in Fig. 4 (b)). A shift found in the I_D - V_{LG} curve on the left side after incubation of 100 CFU/mL of *E. coli* (Magenta curve in Fig. 4 (b)). This illustrated the current in the MoS_2 channel deteriorated due to the increment of hole concentration induced on the dielectric surface by highly negative charged bacteria wall (Huang et al. 2011).

After achieving considerable performance from the suspended MoS_2 FET device, different *E. coli* concentrations ranging from 0 to 10^3 CFU/mL were prepared and used for detection. The increase in the concentration of the bacteria leads to the decrement of the MoS_2 channel conductance. For buffer solution, I_D at ($V_{LG} = 0V$) is illustrated 0.3 μA , whereas 10 CFU/mL shows the current response 0.274 μA , the change in the conductance of freestanding MoS_2 found to be $\sim 9\%$ with a low concentration of the bacteria. The change in conductance for 100 CFU/mL and 10^3 CFU/mL with buffer solution illustrated 18% and 25%, respectively, as shown in Fig. 4 (c). Another essential factor in the biosensor domain is selectivity, which attributes the other bacterial strain should not be attached with *E. coli* antibodies to achieve false signals. Therefore, *Pseudomonas aeruginosa* (*P. aeruginosa*) bacteria (100 CFU/mL) were considered to measure the I_D - V_{LG} signal by keeping $V_{DS} = 100$ mV. *P. aeruginosa* bacteria did not show any change in conductance as compared to the *E. coli* (100 CFU/mL), as shown in Fig. 4(d). Fig. 4 (e) illustrates the sensitivity of the biosensor, which is defined as shown in equ (2),

$$S_{n(cfU/mL)} = \frac{I_{buffer} - I_{n(CFU/mL)}}{I_{n(CFU/mL)}} \times 100 \quad (2)$$

where $S_{n(CFU/mL)}$ sensitivity, Buffer is a buffer solution current, $I_{n(CFU/mL)}$ is current after bacteria bind to the FET biosensor. It was found that the shift in I_D-V_{LG} is more in the subthreshold region as compare to saturation and linear regions. The sensitivity of the sensor in the subthreshold

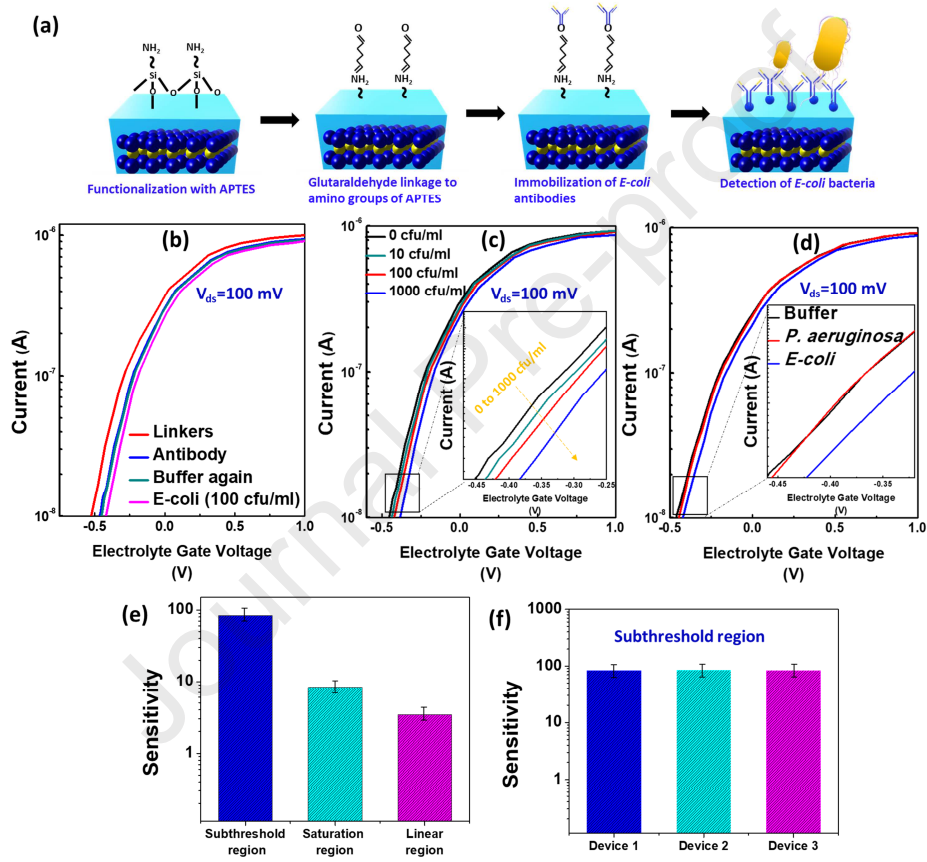


Fig. 4| *E. coli* bacteria detection on suspended MoS_2 FET: (a) Schematic diagram of self-assembly process for immobilization of *E. coli* bacteria on HfO_2 surface. (b) I_D-V_{LG} curve of SAM process at 100 mV bias; Functionalization of APTES (Red), immobilization of *E. coli* antibodies (Blue), buffer solution 0.01 M PBS solution of pH 7 (Cyan), and detection of *E. coli* bacteria of 100 CFU/mL (Magenta). (c) I_D-V_{LG} characterization of *E. coli* bacteria from 0 CFU/mL to 10³ CFU/mL. (d) Comparison of I_D-V_{LG} characterization of *E. coli* and *P. aeruginosa* bacteria (100 CFU/mL) with PBS buffer solution. (e) Sensitivity measurement of *E. coli* bacteria at subthreshold, linear, and saturated regions. (f) Sensitivity comparison of three suspended MoS_2 FET devices by immobilizing the 100 CFU/mL *E. coli* bacteria.

region achieved 83.3 for 100 CFU/mL of bacteria at -0.35 V. After obtaining higher sensitivity, the other most crucial factor in the 2D FET biosensor is the reliability in every fabricated sensor, which can be easily affected by external scattering from supporting substrate or the contact resistance from metal. In our case, we have maintained the constant work function between the electrode and the MoS₂ in every single device as well as eliminate the supporting substrate interaction, which attributes the constant sensitivity in three suspended MoS₂ FET at the subthreshold region (100 CFU/mL and -0.35 V) as demonstrated in Fig. 4 (f).

The sensitivity presented in this report is found to be impressive as compare to the previous reports of biomolecule detection via 2D FET based technology. Graphene-based FET biosensor has shown the conductance change of 3.25% at 10 CFU/mL of *E. coli*, which is almost three times smaller than the presented value (9% at 10 CFU/mL) (Chang et al. 2013; Huang et al. 2011). The detection limits of the graphene are due to the zero band gap in nature, which leads to an increase in the off-state leakage current and the SS value (SS is inversely proportional to the sensitivity) (Schwierz 2010). On the other hand, FET based MoS₂ biosensor has shown promising sensitivity in biomolecule detection due to ~1.8 eV bandgap. However, the uneven morphology from the supporting substrate leads to an unreliable output from every single device due to different interface resistances caused majorly by trapped charges, chemical adsorbates, and unknown functional groups (Jin et al. 2013; Kaushik et al. 2018). This is a significant barrier in proliferating this technology in biosensor industries. Therefore, elimination of the scattering effect in the proposed biosensor shows the overall change of conductance in the 2D film is entirely by the change in concentration of the biomolecule. Table S2 in Supporting Information represents a comparison of different 2D material based FET biosensor in terms of channel materials, detection limits, biomolecules, and sensitivity, whereas Table S3 in Supporting

Information illustrates the comparison between different dimension and materials with respective their sensitivity, feasibility in bulk fabrication, and reliability. Due to these merits i.e., the involvement of the CVD method, direct transfer technique, and optical lithography open the new avenues for the bulk fabrication of suspended MoS₂ biosensor.

4. Conclusion

In summary, we provide an approach for a comprehensive investigation of the highly sensitive and reliable biosensor based on suspended MoS₂ FET to detect the pH as well as *E. coli* bacteria. CVD grown MoS₂ channel material transferred on photolithographically patterned nano-gaps to achieve suspension and covered with Hafnium oxide (HfO₂) as a dielectric material to eliminate direct functionalization on the channel material. The proposed biosensor demonstrated the reliable and quantitative detection of pH and biomolecule by eliminating the scattering effect from the supporting substrate. A control experiment of pH sensing confirms the high sensitivity (~880/pH) as well as the constant I_D - V_{LG} curve shift at different pH range (3 to 9). Suspended MoS₂ FET biosensor provides selective detection of different concentration of the *E. coli* bacteria as low as 10 CFU/mL via changing the conductance by 9% of MoS₂ channel at ultralow power (V_{DS} =100 mV). This architecture can detect the low concentration of biomolecule because of the atomic layer as a channel material, scalable due to the involvement of optical photolithography, dry stamping, and CVD technique as well as reliable and reproducible due to the elimination of the supporting substrate. These merits of suspended MoS₂ FET biosensor makes it promising next-generation sensing device for on-spot detection of biomolecule such as COVID-19 viruses and bioelectronics applications.

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Acknowledgments

This article was supported in part by the National Science Foundation under grant no 1751472 and ACS Petroleum Research Fund (ACS PRF: 57647-DNI10)

Author contributions

NM, GW., and LMRA conceived and designed the experiments; NM and SV fabricated and characterized materials and devices, NM and SV electrical characterization; NM, SY, and NT performed the biological experiments; NM, SV, GW., and LMRA analyzed the data; NM, NT, SY wrote the paper.

Competing interests

The authors declare no competing financial interests.

Highlights:

1. A suspended MoS₂ atomic layer field-effect transistors is reported.
2. Self-assembled nanogap electrodes provide remarkable robust strength to the MoS₂ atomic layer.
3. Suspended MoS₂ field-effect transistor device exhibits impressive pH sensitivity and Escherichia coli bacteria detection.
4. The sensing mechanism highly selective to Escherichia coli bacteria.

Declaration of interests

☒ The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

☐ The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: