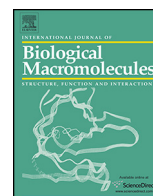




Contents lists available at ScienceDirect

International Journal of Biological Macromolecules

journal homepage: www.elsevier.com/locate/ijbiomac



Designing probe from E6 genome region of human *Papillomavirus* 16 for sensing applications

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ARTICLE INFO

Article history:

Received 12 July 2017

Received in revised form 9 October 2017

Accepted 9 October 2017

Available online xxx

Keywords:

Human *Papillomavirus*

Multiple sequence alignments

BLAST

Probe

Genome

Biosensor

ABSTRACT

Human Papillomavirus (HPV) is a standout amongst the most commonly reported over 100 types, among them genotypes 16, 18, 31 and 45 are the high-risk HPV. Herein, we designed the oligonucleotide probe for the detection of predominant HPV type 16 for the sensing applications. Conserved amino acid sequences within E6 region of the open reading frame in the HPV genome was used as the basis to design oligonucleotide probe to detect cervical cancer. Analyses of E6 amino acid sequences from the high-risk HPVs were done to check the percentage of similarity and consensus regions that cause different cancers, including cervical cancer. Basic local alignment search tools (BLAST) have given extra statistical parameters, for example, desire values (E-values) and score bits. The probe, 'GGG GTC GGT GGA CCG GTC GAT GTA' was designed with 66.7% GC content. This oligonucleotide probe is designed with the length of 24 mer, GC percent is between 40 and 70, and the melting point (T_m) is above 50 °C. The probe needed an acceptable length between 22 and 31 mer. The choice of region is identified here can be used as a probe, has implications for HPV detection techniques in biosensor especially for clinical determination of cervical cancer.

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1. Introduction

Over the last decade, amazing progress has been made to understand the pathogenesis of Human Papillomavirus (HPV). Examining the natural history and the study of disease transmission of HPV is complicated, because there are more than 100 strains of HPVs found [1–4]. Since all HPV sorts are immovably related, tests can be intended to target the conserved region of the genome or areas where sequences can best be used to isolate assorted HPV strains [5]. High-risk types are those like the sorts most of the time, were found in anogenital malignancies; by and large generally the low-risk types are those like the types related in condylomata [6–8]. There are several approaches to distinguish different types of HPV strains in clinical specimens. One of the basic ways is by utilizing primers coordinated to the generally conserved regions of HPV genome and after that, the consensus amplicon is recognized by the proper strategy, for example, dot blot hybridization with particular

probe(s), restriction fragment length polymorphism examination, or sequencing [9–15].

Generally, cervical cancer is created by the high-risk types of HPV [16–18] and the genome association of HPV strain 16, is one of the subtypes with a specific genome is known to cause cervical cancer predominantly (Fig. 1). This HPV genome includes an upstream regulatory region (URR), or long control region (LCR), has early genes (E1, E2, E6, and E7), and late genes (L1 and L2). It ought to be noted, about overlapping adjacent regions: e.g., E6, which is more prominent than E7, covers a huge segment of E7 [19–23]. Integration is essential, goes before cancer growth development, and could be incorporated into the malignant change [24]. In cervical carcinoma cell lines, the viral deoxyribonucleic acid (DNA) is typically incorporated into the human cell genome [25–27]. Previously, integration form had been found as much as 44%, episomal had 44% and both forms had 11% for cervical carcinomas containing HPV 16 [16,28,29]. A technique including real-time PCR has been utilized to screen for mixing status by determination of the E2/E6 proportion and might be appropriate for checking disease progression. Combination includes breakage of the HPV DNA and loss of parts in genome. Early perceptions of cervical carcinoma cell lines uncovered erasure of 2–3 kb that incorporated the E2 to L2 area [30–32]. The most crucial part is E6–E7 transcripts could be recognized in the cells. Open reading frame (ORFs) of E1, E2, E4, and E5

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<https://doi.org/10.1016/j.ijbiomac.2017.10.051>

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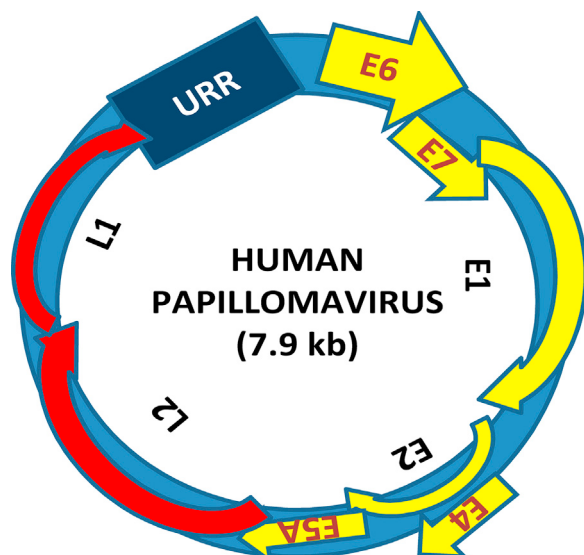


Fig. 1. Genome organization of human papillomavirus type 16. This subtype is known to cause cervical cancer. It has E1–E7 early genes, L1–L2 late genes; and capsid region.

were hindered by flanking the host cell DNA, indicating as sites of combination [33]. HPV mRNAs present hybridized probes for the whole E6 and E7 ORF and a minor part of the E1 ORF, implying that these were the main portions of the HPV genome present [34,35]. Investigation of six cervical carcinoma tests demonstrated that the E6/E7 region was observed to be held on joining, yet the E2 was lost. Viral genome integrated into human chromosomal DNA of cancer cells generally erases E2 ORF, which results in the loss of expression of the E2 gene, however, is connected with the continuation of high expression of E6 and E7 [36–42]. Whatever the rest of the genome may have a part in transmission of the infection to another host cell, yet it is the proteins encoded by E6 and E7 that cause the malignancy. Joining of HPV DNA happens early in cancer development and is vital as an enactment mechanism for movement from the precancer to cancer [43–46]. E6 and E7 regions demonstrate the best contrasts in nucleotide sequence between various HPV types and is constantly held by cells that are contaminated compared to other regions in HPV 16 [47–49].

DNA has a role as biomarker of HPV. Investigation of potential biomarker helps to identify a new pathway involved in the HPV pathogenesis to prevent cervical cancer. DNA probes are stretches of single-stranded used to detect the presence of complementary nucleic acid sequences; in other word, is a target sequence by hybridization process. HPV type-specific probes provide a rapid and cost-effective method to simultaneously detect HPV genotypes. Suitable DNA probe synthesize based on E6 area of the different types of HPV able to answer whether a patient is infected with a high-risk HPV, can be integrated and promptly fused into the sensing purposes [50,51]. The use of E6 DNA probes make the detection is considered to be applicable for early discovery of cervical cancer and expenses will be lower. The general principle of DNA biosensor involves the immobilization of a DNA or DNA-analog probe onto the electrode surface. The importance of DNA probe in the sensing application with biosensor has been shown to give a higher sensitivity and specificity, fast hybridization kinetic strength rather than ionic strength and requires only a shorter probe length [52]. Fig. 2 showed a research overview in the development of highly sensitive and selective nanoparticles based biosensor for early detection of cervical cancer.

To analyse the region of interest, BLAST programs are widely used as tools for looking protein and DNA databases for group-

ing similarities. Table 1 showed Basic Local Alignment Search Tool (BLAST) results of Oligonucleotide A for the identification of HPV 16 target. BLAST have been figured to differentiate protein or DNA queries with databases, with DNA sequences frequently experiencing reasonable interpretation before any experimental procedures is performed. The aim of this research was to analyze the importance and usage of probe resides in E6 genome region of HPV 16 for sensing purposes through bioinformatics analysis such as BLAST and CLUSTALW. This type of probe identification can be used as a biomarker for biosensing receptor element in biosensor application for early detection of cervical cancer (Fig. 3). Fig. 3 is overall figure revealing the sensing application for the detection of HPV. DNA biomarker have been selected from the gene profiling and involved 4 strains of HPV that cause cervical cancer, HPV 16 (AAA46939.1), HPV 18 (P06463.1), HPV 31 (AGM34418.1), and HPV 45 (AGM34429.1). The biosensor device requires the efficient immobilization of nucleic acids over the surface of a transducer, which is responsible for the analyte recognition and executed a response signal proportional that can be measured the analyte concentration. The challenge for biosensor device relies on the importance of preserving the biological activity of probe as well as the transducing activities for measurable electrical biosensor.

In this research work, designing a probe from E6 genome region of HPV 16 for sensing applications was developed using HPV DNA probe complementary to a 24 bases region unique to E6 region gene of HPV type 16. Gravitt et al. [53] conclude that a high viral load is associated with prevalent cervical cancer precursors for the most HR-HPV genotypes, but only HPV 16 load predicts the development of cervical cancer disease.

2. Materials and methods

2.1. Analysis of HPV sequence from high-risk HPVs

Different strains of HPV sequences were searched in GenBank by using FASTA. Nucleotide sequences of HPV in GenBank were identified. A sequence was viewed as a match if it had more than 70% nucleotide comparability [54]. The reliability of this method for generating other probes, rather than HPV with no less than 80% comparability to the relating HPV sequences in GenBank were thought to be available. If a sequence search produced 95% similitude to HPV type 16 (HPV 16) and 85% closeness to HPV 18, the sample was viewed as positive for both types. HPV16 oligonucleotide probe sequence can be utilized to cross-examine a grouping database utilizing a homology search by utilizing BLAST programming. Primer-BLAST programming [55] allow quick homology finding of a sequence within a constantly updated sequence database.

2.2. Identification of HPV 16 target DNA

HPV16 oligonucleotide probe sequence can be utilized to investigate a sequence database utilizing a homology search by BLAST programming. Broad databases are accessible on the web and can be openly obtained (<http://www.ncbi.nlm.nih.gov>). Primer-BLAST software [55] allows quick homology finding of a sequence inside a consistently redesigned sequence database. At the point when the query is submitted, either as a sequence in FASTA group or as an arrangement identifier, e.g. GenBank accession form, the inquiry is sent to the BLAST server and a 'Request Identifier' (RID) is returned. The inquiry and results are put away in an organized arrangement for up to 24 h after a RID is issued. The RID distinguishes the query and permits the outcome to be seen in a few organizations, which incorporate the natural BLAST report, an improved 'hittable', XML and ASN.1. [56]. Single-stranded carboxylate 24 mer synthetic

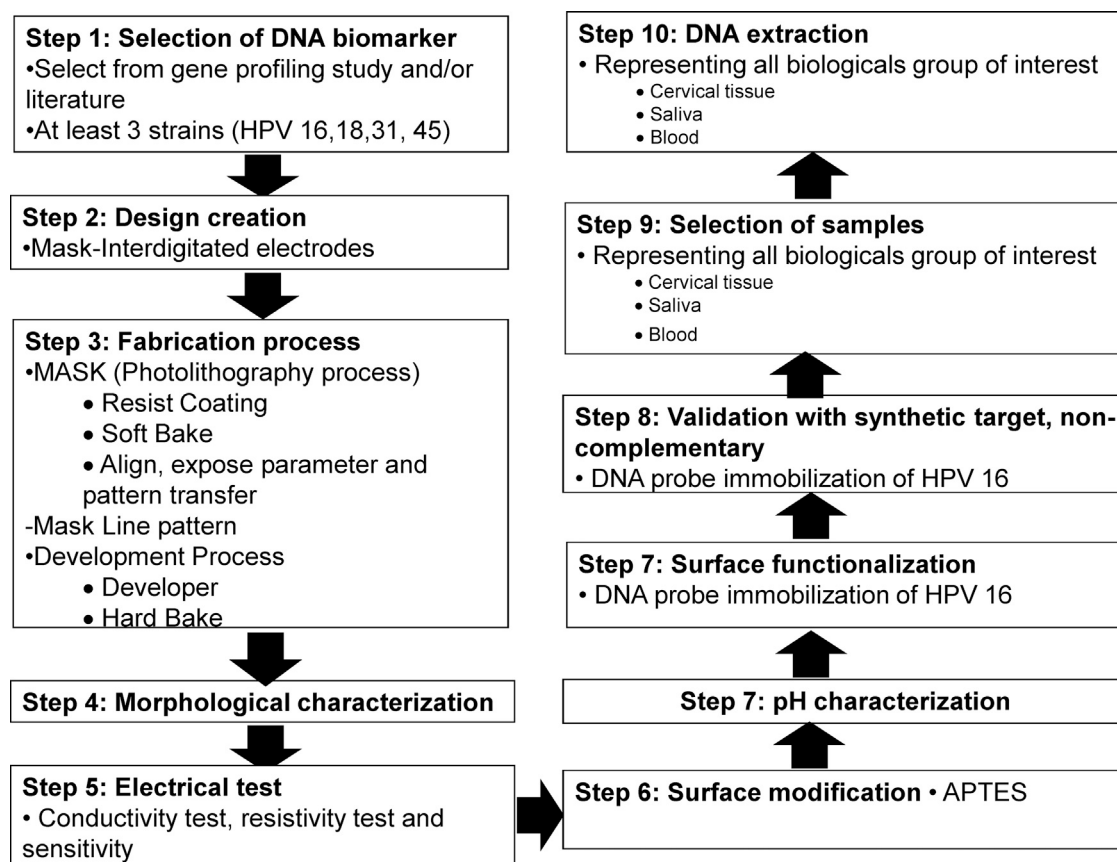


Fig. 2. Research overview in development of highly sensitive and selective nanoparticles based biosensor for early detection of cervical cancer.

Table 1

Basic Local Alignment Search Tool (BLAST) results of Oligonucleotide A for the identification of HPV 16 target DNA.

Accession	Description	Max Score	Total score	Query coverage	E value	Max identity
KF700140.1	Human papillomavirus type 16 isolate 85 E6 protein (E6) gene, complete CDS	48.1	48.1	100%	0.001	100%
KF700139.1	Human papillomavirus type 16 isolate 67 E6 protein (E6) gene, complete CDS	48.1	48.1	100%	0.001	100%
KF700134.1	Human papillomavirus type 16 isolate 28 E6 protein (E6) gene, complete CDS	48.1	48.1	100%	0.001	100%
KM030565.1	Alphapapillomavirus 9 isolate Afr4 E6 oncoprotein (E6) gene, complete CDS	48.1	48.1	100%	0.001	100%
KM030564.1	Alphapapillomavirus 9 isolate Eur18 E6 oncoprotein (E6) gene, complete CDS	48.1	48.1	100%	0.001	100%
KM030563.1	Alphapapillomavirus 9 isolate Eur14 E6 oncoprotein (E6) gene, complete CDS	48.1	48.1	100%	0.001	100%
KJ869425.1	Human papillomavirus type 16 isolate HPV BHU 3 E6 protein (E6) and E7 protein (E7) genes, partial CDS	48.1	48.1	100%	0.001	100%
KJ922765.1	Human papillomavirus type 16 strain 7 isolate salem-2 transforming protein (E6) gene, complete CDS	48.1	48.1	100%	0.001	100%
KJ922764.1	Human papillomavirus type 16 strain 7 isolate salem-1 transforming protein (E6) gene, complete CDS	48.1	48.1	100%	0.001	100%

oligonucleotide probe designed from HPV 16 for recognizable probe of cervical cancer (Accession No.: A18875.1).

3. Results and discussion

3.1. Computational biology tools for analysing HPV sequences

Computational science apparatuses have opened new bits of knowledge concerning the portrayal and protection of biological molecule sequences among the particular viral genomes [57]. By utilizing different sequence alignments (CLUSTALW), the target of present study is to perform the relative analysis of E6 protein sections among the high-risk HPV genotypes that cause cervical cancer. For these analyses, 4 types of E6 protein segments present in HPV high-risk genotypes that cause cervical cancer, namely; 16, 18, 31 and 45 were considered. At first, amino acid sequences of E6 protein from these fourth genotypes of high-risk HPV were

searched utilizing GenBank database (<http://www.ncbi.nlm.nih.gov/protein>). Recognizable proof of amino acid sequences comparing to E6 protein was saved in Biology Workbench database (<http://workbench.sdsc.edu>) with accession numbers: AIQ82781.1, P06463.1, AGM34418.1 and AGM34429.1 for HPV16, 18, 31 and HPV 45, individually. The numerous alignment sequences of the HPV high-risk amino acid residues in the E6 viral protein among unmistakable HPV genotypes were created utilizing the CLUSTALW database apparatuses (Fig. 4), giving significant factual parameters, for example, similarity and identity. Multiple sequence alignments of the nucleotide residues in the E6 in a complete genome of HPV 16 by using CLUSTALW were displayed in Fig. 5. Columns showing indistinguishable nucleotide similarly situated in numerous types 'i' were analyzed exclusively highlighted by an asterisk (*) mark which showed fully conserved residues. Symbol (–) showed no consensus of the groups. Specific HPV E6 region as oligonucleotide probe for the detection of HPV 16 is indicated in red, which prove

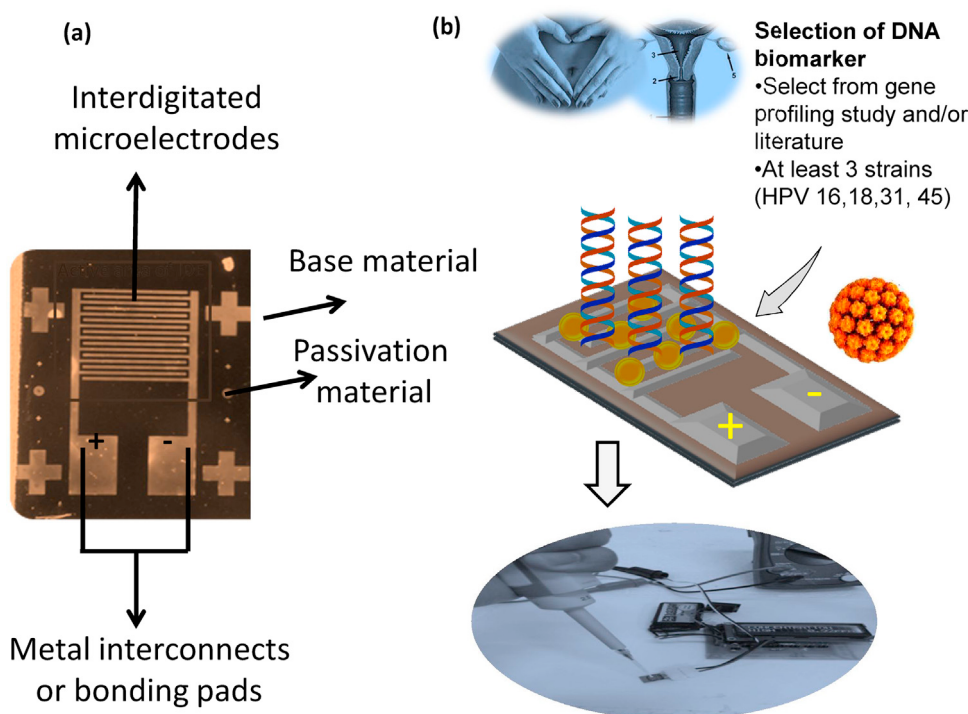


Fig. 3. Overall figure for sensing application for the detection of HPV. DNA biomarker have been select from gene profiling and involved 4 strains of HPV that cause cervical cancer, HPV 16 (AAA46939.1), HPV 18(P06463.1), HPV 31(AGM34418.1), and HPV 45 (AGM34429.1).

AGM34429.1_HPV_45_E6	-----MARFDDPKQRPYKLPDLCTELNTSLQDVSACVYCKATLERTEVY
P06463.1_HPV18_E6	-----MARFEDPTRRPYKLPDLCTELNTSLQDIEITCVYCKTVLELTVF
AAA46939.1_HPV_16_E6	MHQKRTAMFQDPQERPRKLPQLCTELQTTIHDIILECVYCKQQLRREVY
AGM34418.1_HPV_31_E6	-----MFKNPAERPRKLHELSSALEIPYDELRLNCVYCKGQLTETEV
	.: .** ** :*.: *: . :.: : ***** *
AGM34429.1_HPV_45_E6	QFAFKDLCIVYRDCIAYAACHKCIDFYSRIRELRYYSNVYGETLEKITN
P06463.1_HPV18_E6	EFAFKDLFVYRDSIPHAACHKCIDFYSRIRELRYSDSVYGDITLEKITN
AAA46939.1_HPV_16_E6	DFAFRDLCIVYRDGNPYAVCDKCLKFYISKISEYRHYCYSLYGTTLEQQYN
AGM34418.1_HPV_31_E6	DFAFTDLTIVYRDTPIYGVCTKCLRFYSKVSFRWYRYSVYGTITLEKITN
	:*** ** :**** .:..* **: ***: * * * *:*** **:
AGM34429.1_HPV_45_E6	TELYNLLIRCLRCQKPLNPAEKRRHLKDKRRFHNIAGQYRGQCNTCCDQA
P06463.1_HPV18_E6	TGLYNLLIRCLRCQKPLNPAEKLRLHNEKRRFHNIAGHYRGQCHSCCNRA
AAA46939.1_HPV_16_E6	KPLCDLLIRCLNCQKPLCPEEKQRHLDDKQRFHNIIGRWTGRCMSSCRSS
AGM34418.1_HPV_31_E6	KGICDLLIRCLTCQRPCLPEEKQRHLDDKKRFHNIIGRWTGRCIVCWR--
	. : :*****: **:* * * * ***. :***** *: : * : *
AGM34429.1_HPV_45_E6	RQERLRRRRRETQV
P06463.1_HPV18_E6	RQERLQRRRETQV
AAA46939.1_HPV_16_E6	R-----TRRETQL
AGM34418.1_HPV_31_E6	R-----PRTETQV
	* * * * *

Fig. 4. CLUSTAL W (1.81) multiple sequence alignments of the amino acid residues in the E6 protein sequences among high-risk HPV types cause cervical cancer. Four high-risk genotypes was identified; 16, 18, 31, 45. Columns indicate identical amino acids in the same position in all types analyzed are exclusively indicated by an asterisk (*) which showed fully conserved residues, whereas alignments showing the presence of similar amino acids are represented by other symbols (: for conservation of strong groups or .) for conservation of weak groups. Symbol (–) showed no consensus of the groups.

that oligonucleotide probe for the detection of HPV16 was present in the sequences. Nucleotide and amino acid sequences encoded HPV 16 isolate IR-32 E6 (E6) genes; complete coding sequence

(CDS) is shown in Fig. 6. Specific HPV E6 region as oligonucleotide probe for the detection of HPV 16 is indicated by underline. RGRWT-GRC was identified as a biomarker region for E6 region of HPV 16.

```
K02718.1_HPV_16_COMPLETE_GEN      ACTACAATAATTCATGTATAAACTAAGGGCGTAACCGAAATCGGTTGAA
KM058604.1_HPV16_E6                -----
EU869318.1_HPV_16_E6_and_E7        -----CGTAACCGAAATCGGTTGAA

K02718.1_HPV_16_COMPLETE_GEN      CCGAAACCGGTTAGTATAAAAGCAGACATTTTATGCACCAAAAGAGAACT
KM058604.1_HPV16_E6                -----ATGCACCAAAAGAGAACT
EU869318.1_HPV_16_E6_and_E7        CCGAAACCGGTTAGTATAAAAGCAGACATTTTATGCACCAAAAGAGAACT
                                     *****

K02718.1_HPV_16_COMPLETE_GEN      GCAATGTTTCAGGACCCACAGGAGCGACCCAGAAAGTTACCACAGTTATG
KM058604.1_HPV16_E6                GCAATGTTTCAGGACCCACAGGAGCGACCCAGAAAGTTACCACAGTTATG
EU869318.1_HPV_16_E6_and_E7        GCAATGTTTCAGGACCCACAGGAGCGACCCAGAAAGTTACCACAGTTATG
                                     *****

K02718.1_HPV_16_COMPLETE_GEN      CACAGAGCTGCAAAACAACATACATGATATAATATTAGAATGTGTGTACT
KM058604.1_HPV16_E6                CACAGAGCTGCAAAACAACATACATGATATAATATTAGAATGTGTGTACT
EU869318.1_HPV_16_E6_and_E7        CACAGAGCTGCAAAACAACATACATGATATAATATTAGAATGTGTGTACT
                                     *****

K02718.1_HPV_16_COMPLETE_GEN      GCAAGCAACAGTTACTGCGACGTGAGGTATATGACTTTGCTTTTCGGGAT
KM058604.1_HPV16_E6                GCAAGCAACAGTTACTGCGACGTGAGGTATATGACTTTGCTTTTCGGGAT
EU869318.1_HPV_16_E6_and_E7        GCAAGCAACAGTTACTGCGACGTGAGGTATATGACTTTGCTTTTCGGGAT
                                     *****

K02718.1_HPV_16_COMPLETE_GEN      TTATGCATAGTATATAGAGATGGGAATCCATATGCTGTATGTGATAAATG
KM058604.1_HPV16_E6                TTATGCATAGTATATAGAGATGGGAATCCATATGCTGTATGTGATAAATG
EU869318.1_HPV_16_E6_and_E7        TTATGCATAGTATATAGAGATGGGAATCCATATGCTGTATGTGATAAATG
                                     *****

K02718.1_HPV_16_COMPLETE_GEN      TTTAAAGTTTATCTCAAATTAGTGAGTATAGACATTATTGTTATAGTT
KM058604.1_HPV16_E6                TTTAAAGTTTATCTCAAATTAGTGAGTATAGACATTATTGTTATAGTT
EU869318.1_HPV_16_E6_and_E7        TTTAAAGTTTATCTCAAATTAGTGAGTATAGACATTATTGTTATAGTT
                                     *****

K02718.1_HPV_16_COMPLETE_GEN      TGTATGGAACAACATTAGAACAGCAATACAAACCGTTGTGTGATTG
KM058604.1_HPV16_E6                TGTATGGAACAACATTAGAACAGCAATACAAACCGTTGTGTGATTG
EU869318.1_HPV_16_E6_and_E7        TGTATGGAACAACATTAGAACAGCAATACAAACCGTTGTGTGATTG
                                     *****

K02718.1_HPV_16_COMPLETE_GEN      TTAATTAGGTGTATTAACTGTCAAAGCCACTGTGTCCTGAAGAAAAGCA
KM058604.1_HPV16_E6                TTAATTAGGTGTATTAACTGTCAAAGCCACTGTGTCCTGAAGAAAAGCA
EU869318.1_HPV_16_E6_and_E7        TTAATTAGGTGTATTAACTGTCAAAGCCACTGTGTCCTGAAGAAAAGCA
                                     *****

K02718.1_HPV_16_COMPLETE_GEN      AAGACATCTGGACAAAAGCAAGATTCCATAATATAAGGGTCGGTGGA
KM058604.1_HPV16_E6                AAGACATCTGGACAAAAGCAAGATTCCATAATATAAGGGTCGGTGGA
EU869318.1_HPV_16_E6_and_E7        AAGACATCTGGACAAAAGCAAGATTCCATAACATGAGGGTCGGTGGA
                                     *****

K02718.1_HPV_16_COMPLETE_GEN      CCGGTCGATGTATGTCTTGTGTCAGATCATCAAGAACCGTAGAGAAACC
KM058604.1_HPV16_E6                CCGGTCGATGTATGTCTTGTGTCAGATCATCAAGAACCGTAGAGAAACC
EU869318.1_HPV_16_E6_and_E7        CCGGTCGATGTATGTCTTGTGTCAGATCATCAAGAACCGTAGAGAAACC
                                     *****

K02718.1_HPV_16_COMPLETE_GEN      CAGCTGTAATCATGCATGGAGATACACCTACATTGCATGAATATATGTTA
KM058604.1_HPV16_E6                CAGCTGTAATCATGCATGGAGATACACCTACATTGCATGAATATATGTTA
EU869318.1_HPV_16_E6_and_E7        CAGCTGTAATCATGCATGGAGATACACCTACATTGCATGAATATATGTTA
                                     *****

K02718.1_HPV_16_COMPLETE_GEN      GATTTGCAACACAGAGACAACATGATCTCTACTGTTATGAGCAATTAATGA
KM058604.1_HPV16_E6                -----
EU869318.1_HPV_16_E6_and_E7        GATTTGCAACACAGAGACAACATGATCTCTACTGTTATGAGCAATTAATGA
```

GGG GTC GGT
GGA CCG GTC
GAT GTA

Fig. 5. CLUSTAL W (1.81) multiple sequence alignments of the nucleotide residues in the E6 and E7 in a complete genome of HPV 16. Columns indicating identical nucleotide in the same position in all types analyzed are exclusively indicated by an asterisk (*) which showed fully conserved residues. Symbol (–) showed no consensus of the groups. Specific HPV E6 region as oligonucleotide probe for detection of HPV 16 are indicated by red. Comparative analysis of the E6 viral protein from HPV strains 16, 18, 31 and 45. In order of appearance of E6 region for different HPV strains; HPV 16 (AAA46939.1), HPV 18 (P06463.1), HPV 31 (AGM34418.1), and HPV 45 (AGM34429.1).

HPV16 was the most frequent genotype in cervical and oropharyngeal cancer [58].

3.2. Comparative analysis of HPV E6 viral region cause cervical cancer

The relative examination of the E6 viral protein demonstrated an 81% comparability between HPV 18 and 45 types, 65% between HPV 16 and 31, 54% between HPV 16 and 45, 53% between HPV 16 and 18, 51% between HPV 18 and 31, 49% between HPV 31 and 45 (Fig. 7). The amino acids alignment between HPV 18 and 45 genotypes revealed the similarity value corresponding 81% and showed the highest similarity among pairwise alignments, even though both strain are too far from each other, followed by 65% between HPV 16 and 31. The relative examination of the E6 viral protein demonstrated 53% comparability between the regular HPV types 16 and 18. The most reduced comparability is between HPV 31 and 45.

3.3. Basic local alignment search tools (BLAST) for HPV 16

Further examination by utilizing basic local alignment search tools (BLAST) gave extra statistical parameters, for example, expectation values (e-values) and score bits [59]. E-values in blast results represent to the probability of the alignment has happened by chance, while the estimation of score bits relies on upon size of

alignment, the number of matches/mismatches/gap and matrix utilized for the correlation of sequences and is normalized by the mean for variable statistics [17]. Blast analysis of oligonucleotide A for recognizable proof of HPV 16 target DNA produced huge hits with E-values comparing to 0.001% and the greatest identity of 100% and 48.1 score bits based on HPV 16 isolate 85, E6 protein (E6) gene (Fig. 8). According to Brzeski [60], E-values near zero shows that the match of E6 proteins, is noteworthy in short sequences and subsequently, the discoveries introduced in this study exhibit complemented likenesses in the alignments of the amino acid residues deposits in the E6 protein among high-risk HPV. While Basic Local Alignment Search Tool (BLAST) outflanks accurate techniques through its utilization of heuristics, the pace of the BLAST programming is imperfect for long inquiries or database arrangements. There are a few weaknesses in the User Interface (UI) of the present charge line applications. Notwithstanding performing alignments, BLAST gives an expect value, statistical data about the significance of every arrangement of HPV E6 region.

3.4. Efficiency of E6 region of oligonucleotide probe

Particular probe that coordinated at the E6 region reported to have an especially more prominent probability annealing than consensus probe focusing on the L1 region [61]. Degenerating probe used to distinguish L1 sequences are not 100% dependable, owing to the more sequences succession varieties in this region of the HPV

```

atgcaccaaagagaactgcaatgtttcaggacccacaggagcgacccagaaagttacca
M H Q K R T A M F Q D P Q E R P R K L P
cagttatgcacagagctgcaaacaactatacatgatataatattagaatgtgtgtactgc
Q L C T E L Q T T I H D I I L E C V Y C
aagcaacagttactgcgacgtgaggtatatgactttgcttttcgggatttatgcatagta
K Q Q L L R R E V Y D F A F R D L C I V
tatagagatgggaatccatattgctgtatgtgataaatgtttaagttttattctaaaatt
Y R D G N P Y A V C D K C L K F Y S K I
agtgagtatagacattattgttatagtttgatggaacaacattagaacagcaataacaac
S E Y R H Y C Y S L Y G T T L E Q Q Y N
aaaccgttgtgtgatttgttaattaggtgtattaactgtcaaaagccactgtgtcctgaa
K P L C D L L I R C I N C Q K P L C P E
Gaaaagcaaagacatctggacaaaaagcaaagattccataatataaggggtcgggtggacc
E K Q R H L D K K Q R F H N I R G R W T
ggtcgatgtatgtcttgttgcagatcatcaagaacacgtagagaaaccagctgtaa
G R C M S C C R S S R T R R E T Q L -

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Fig. 6. Nucleotide and amino acid sequences encoded HPV type 16 isolate IR-32 E6 (E6) genes, complete coding sequence (CDS). Specific HPV E6 region as oligonucleotide probe for detection of HPV 16 are indicated by underline.

```

Sequence type explicitly set to Protein
Sequence format is Pearson
Sequence 1: AAA46939.1_HPV_16_E6          158 aa
Sequence 2: AGM34429.1_HPV_45_E6          158 aa
Sequence 3: AGM34418.1_HPV_31_E6          149 aa
Sequence 4: P06463.1_HPV18_E6             158 aa
Start of Pairwise alignments
Aligning...
Sequences (1:2) Aligned. Score: 54
Sequences (1:3) Aligned. Score: 65
Sequences (1:4) Aligned. Score: 53
Sequences (2:3) Aligned. Score: 49
Sequences (2:4) Aligned. Score: 81
Sequences (3:4) Aligned. Score: 1
Time for pairwise alignment: 0.038811

```

Fig. 7. Comparative analysis of the E6 viral protein from HPV strains 16, 18, 31 and 45. In order of appearance of E6 region for different HPV strains, for HPV 16 (AAA46939.1), HPV 18 (P06463.1), HPV 31 (AGM34418.1), and HPV 45 (AGM34429.1).

genome. Contrasted with the E6 region, L1 probe may have higher chances for disappointment in hybridizing and may prompt mispriming. Sequence closeness seeking is an essential undertaking in bioinformatics apparatuses [62]. Comparability searches are a key part and they form the bases of basic structural motif identification, gene identification, and bits of knowledge into useful affiliations. With the fast increment in the accessible genetic data information through a wide assortment of databases, closeness searches are a crucial apparatus down getting to this information in an instructive and beneficial way.

A few studies have demonstrated a noteworthy part of the E6 protein in the induction and maintenance of cell change, acting

basically as an oncoprotein by fortifying the decimation of numerous host cell key administrative proteins, coming full circle in the cervical cancer side effects [63]. Steady proof proposes that the genome of papillomaviruses is static, and changes in its sequence by mutation or recombination are exceptionally uncommon occasions [64]. Diverse virus species within the same genus offer around 60% to 70% similarity [65]. Thus, the discoveries introduced in this study, exhibiting the presence of a highly conserved region with critical comparability of the amino acid sequences in the E6 protein among distinct HPV regions that cause cervical cancer uncovered more grounded proof in regards to the molecular characterization and developmental protection of viral genomes utilizing compar-

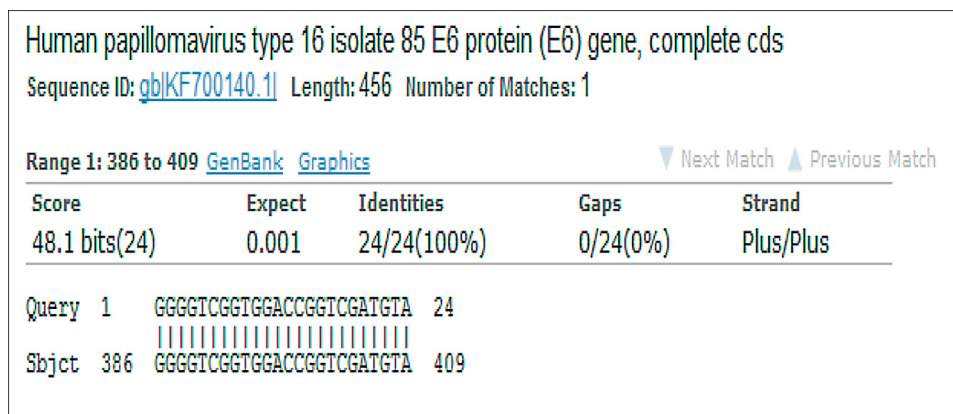


Fig. 8. Alignment statistics for match of Oligonucleotide A for the identification of HPV 16 target DNA and Human *Papillomavirus* (HPV) type 16 isolate 85 E6 proteins (E6) gene.

active investigation of various sequence alignments. The probe was designed with the sequence “GGG GTC GGT GGA CCG GTC GAT GTA”, having 66.7% GC content. The oligonucleotide probe can be used as a biomarker for detection E6 genome region of HPV 16 due to the characteristics; length between 22 and 30 mers, % GC between 40 and 70, and melting point (T_m) is above 50 °C. The probe needed an acceptable length between 22 and 31 mers. Since the higher degeneracy lowers the primer concentration, and affecting the T_m of probe. Besides that, oligonucleotides probe must have a GC content of 40–70% which is known to facilitate stronger binding to target annealing [66].

3.5. Future work of interdigitated electrodes sensor efficiency

High quality of cytology-based method requires a trained personnel and specialized equipment. HPV vaccines may provide the required potential approach for the prevention of cervical cancer. HPV testing mostly required a high technology laboratory-based molecular analysis involved molecular biologist and costly. Currently, several aims of the development of rapid, simple, accurate and affordable HPV tests. Due to rapid results, women can be screened and treated during the same visit to the hospital for examination by the doctor.

Innovative HPV-based biosensor enables the establishment and effective screening detection. The interdigitated electrodes (IDEs) discussed in this study consisted of a two-electrode system prepares as previously described [50,51,67]. We reported the gold nanoparticle mediated method for the spatially resolved deposition of DNA on nano-gapped interdigitated electrodes [50], and its application to the detection of the human Papillomavirus. We also reported a direct detection of human papillomavirus 16 genomic DNA using gold nanoprobe [51]. We summarized the main steps involved in fabrication (wafer cleaning, oxide layer growth, negative photoresist spin coating, photoresist patterning, aluminium (Al) deposition, photoresist removing and wafer dicing) [67]. Briefly, a 4 inch p-type SOI wafer was used as a substrate and cleaned using buffered oxide etchant (BOE) to remove a native oxide layer on SOI surface. After the cleaning process, the SOI wafer was cut into small pieces (2 cm × 2 cm), and followed by EBL process. The IDE nanosensor was fabricated on a wafer with a 145 nm buried oxide layer. Silicon wafer was used as a main substrate in order to form silicon dioxide (SiO_2) as an insulation layer of an electrical device. The wafer was cleaned using BOE to remove the native oxide, which had been naturally grown on it. Growth of SiO_2 using wet oxidation process provides a thicker insulation layer and shorter time consuming compared to dry oxidation process. In order to transfer pattern from a mask on a wafer, photoresist was

spin-coated on the growth SiO_2 wafer using spin-coater. Photoresist is a light-sensitive material used to form a patterned coating on a surface. By using deep ultraviolet lithography, a 50 nm silicon layer was patterned and etched when UV-light was exposed on the photoresist; pattern from chrome mask was directly transferred onto the photoresist. After development process, aluminum metal was deposited using sputter-coater and acetone was used to strip the unwanted photoresist. The designed study here can also be implemented in other potential studies have been performed in past for further expansion [68–70].

4. Conclusion

In this research study, designation of oligonucleotides probe designed for the detection of Human Papillomavirus (HPV) 16 towards nanobiosensing. E6 region of HPV16 was considered and revealed the possible specific probe by comparison analyses with other potent subtypes 18, 31 and 45. The probe designed has all appealing characteristics, can be used as a biomarker for detection E6 genome region of HPV 16. By availing the oligonucleotide regions predicted for probe designing, the profoundly sensitive HPV identification by nanobiosensor will significantly overestimate the extent of cervical malignancy patients who have low-quality cytological variations from the norm.

Conflict of interests

None declared.

Acknowledgements

This research was supported by the Ministry of Higher Education (MOHE) under Grant Nos. 9003-00481 and the author would like to thank all staff members of the Institute of Nanoelectronic Engineering in Universiti Malaysia Perlis for their technical advice.

References

- [1] B. Serrano, S. De Sanjosé, S. Tous, B. Quiros, N. Muñoz, X. Bosch, L. Alemany, Human papillomavirus genotype attribution for HPVs 6, 11, 16, 18, 31, 33, 45, 52 and 58 in female anogenital lesions, *Eur. J. Cancer* 51 (2015) 1732–1741, <http://dx.doi.org/10.1016/j.ejca.2015.06.001>.
- [2] J. Doorbar, N. Egawa, H. Griffin, C. Kranjec, I. Murakami, Human papillomavirus molecular biology and disease association, *Rev. Med. Virol.* 25 (2015) 2–23, <http://dx.doi.org/10.1002/rmv.1822>.
- [3] N.D. Christensen, HPV disease transmission protection and control, *Microb. Cell.* 3 (2016) 475–489, <http://dx.doi.org/10.15698/mic2016.09.530>.
- [4] S. Ljubojevic, M. Skerlev, HPV-associated diseases, *Clin. Dermatol.* 32 (2014) 227–234, <http://dx.doi.org/10.1016/j.clindermatol.2013.08.007>.

- [5] C. Flake, J. Arafa, A. Hall, E. Ence, K. Howard, K. Kingsley, Screening and detection of human papillomavirus (HPV) high-risk strains HPV16 and HPV18 in saliva samples from subjects under 18 years old in Nevada: a pilot study, *BMC Oral Health* 12 (2012) 43, <http://dx.doi.org/10.1186/1472-6831-12-43>.
- [6] M.G. Hawkins, D.M. Winder, S.L.R. Ball, K. Vaughan, C. Sonnex, M. a Stanley, J.C. Sterling, P.K.C. Goon, Detection of specific HPV subtypes responsible for the pathogenesis of condylomata acuminata, *Viol. J.* 10 (2013) 1–9, <http://dx.doi.org/10.1186/1743-422X-10-137>.
- [7] J. Leszczyszyn, I. Łębski, L. Łysenko, L. Hirnle, H. Gerber, Anal warts (condylomata acuminata)—current issues and treatment modalities, *Adv. Clin. Exp. Med.* 23 (2014) 307–311, <http://www.ncbi.nlm.nih.gov/pubmed/24913124>, <http://www.advances.am.wroc.pl/pdf/2014/23/2/307.pdf>.
- [8] S. Panidis, D. Paramythiotis, V.N. Papadopoulos, A. Michalopoulos, Condylomata acuminata within perianal fistulae tracts: report of two cases, *Int. J. STD AIDS* 26 (2015) 364–366, <http://dx.doi.org/10.1177/0956462414537483>.
- [9] Y. Ma, R. Madupu, U. Karaoz, C.W. Nossa, L. Yang, S. Yooseph, P.S. Yachinski, E.L. Brodie, K.E. Nelson, Z. Pei, Human papillomavirus community in healthy persons, defined by metagenomics analysis of human microbiome project shotgun sequencing data sets, *J. Virol.* 88 (2014) 4786–4797, <http://dx.doi.org/10.1128/JVI.00093-14>.
- [10] T.L. Meiring, A.T. Salimo, B. Coetzee, H.J. Maree, J. Moodley, I.I. Hitzeroth, M.-J. Freeborough, E.P. Rybicki, A.-L. Williamson, Next-generation sequencing of cervical DNA detects human papillomavirus types not detected by commercial kits, *Viol. J.* 9 (2012) 1, <http://dx.doi.org/10.1186/1743-422X-9-164>.
- [11] Y.P. Cai, Y. Yang, B.L. Zhu, Y. Li, X.Y. Xia, R.F. Zhang, Y. Xiang, Comparison of human papillomavirus detection and genotyping with four different prime sets by PCR-sequencing, *Biomed. Environ. Sci.* 26 (2013) 40–47, <http://dx.doi.org/10.3967/0895-3988.2013.01.005>.
- [12] J.D. Siqueira, B.M. Alves, I.M. Prellwitz, C. Furtado, Â.R. Meyrelles, E.S. Machado, H.N. Seuanez, M.A. Soares, E.A. Soares, Identification of novel human papillomavirus lineages and sublineages in HIV/HPV-coinfected pregnant women by next-generation sequencing, *Virology* 493 (2016) 202–208, <http://dx.doi.org/10.1016/j.virol.2016.03.027>.
- [13] G. Gao, S.H. Johnson, J.L. Kasperbauer, B.W. Eckloff, N.M. Tombers, G. Vasmataz, D.I. Smith, Mate pair sequencing of oropharyngeal squamous cell carcinomas reveals that HPV integration occurs much less frequently than in cervical cancer, *J. Clin. Virol.* 59 (2014) 195–200, <http://dx.doi.org/10.1016/j.jcv.2013.12.006>.
- [14] M. Cullen, J.F. Boland, M. Schiffman, X. Zhang, N. Wentzensen, Q. Yang, Z. Chen, K. Yu, J. Mitchell, D. Robertson, S. Bass, L. Burdette, M. Machado, S. Ravichandran, B. Luke, M.J. Machiela, M. Andersen, M. Osentoski, M. Laptewicz, S. Wacholder, A. Feldman, T. Raine-Bennett, T. Lorey, P.E. Castle, M. Yeager, R.D. Burk, L. Mirabello, Deep sequencing of HPV16 genomes: a new high-throughput tool for exploring the carcinogenicity and natural history of HPV16 infection, *Papillomavirus Res.* 1 (2015) 3–11, <http://dx.doi.org/10.1016/j.pvr.2015.05.004>.
- [15] N.P. Ambulos, L.M. Schumaker, T.J. Mathias, R. White, J. Troyer, D. Wells, K.J. Cullen, Next-generation sequencing-based HPV genotyping assay validated in formalin-fixed, paraffin-embedded oropharyngeal and cervical cancer specimens, *J. Biomol. Tech.* 27 (2016) 46–52, <http://dx.doi.org/10.7171/jbt.16-2702-004>.
- [16] T. Agorastos, K. Chatzistamatiou, T. Katsamagkas, G. Koliopoulos, A. Daponte, T. Constantinidis, T.C. Constantinidis, Primary screening for cervical cancer based on high-risk human papillomavirus (HPV) detection and HPV 16 and HPV 18 genotyping, in comparison to cytology, *PLoS One* 10 (2015), <http://dx.doi.org/10.1371/journal.pone.0119755>.
- [17] H.C. Kitchener, K. Denton, K. Soldan, E.J. Crosbie, Developing role of HPV in cervical cancer prevention, *BMJ* 347 (2013) f4781, <http://dx.doi.org/10.1136/bmj.f4781>.
- [18] G. Orlando, M. Fasolo, F. Mazza, E. Ricci, S. Esposito, E. Frati, G.V. Zuccotti, I. Cetin, M. Gramegna, G. Rizzardini, E. Tanzi, M.C. Antonacci, I. Arcidiacono, S. Bianchi, V. Boero, G. Cambi?, E. Casolati, V. Montinaro, M. Falchetti, M. Martinelli, C. Galli, E. Bertazzoli, G. Lunghi, A. Matteelli, G. Tisi, A.M. Villa, N. Zanchetta, L. Pogliani, Risk of cervical HPV infection and prevalence of vaccine-type and other high-risk HPV types among sexually active teens and young women (13–26 years) enrolled in the VALHIDATE study, *Hum. Vaccines Immunother.* 10 (2014) 986–994, <https://doi.org/10.1159/000355959>.
- [19] W.A.A. Tjalma, C.E. Depuydt, Cervical cancer screening: which HPV test should be used—L1 or E6/E7? *Eur. J. Obstet. Gynecol. Reprod. Biol.* 170 (2013) 45–46, <http://dx.doi.org/10.1016/j.ejogrb.2013.06.027>.
- [20] M. Ajiro, Z.M. Zheng, E6E7, a novel splice isoform protein of human papillomavirus 16, stabilizes viral E6 and E7 oncoproteins via HSP90 and GRP78, *MBio* 6 (2015), <http://dx.doi.org/10.1128/mBio.02068-14>.
- [21] C. a Ramos, N. Narala, G.M. Vyas, A.M. Leen, U. Gerdemann, E.M. Sturgis, M.L. Anderson, B. Savoldo, H.E. Heslop, M.K. Brenner, C.M. Rooney, Human papillomavirus type 16 E6/E7-specific cytotoxic T lymphocytes for adoptive immunotherapy of HPV-associated malignancies, *J. Immunother.* 36 (2013) 66–76, <http://dx.doi.org/10.1097/JCI.0b013e318279652e>.
- [22] M. Orioni, P. Cristoforoni, G. Carminati, C. Stefani, S. Costa, M.T. Sandri, L. Mariani, M. Preti, E6/E7 mRNA testing for human papilloma virus-induced high-grade cervical intraepithelial disease (CIN2/CIN3): a promising perspective, *Cancermedicallscience* 9 (2015) 533, <http://dx.doi.org/10.3332/cancr.2015.533>.
- [23] S. Tan, E.G.E. de Vries, a G.J. van der Zee, S. de Jong, Anticancer drugs aimed at E6 and E7 activity in HPV-positive cervical cancer, *Curr. Cancer Drug Targets* 12 (2012) 170–184, <http://dx.doi.org/10.2174/156800912799095135>.
- [24] M. Schmitz, C. Driesch, L. Jansen, I.B. Runnebaum, M. Dürst, Non-random integration of the HPV genome in cervical cancer, *PLoS One* 7 (2012), <http://dx.doi.org/10.1371/journal.pone.0039632>.
- [25] Z. Hu, D. Zhu, W. Wang, W. Li, W. Jia, X. Zeng, W. Ding, L. Yu, X. Wang, L. Wang, H. Shen, C. Zhang, H. Liu, X. Liu, Y. Zhao, X. Fang, S. Li, W. Chen, T. Tang, A. Fu, Z. Wang, G. Chen, Q. Gao, S. Li, L. Xi, C. Wang, S. Liao, X. Ma, P. Wu, K. Li, S. Wang, J. Zhou, J. Wang, X. Xu, H. Wang, D. Ma, Genome-wide profiling of HPV integration in cervical cancer identifies clustered genomic hot spots and a potential microhomology-mediated integration mechanism, *Nat. Genet.* 47 (2015) 158–163, <http://dx.doi.org/10.1038/ng.3178>.
- [26] F. Xu, M. Cao, Q. Shi, H. Chen, Y. Wang, X. Li, Integration of the full-length HPV16 genome in cervical cancer and Caski and Siha cell lines and the possible ways of HPV integration, *Virus Genes* 50 (2015) 210–220, <http://dx.doi.org/10.1007/s11262-014-1164-7>.
- [27] S.A. Farkas, N. Milutin-Gašperov, M. Grce, T.K. Nilsson, Genome-wide DNA methylation assay reveals novel candidate biomarker genes in cervical cancer, *Epigenetics* 8 (2013) 1213–1225, <http://dx.doi.org/10.4161/epi.26346>.
- [28] B. Xu, S. Chotewutmontri, S. Wolf, U. Klos, M. Schmitz, M. Dürst, E. Schwarz, Multiplex identification of human papillomavirus 16 DNA integration sites in cervical carcinomas, *PLoS One* 8 (2013), <http://dx.doi.org/10.1371/journal.pone.0066693>.
- [29] G.L. Larsson, G. Helenius, S. Andersson, B. Sorbe, M.G. Karlsson, Prognostic impact of human papilloma virus (HPV) genotyping and HPV-16 subtyping in vaginal carcinoma, *Gynecol. Oncol.* 129 (2013) 406–411, <http://dx.doi.org/10.1016/j.ygyng.2013.02.004>.
- [30] E.J. Ulbrich, R. Aeberhard, S. Wetli, A. Busato, C. Boesch, H. Zimmermann, J. Hodler, S.E. Anderson, M. Sturzenegger, Cervical muscle area measurements in whiplash patients: acute, 3, and 6 months of follow-up, *J. Magn. Reson. Imaging* 36 (2012) 1413–1420, <http://dx.doi.org/10.1002/jmri.23769>.
- [31] A. Yemelyanova, P.E. Gravitt, B.M. Ronnett, A.F. Rositch, A. Ogurtsova, J. Seidman, R.B.S. Roden, Immunohistochemical detection of human papillomavirus capsid proteins L1 and L2 in squamous intraepithelial lesions: potential utility in diagnosis and management, *Mod. Pathol.* 26 (2013) 268–274, <http://dx.doi.org/10.1038/modpathol.2012.156>.
- [32] H.P. Nguyen, M.K. Ramirez-Fort, P.L. Rady, The biology of human papillomaviruses, *Curr. Probl. Dermatol.* 45 (2014) 19–32, <http://dx.doi.org/10.1159/000355959>.
- [33] T. Ramqvist, M. Mints, N. Tertipis, A. Näsman, M. Romanitan, T. Dalianis, Studies on human papillomavirus (HPV) 16 E2, E5 and E7 mRNA in HPV-positive tonsillar and base of tongue cancer in relation to clinical outcome and immunological parameters, *Oral Oncol.* 51 (2015) 1126–1131, <http://dx.doi.org/10.1016/j.oraloncology.2015.09.007>.
- [34] K. Akagi, J. Li, T.R. Broutian, H. Padilla-Nash, W. Xiao, B. Jiang, J.W. Rocco, T.N. Teknos, B. Kumar, D. Wangsa, D. He, T. Ried, D.E. Symer, M.L. Gillison, Genome-wide analysis of HPV integration in human cancers reveals recurrent, focal genomic instability, *Genome Res.* 24 (2014) 185–199, <http://dx.doi.org/10.1101/gr.164806.113>.
- [35] H.Y. Wang, D. Lee, S. Park, G. Kim, S. Kim, L. Han, R. Yubo, Y. Li, K.H. Park, H. Lee, Diagnostic performance of HPV E6/E7 mRNA and HPV DNA assays for the detection and screening of oncogenic human papillomavirus infection among women with cervical lesions in China, *Asian Pacific J. Cancer Prev.* 16 (2015) 7633–7640, <http://dx.doi.org/10.7314/APJCP.2015.16.17.7633>.
- [36] T.G. Magaldi, L.L. Almstead, S. Bellone, E.G. Prevatt, A.D. Santin, D. DiMaio, Primary human cervical carcinoma cells require human papillomavirus E6 and E7 expression for ongoing proliferation, *Virology* 422 (2012) 114–124, <http://dx.doi.org/10.1016/j.virol.2011.10.012>.
- [37] S.F. Jabbar, S. Park, J. Schweizer, M. Berard-Bergery, H.C. Pitot, D. Lee, P.F. Lambert, Cervical cancers require the continuous expression of the human papillomavirus type 16 E7 oncoprotein even in the presence of the viral E6 oncoprotein, *Cancer Res.* 72 (2012) 4008–4016, <http://dx.doi.org/10.1158/0008-5472.CAN-11-3085>.
- [38] N. Ramirez, F. Guerra, G. Camporeale, S. Quintana, L. Diaz, N. Cuneo, J. Villacorta Hidalgo, S. Tatti, L. Alonso, S. Borkosky, G. Prat Gay, L. Palaoro, Expressions of E2 and E7-HPV16 proteins in pre-malignant and malignant lesions of the uterine cervix, *Biotech. Histochem.* 90 (2015) 573–580, <http://dx.doi.org/10.3109/10520295.2015.1047794>.
- [39] D. Hu, J. Zhou, F. Wang, H. Shi, Y. Li, B. Li, HPV-16 E6/E7 promotes cell migration and invasion in cervical cancer via regulating cadherin switch in vitro and in vivo, *Arch. Gynecol. Obstet.* 292 (2015) 1345–1354, <http://dx.doi.org/10.1007/s00404-015-3787-x>.
- [40] T. Ma, Z. Su, L. Chen, S. Liu, N. Zhu, L. Wen, Y. Yuan, L. Lv, X. Chen, J. Huang, H. Chen, Human papillomavirus type 18 E6 and E7 genes integrate into human hepatoma derived cell line Hep G2, *PLoS One* 7 (2012), <http://dx.doi.org/10.1371/journal.pone.0037964>.
- [41] W. Villavicencio-Torres, M.Y. Lacourt-Ventura, S.C. Fonseca-Williams, M. Del Mar Gonzalez-Pons, Y. Diaz-Algori, C. Torres-Ramos, R.D. Bernabe-Dones, M.R. Cruz-Correa, Expression of Human Papillomavirus E6 and E7 genes in colorectal cancer, *Cancer Res.* 73 (2013) <http://www.embase.com/search/results?subaction=viewrecord&from=export&id=L71342920%5Cnhttps://doi.org/10.1158/1538-7445.AM2013-4776%5Cnhttp://sfx.library.uu.nl/utrecht?sid=EMBASE&issn=00085472&id=10.1158%2F1538-7445.AM2013-4776&title=Expression+of+>

- [42] A. Plesa, G. Anton, I.V. Iancu, C.C. Diaconu, I. Huica, A.D. Stanescu, D. Socolov, E. Nistor, E. Popa, M. Stoian, A. Botezatu, Molecular variants of human papilloma virus 16 E2, E4, E5, E6 and E7 genes associated with cervical neoplasia in Romanian patients, *Arch. Virol.* 159 (2014) 3305–3320, <http://dx.doi.org/10.1007/s00705-014-2199-8>.
- [43] F.X. Bosch, C. Robles, M. Diaz, M. Arbyn, I. Baussano, C. Clavel, G. Ronco, J. Dillner, M. Lehtinen, K.-U. Petry, M. Poljak, S.K. Kjaer, Meijer C.J.L.M, S.M. Garland, J. Salmerón, X. Castellsagué, L. Bruni, S. de Sanjosé, J. Cuzick, HPV-FASTER: broadening the scope for prevention of HPV-related cancer, *Nat. Rev. Clin. Oncol.* 13 (2015) 1–14, <http://dx.doi.org/10.1038/nrclinonc.2015.146>.
- [44] J.A. den Boon, D. Pyeon, S.S. Wang, M. Horswill, M. Schiffman, M. Sherman, R.E. Zuna, Z. Wang, S.M. Hewitt, R. Pearson, M. Schott, L. Chung, Q. He, P. Lambert, J. Walker, M.A. Newton, N. Wentzensen, P. Ahlquist, Molecular transitions from papillomavirus infection to cervical precancer and cancer: role of stromal estrogen receptor signaling, *Proc. Natl. Acad. Sci.* 112 (2015) E3255–E3264, <http://dx.doi.org/10.1073/pnas.1509322112>.
- [45] J.C. Gage, M. Schiffman, H.A. Katki, P.E. Castle, B. Fetterman, N. Wentzensen, N.E. Poitras, T. Lorey, L.C. Cheung, W.K. Kinney, Reassurance against future risk of precancer and cancer conferred by a negative human papillomavirus test, *J. Natl. Cancer Inst.* 106 (2014), <http://dx.doi.org/10.1093/jnci/dju153>.
- [46] C. Bodelon, S. Vinokurova, J.N. Sampson, J.A. den Boon, J.L. Walker, M.A. Horswill, K. Korthauer, M. Schiffman, M.E. Sherman, R.E. Zuna, J. Mitchell, X. Zhang, J.F. Boland, A.K. Chaturvedi, S.T. Dunn, M.A. Newton, P. Ahlquist, S.S. Wang, N. Wentzensen, Chromosomal copy number alterations and HPV integration in cervical precancer and invasive cancer, *Carcinogenesis* 37 (2015) 188–196, <http://dx.doi.org/10.1093/carcin/bgv171>.
- [47] F.A. Gourronc, W.M. Rockey, W.H. Thiel, P.H. Giangrande, A.J. Klingelutz, Identification of RNA aptamers that internalize into HPV-16 E6/E7 transformed tonsillar epithelial cells, *Virology* 446 (2013) 325–333, <http://dx.doi.org/10.1016/j.virol.2013.08.015>.
- [48] V.M. Williams, M. Filippova, V. Filippov, K.J. Payne, P. Duerksen-Hughes, Human papillomavirus type 16 E6* induces oxidative stress and DNA damage, *J. Virol.* 88 (2014) 6751–6761, <http://dx.doi.org/10.1128/JVI.03355-13>.
- [49] Y.M. Cheng, C.Y. Chou, Y.C. Hsu, M.J. Chen, L.Y.C. Wing, The role of human Papillomavirus type 16 E6/E7 oncoproteins in cervical epithelial-mesenchymal transition and carcinogenesis, *Oncol. Lett.* 3 (2012) 667–671, <http://dx.doi.org/10.3892/ol.2011.512>.
- [50] N. Azizah, U. Hashim, S.C.B. Gopinath, S. Nadzirah, Gold nanoparticle mediated method for spatially resolved deposition of DNA on nano-gapped interdigitated electrodes, and its application to the detection of the human Papillomavirus, *Microchim. Acta* (2016), <http://dx.doi.org/10.1007/s00604-016-1954-9>.
- [51] N. Azizah, U. Hashim, S.C.B. Gopinath, S. Nadzirah, A direct detection of human papillomavirus 16 genomic DNA using gold nanoprobe, *Int. J. Biol. Macromol.* 94 (2017) 571–575, <http://dx.doi.org/10.1016/j.ijbiomac.2016.10.060>.
- [52] D.K. Corrigan, H. Schulze, R.A. McDermott, I. Schmüser, G. Henihan, J.B. Henry, T.T. Bachmann, A.R. Mount, Improving electrochemical biosensor performance by understanding the influence of target DNA length on assay sensitivity, *J. Electroanal. Chem.* 732 (2014) 25–29, <http://dx.doi.org/10.1016/j.jelechem.2014.08.026>.
- [53] P.E. Gravitt, M.B. Kovacic, R. Herrero, M. Schiffman, C. Bratti, A. Hildesheim, J. Morales, M. Alfaro, M.E. Sherman, S. Wacholder, A.C. Rodriguez, R.D. Burk, High load for most high risk human papillomavirus genotypes is associated with prevalent cervical cancer precursors but only HPV16 load predicts the development of incident disease, *Int. J. Cancer* 121 (2007) 2787–2793, <http://dx.doi.org/10.1002/ijc.23012>.
- [54] S.H. Lee, V.S. Vigliotti, J.S. Vigliotti, S. Pappu, Validation of human papillomavirus genotyping by signature DNA sequence analysis, *BMC Clin. Pathol.* 9 (2009) 3, <http://dx.doi.org/10.1186/1472-6890-9-3>.
- [55] J. Ye, G. Coulouris, I. Zaretskaya, I. Cutcutache, S. Rozen, T.L. Madden, Primer-BLAST: a tool to design target-specific primers for polymerase chain reaction, *BMC Bioinf.* 13 (2012) 134, <http://dx.doi.org/10.1186/1471-2105-13-134>.
- [56] S. McGinnis, T.L. Madden, BLAST: At the core of a powerful and diverse set of sequence analysis tools, *Nucleic Acids Res.* 32 (2004), <http://dx.doi.org/10.1093/nar/gkh435>.
- [57] L. Fancello, D. Raoult, C. Desnues, Computational tools for viral metagenomics and their application in clinical research, *Virology* 434 (2012) 162–174, <http://dx.doi.org/10.1016/j.virol.2012.09.025>.
- [58] I. Agalliu, S. Gapstur, Z. Chen, T. Wang, R. Anderson, L. Teras, A. Kreimer, R. Hayes, N. Freedman, R. Burk, Associations of oral α -, β -, and γ -human papillomavirus types with risk of incident head and neck cancer, *JAMA Oncol.* 10461 (2016) 1–8, <http://dx.doi.org/10.1001/jamaoncol.2015.5504>.
- [59] M.D. Healy, Using BLAST for performing sequence alignment, *Curr. Protoc. Hum. Genet.* (2007), <http://dx.doi.org/10.1002/0471142905.hg0608s52>, Chapter 6 Unit 6.8.
- [60] H. Brzeski, An introduction to bioinformatics, *Methods Mol. Biol.* 187 (2002) 193–208, <http://dx.doi.org/10.1198/jasa.2006.s110>.
- [61] B.J. Morris, Cervical human papillomavirus screening by PCR: advantages of targeting the E6/E7 region, *Clin. Chem. Lab. Med.* 43 (2005) 1171–1177, <http://dx.doi.org/10.1515/CCLM.2005.203>.
- [62] C. Camacho, G. Coulouris, V. Avagyan, N. Ma, J. Papadopoulos, K. Bealer, T.L. Madden, BLAST+: architecture and applications, *BMC Bioinf.* 10 (2009) 421, <http://dx.doi.org/10.1186/1471-2105-10-421>.
- [63] X. Liu, A. Dakic, Y. Zhang, Y. Dai, R. Chen, R. Schlegel, HPV E6 protein interacts physically and functionally with the cellular telomerase complex, *Proc. Natl. Acad. Sci. U. S. A.* 106 (2009) 18780–18785, <http://dx.doi.org/10.1073/pnas.0906357106>.
- [64] P.E. Gravitt, The known unknowns of HPV natural history, *J. Clin. Invest.* 121 (2011) 4593–4599, <http://dx.doi.org/10.1172/JCI57149>.
- [65] M. das, G.P. Leto, G.F. Dos Santos Júnior, A.M. Porro, J. Tomimori, Human papillomavirus infection: etiopathogenesis, molecular biology and clinical manifestations, *An. Bras. Dermatol.* 86 (2011) 306–317.
- [66] T.G. Mamedov, E. Pienaar, S.E. Whitney, J.R. TerMaat, G. Carvill, R. Goliath, A. Subramanian, H.J. Viljoen, A fundamental study of the PCR amplification of GC-rich DNA templates, *Comput. Biol. Chem.* 32 (2008) 452–457, <http://dx.doi.org/10.1016/j.compbiolchem.2008.07.021>.
- [67] S. Nadzirah, N. Azizah, U. Hashim, S.C.B. Gopinath, M. Kashif, Titanium dioxide nanoparticle-based interdigitated electrodes: a novel current to voltage DNA biosensor recognizes E. coli O157:H7, *PLoS One* 10 (2015) e0139766, <http://dx.doi.org/10.1371/journal.pone.0139766>.
- [68] A.A. Odeh, Y. Al-Douri, C.H. Voon, R. Mat Ayub, S.C.B. Gopinath, R.A. Odeh, M. Ameri, A. Bouhemadou, A needle-like Cu₂CdSnS₄ alloy nanostructure-based integrated electrochemical biosensor for detecting the DNA of Dengue serotype 2, *Microchim. Acta* 184 (2017) 2211–2218, <http://dx.doi.org/10.1007/s00604-017-2249-5>.
- [69] A.S. Ibraheem, Y. Al-Douri, U. Hashim, M. Ameri, A. Bouhemadou, R. Khenata, Structural, optical and electrical investigations of Cu₂Zn_{1-x}Cd_xSnS₄/Si quaternary alloy nanostructures synthesized by spin coating technique, *Microsyst. Technol.* 23 (2017) 2223–2232, <http://dx.doi.org/10.1007/s00542-016-2986-0>.
- [70] C.Y.S. Yean, K.S. Raju, R. Xavier, S. Subramaniam, S.C.B. Gopinath, S.V. Chinni, Molecular detection of methicillin-resistant staphylococcus aureus by non-protein coding RNA-mediated monoplex polymerase chain reaction, *PLoS One* 11 (2016), <http://dx.doi.org/10.1371/journal.pone.0158736>.