



Enzyme free glucose sensing by amino-functionalized silicon quantum dot

Liqing Du^a, Zhongping Li^{a,*}, Jiaoli Yao^a, Guangming Wen^a, Chuan Dong^a, Hung-Wing Li^{b,*}

^a Institute of Environmental Science, School of Chemistry and Chemical Engineering, Shanxi University, Taiyuan 030006, PR China

^b Department of Chemistry, Hong Kong Baptist University, Kowloon Tong, Hong Kong, China

ARTICLE INFO

Article history:

Received 13 November 2018

Received in revised form 5 March 2019

Accepted 18 March 2019

Available online 19 March 2019

Keywords:

Silicon quantum dots

Enzyme free

A glucose sensor

Fluorescence quenching

ABSTRACT

Silicon quantum dots have become one of the most popular nanomaterials in biological applications for their excellent biocompatibility and optical properties. Herein, we synthesized amino-functionalized silicon quantum dots (NH₂@SiQDs) via a simple microemulsion method, in which silicon tetrachloride and allylamine were used as source of silicon and functional group. NH₂@SiQDs exhibits good water-solubility, high fluorescence quantum yield and optical stability. A non-enzymatic biosensor of glucose was developed based on the fluorescence quenching of NH₂@SiQDs in response to glucose. The fluorescence response was linearly proportional to glucose in the concentration range of 1.0×10^{-6} – 9.0×10^{-5} mol/L and the detection limit was determined to be 3.0×10^{-7} mol/L. The developed glucose sensor was successfully applied in blood glucose analysis of human serum. Satisfactory result that agreed very well with traditional method was obtained.

© 2019 Elsevier B.V. All rights reserved.

1. Introduction

Glucose is the most widely distributed and important monosaccharide in nature, and plays an important role in biological metabolisms [1–4]. Glucose has the function of strengthening memory, stimulating calcium absorption and increasing intercellular communication. High concentration of glucose however easily increases insulin levels and sequentially leads to obesity and diabetes [5,6], while a low level may induce hypoglycemia and even insulin shock (diabetic coma) [7,8]. Therefore, strict control of glucose content in the blood is essential to prevent long-term chronic diseases, including neurological diseases, cardiovascular diseases, color blindness and etc. [9,10]. To date, the detection of glucose are mainly conducted by principles of electrochemistry [11–14], spectrophotometry [15,16], capillary electrophoresis [17] and high performance liquid chromatography (HPLC) [18]. Recently, various enzymatic biosensors [19–21] have attracted increasing interests for their high sensitivity, selectivity and simplicity. However, enzyme is fragile to be inactivated, and sensitive to temperature and external conditions, which would lead to poor reproducibility in glucose sensing accuracy. Meanwhile, enzyme-based sensors are also of relatively high cost and short shelf life.

Research in developing novel nanomaterials today is attractive. People now are provided with a wide range of choices of nanomaterials based on their desired properties. For example, SiQDs are made of

non-toxic and abundant material, offering one of the broadest spectrally ranges with emission tunabilities. Furthermore, colloidal SiQDs are feasible for low-temperature deposition on any substrates (such as glass, plastic, wood, and textile) using techniques like printing, roll-to-roll, stamping, spraying, or painting. Given their bright and tunable emission, colloidal SiQDs are found useful in large-area (flexible) displays, new types of lighting, and third generation photovoltaics-as “light paint” or “solar paint”. Additionally, the nontoxicity and biodegradability of SiQDs opens up new opportunities for applications in fields of strict environmental/health concerns, for instances, in-vivo bioimaging, drug delivery, lab-on-chip sensing, optoelectronic toys, smart textiles, photocatalysis for water/waste/food sterilization, phototherapeutics, cosmetics and etc.

Silicon quantum dots (SiQDs) has excellent biocompatibility and can be readily functionalized and conjugated with biomolecules by various chemical modifications [22,23], making it suitable for labeling and detection in vivo [24–26]. Nevertheless, most native SiQDs are rather poor in water-solubility or incompatible with hydrophilic solvents due to the surface Si—H bonds [27], which limits its applications in biological and biomedical field [28,29]. However, appropriate functionalization of SiQDs would provide active morphology for beneficial electron and charge transport [30–33]. Hence, native SiQDs have to be modified with hydrophilic ligands to boost their water dispersibility and photostability, as well as fluorescence quantum yield if appropriate. In this paper, water-soluble allylamine-modified silicon quantum dots (NH₂@SiQDs) were synthesized by the micro-emulsion method [34]. They were further applied to construct a simple enzyme-free biosensor

* Corresponding authors.

E-mail address: zli04@sxu.edu.cn (Z. Li).

for sensitive glucose detection. The newly developed strategy not only eliminates the use of an enzyme, but also offers high stability, good reproducibility and cost-effectiveness for glucose sensing.

2. Experimental section

2.1. Material

Tetraoctylammonium bromide (TOAB, 98%), carbon tetrachloride (SiCl_4 , 99.5%), lithium (LiAlH_4 , 97%) and chloroplatinic acid ($\text{H}_2\text{PtCl}_6 \cdot 6\text{H}_2\text{O}$) as a reducing and catalytic agents, were obtained from Aladdin. Tetrahydrofuran (THF, allylamine ($\text{C}_3\text{H}_7\text{N}$, 99%) and isopropyl alcohol ($\text{C}_3\text{H}_8\text{O}$) were purchased respectively from Tianjin Kaitong, Chengdou Best and Beijing Chemical Company. Anhydrous toluene (C_7H_8), Dopamine (DA), ascorbic acid (AA), cysteine (Cys), glutathione (GSH) sucrose, D-fructose, glucose, sodium chloride (NaCl), potassium chloride (KCl), magnesium chloride (MgCl_2), calcium chloride (CaCl_2) were purchased from Aladdin. All other chemicals were of analytical reagent grade. Double distilled water (18.25 M Ω cm) was used throughout the experiments.

2.2. Synthesis of $\text{NH}_2\text{@SiQDs}$

In nitrogen environment, tetraoctylammonium bromide (TOAB, 1.5 g) was dissolved in anhydrous toluene (100 mL), and then mixed with 92 μL carbon tetrachloride (0.8 mmol) and twice excess reducing agent lithium aluminum hydride (1 mol/L) in tetrahydrofuran solution. The mixture was allowed to react at room temperature for 3 h. Then 20 mL anhydrous methanol was slowly added to quench excess reductant. Hydrophilic $\text{NH}_2\text{@SiQDs}$ particles were formed by modifying the surface silicon-hydrogen bonds by reacting with 2 mL allylamine and 40 μL chloroplatinic acid (H_2PtCl_6 0.05 mol/L) as catalyst in isopropanol. After surface modification, the white precipitate was collected through rotary evaporation and distillation to obtain hydrophilic $\text{NH}_2\text{@SiQDs}$. Finally, $\text{NH}_2\text{@SiQDs}$ was dissolved in double distilled water (50 mL) and further filtered by 0.22 μm filter to remove reminding TOAB. $\text{NH}_2\text{@SiQDs}$ solution was stocked at 4 $^\circ\text{C}$.

2.3. Characterization of $\text{NH}_2\text{@SiQDs}$

The prepared amino-functionalized silicon quantum dots ($\text{NH}_2\text{@SiQDs}$) in aqueous solution was dripped on a copper grid and dried over 48 h. Then the feature and size distribution of the $\text{NH}_2\text{@SiQDs}$ were measured by Transmission Electron Microscope (TEM). The absorption spectra were recorded by ultraviolet visible spectrophotometer (UV-vis) in the wavelength range of 200–600 nm. Infrared spectrum (IR) was measured by Bruker Tensor II Fourier Transform Infrared spectroscopy (Bruker, Germany). Cary Eclipse fluorescence spectrophotometer (Varian, America) was used to measure fluorescence spectra. The excitation and emission slit was fixed to be 10 nm, respectively. The fluorescence lifetime of the functionalized quantum dots was measured using a FLS 9200 fluorescence spectrometer. The fluorescence quantum yield of quantum dots was calculated by Sankaran formula:

$$\Phi_S = \Phi_R (I_S/I_R)(OD_R/O_D) (\eta_S^2/\eta_R^2) \quad (1)$$

where, Φ_R is the fluorescence quantum yield of the standard substance, I_R is the integral area under the fluorescence curve of the standard substance, and η_R is refractive index of standard substance. In this experiment, quinine sulfate in 0.1 mol/L H_2SO_4 was used as the reference to calculate the quantum yield of $\text{NH}_2\text{@SiQDs}$, in which the fluorescence quantum yield of quinine sulfate is 0.54, refractive index of water is 1.33.

2.4. Detection of glucose

Silicon quantum dots solution (700 μL) was mixed separately with different amounts of glucose solutions of 5.0 mg/mL (0, 1, 5, 10, 20, 30, 40, 50, 60, 70, 80, 90, 100 and 200 μL) in phosphate buffer (10 mmol/L, pH = 7.0). Total volume was adjusted into 3.00 mL. After the mixture was reacted at 30 $^\circ\text{C}$ for 40 min, its emission spectrum was scanned at excitation wavelength of 363 nm with fixed excitation and emission slit 10/10 nm.

3. Results and discussion

3.1. Synthesis and characterization

The transmission electron microscopy (TEM) images in Fig. 1a reveals that the $\text{NH}_2\text{@SiQDs}$ is uniformly dispersed in aqueous solution and its size is approximately 2.0 ± 0.3 nm (50 quantum dots). Fourier infrared spectroscopy (FTIR) of silicon quantum dots modified with allylamine was shown in Fig. 1b. The characteristic peaks at 1460 and 1260 cm^{-1} indicated the presence of Si-CH₂ [35]. The peaks between 2500 and 3500 cm^{-1} are attributed to the vibrational peaks of C-CH₂ and C-NH₂. The peaks between 1000 and 1100 cm^{-1} are attributed to the vibrational stretching of Si-OR. The vibrational peak of Si-H was not observed at 2100 cm^{-1} . It is suggested that amphetamine has been successfully decorated on silicon quantum dots. The fluorescence quantum yield of $\text{NH}_2\text{@SiQDs}$ is calculated to 10.06% based on fluorescence and ultraviolet measurement. Moreover, the $\text{NH}_2\text{@SiQDs}$ quantum dots have higher resistance against photobleaching as compared to a traditional fluorescent dyes, Rhodamine 6G (see Fig. 1c). Fig. 1d displays the UV-vis absorption spectra of $\text{NH}_2\text{@SiQDs}$, a broad absorption wavelength ranging from 200 to 600 nm was observed, and there were two distinct shoulder peaks at 265 nm and 310 nm. $\text{NH}_2\text{@SiQDs}$ fluoresces at peak wavelength of 455 nm with the excitation 363 nm. As displayed in Fig. 1d inset, $\text{NH}_2\text{@SiQDs}$ solution gives strong blue light under the irradiation of 365 nm ultraviolet light. Simultaneously, $\text{NH}_2\text{@SiQDs}$ shows strong photoluminescence in the visible region, while the maximum emission peak is not affected much by the excitation wavelength (Fig. 1e), which supports that the quantum dots $\text{NH}_2\text{@SiQDs}$ are appropriate candidate for applications such as bioimaging and biolabeling [36]. (See Scheme 1.)

3.2. Detection of glucose

Here, we report a simple strategy for detecting glucose based on the fluorescence quenching of silicon quantum dots by glucose. Fig. 2 shows the changes in the fluorescence emission of silicon quantum dots with response to different concentrations of glucose. It is clear that the fluorescence intensity of the $\text{NH}_2\text{@SiQDs}$ gradually declines as of glucose concentrations increases (from 0 to 100 $\mu\text{mol/L}$), suggesting that glucose has a quenching effect on the fluorescence of $\text{NH}_2\text{@SiQDs}$. Moreover, the relative fluorescence intensity of $\text{NH}_2\text{@SiQDs}$ displayed a good linear relationship with the glucose concentration. According to the Stern-Volmer equation, the linear regression equation is $F/F_0 = -0.00908[\text{C}] + 0.98997$. The linear range was 1.0×10^{-6} – 9.0×10^{-5} mol/L, and the detection limit ($S/N = 3$) was estimated to be 0.3×10^{-6} mol/L, which is lower than other reported silicon materials for fluorescence based detection.

3.3. Optimization of the detection conditions

In order to optimize the performance of $\text{NH}_2\text{@SiQDs}$ for glucose detection, we investigated the effects of reaction temperature, time, and pH value on the glucose sensing condition. The results showed that the temperature and pH value had no significant effect on the fluorescence response. The optimal temperature and pH (PBS buffer solutions) were then determined to be 30 $^\circ\text{C}$ and pH 7.0 respectively where

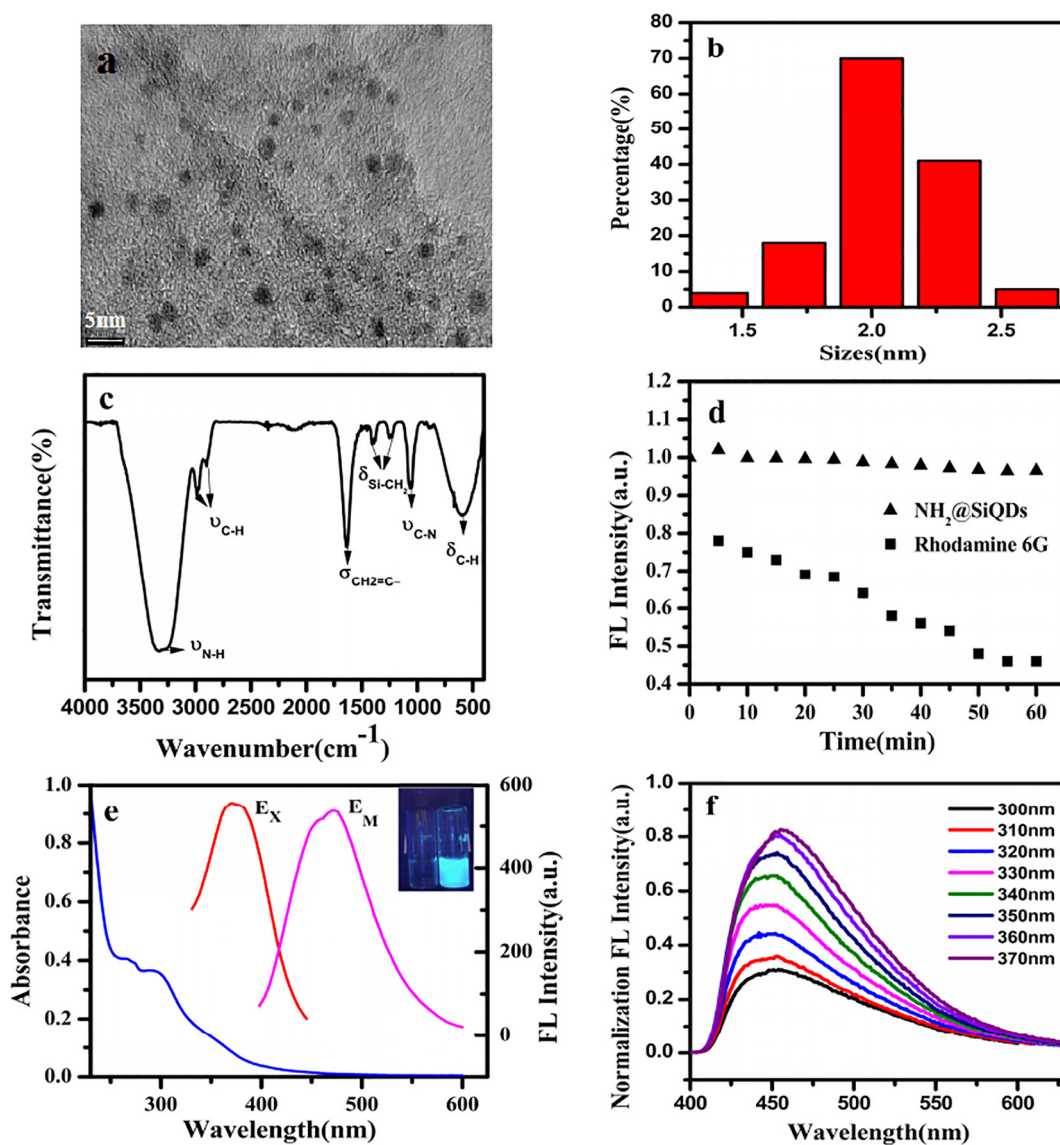
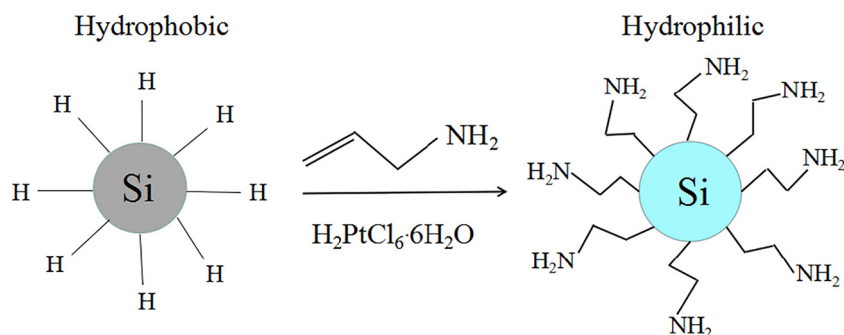


Fig. 1. (a) The TEM image of NH₂@SiQDs (inset plot of size distribution), the scale bar is 5 nm. (b) The IR image of NH₂@SiQDs. (c) Time-resolved photoluminescence decays of in aqueous solution. NH₂@SiQDs (Triangle) and Rhodamine 6G (square) in aqueous solution under the excitation wavelength at 363 nm. (d) UV-vis absorption spectrum and emission spectrum of the NH₂@SiQDs. (e) Excitation-independent FL behavior of NH₂@SiQDs.

maximal fluorescence quenching was given (Fig. 3a, c). The fluorescence signal of sensing system was stable beyond reaction time of 40 min, indicating a completed reaction (Fig. 3b). Therefore, the optimal conditions for glucose detection were performed at 30 °C, pH 7.0 and reaction time of 40 min.

3.4. Selective sensing of glucose

To evaluate the selectivity of the developed glucose biosensor, a series of major relevant carbohydrates, metal ions and amino acids were



Scheme 1. Schematic diagram of the amino-functionalized silicon quantum dot fabricated by H₂PtCl₆ catalyst.

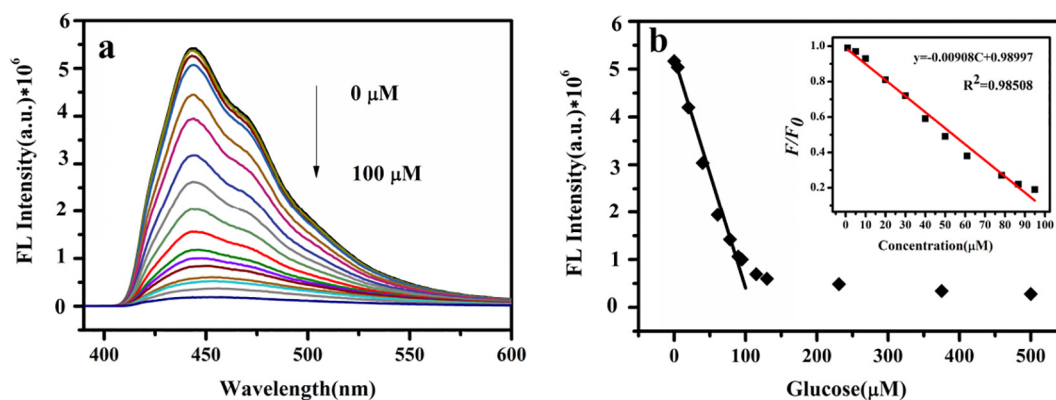


Fig. 2. (a) Fluorescence spectra of NH₂@SiQDs indicated with addition of the concentration of glucose (0–100 μmol/L). (b) The fluorescence intensity NH₂@SiQDs in the presence of glucose solution of different concentrations (excited at 363 nm). Insert: A plot of F/F₀ against the concentrations of glucose. Linear relationship was displayed.

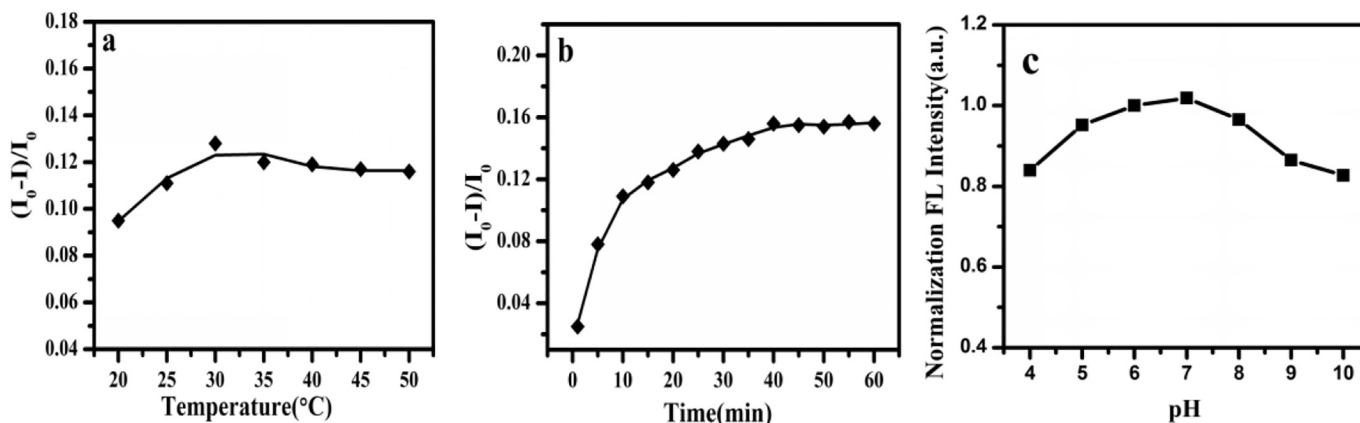


Fig. 3. The effects of temperature (a), incubation time (b), and pH value (c) on the glucose detection performance as indicated by the changes in fluorescence intensity.

adopted and studied under the optimized conditions. As elucidated in Fig. 4, several glucose analogues, such as sucrose and fructose, exhibited little influence on glucose detection even at a concentration 20 times higher than that of glucose. Common metal ions and amino acids also showed minimal interference in glucose determination. These results clearly demonstrated that the NH₂@SiQDs can serve as novel fluorescence probes for highly selective and reliable glucose monitoring.

3.5. Detection mechanism guess

TEM images of the NH₂@SiQDs in the presence of glucose were taken to investigate the sensing mechanism. Comparing the sizes of the particles before and after reaction, the TEM images (Fig. 1a and Fig. 5c) indicated the average size significantly increased in the presence of glucose. It was readily to find large “clusters” of the NH₂@SiQDs (average particle size is 3.59 nm). But the fluorescence lifetime and ultraviolet absorption spectrum of the SiQDs in the absence and presence of glucose were essentially the same, indicating no new material formation. It can be attributed to the fact that glucose molecules linked the NH₂@SiQDs together via H-bonding and hydrophobic interactions that induced the self-aggregation of the QDs and thus quenched the fluorescence (Scheme 2). Upon sonication, the linkage among NH₂@SiQDs and glucose are broken and the fluorescence of NH₂@SiQDs recovered (Fig. 5d). The native fluorescence intensity of NH₂@SiQDs recovered to 98% after 25 min of sonication.

As summarized in Table 1, the performance of the NH₂@SiQDs glucose sensor was compared with other silicon nanomaterials in the

aspects of linear range and detection limit. Compared with other enzymatic Si-based fluorescence nanosensors, this sensitive glucose sensor provides an enzyme-free, simple, direct but low-cost detection strategy for glucose detection.

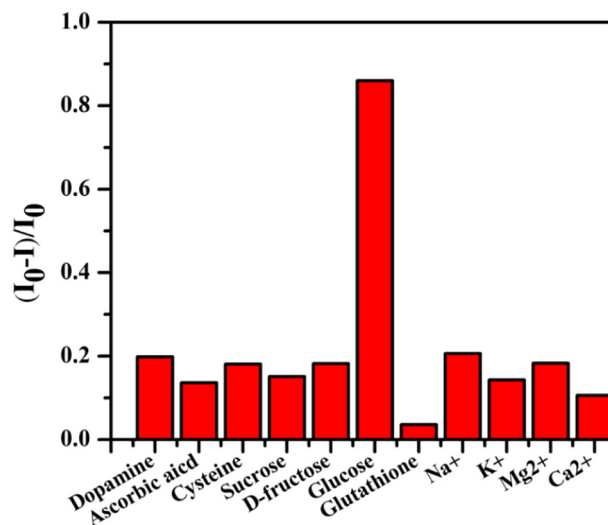


Fig. 4. Selective responses of NH₂@SiQDs to glucose over other relevant carbohydrate, metal ions and amino acids.

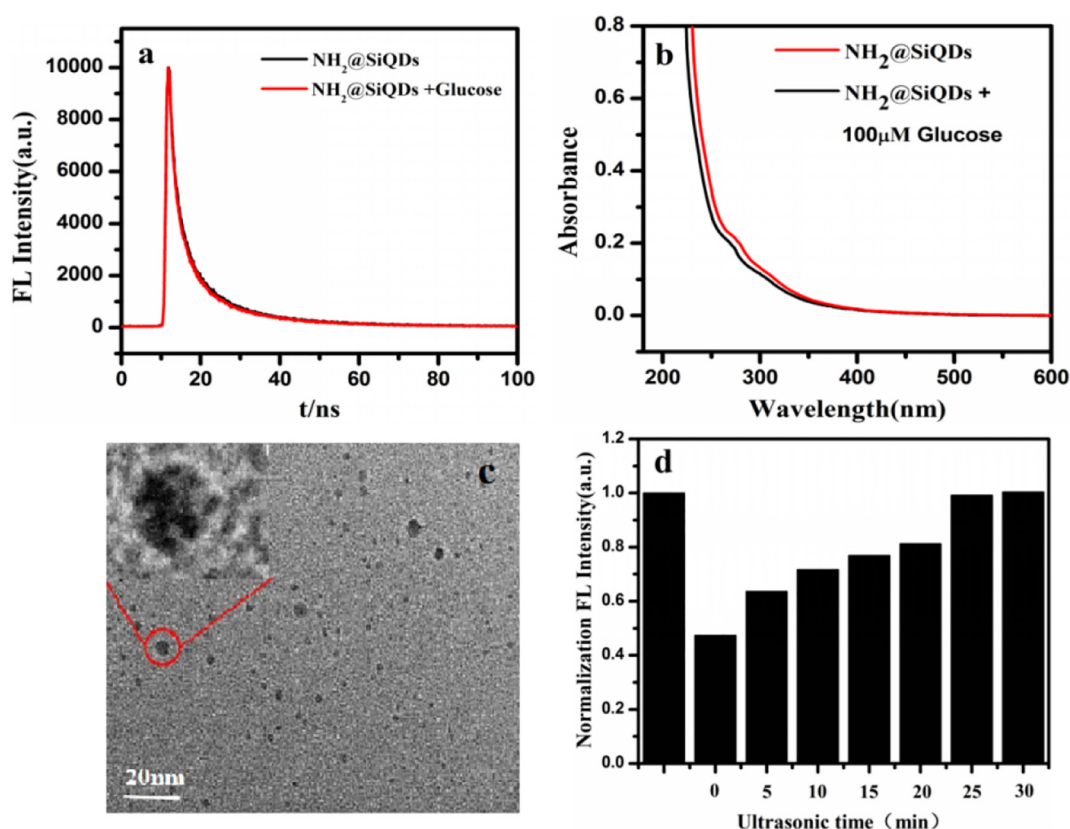


Fig. 5. (a) Photoluminescence decays of NH₂@SiQDs and NH₂@SiQDs with 0.3 mM glucose in water measured at emission wavelengths 455 nm; (b) UV-vis absorption spectra of NH₂@SiQDs and NH₂@SiQDs with 100 μM glucose; (c) TEM image of the NH₂@SiQDs with 0.3 mM glucose, the scale bar is 20 nm; (d) Fluorescence intensity of NH₂@SiQDs gradually recovered upon lengthened sonication duration (glucose concentration of 50 μM).

3.6. Real sample detection

To evaluate the practicability of NH₂@SiQDs biosensors, glucose content in human serum was determined by standard addition method using NH₂@SiQDs. The sample of human serum was provided by the

Hospital of Shanxi University and was diluted by 100 times with ultra-pure water for direct determination. As expected, the fluorescence intensity of NH₂@SiQDs was gradually reduced with increasing glucose spiked concentrations (Table 2). The recovery rate was between 92.5%–98.7%, and the RSD value is less than 7%. The result shows that



Scheme 2. Schematic illustration of the free-enzyme biosensor for sensitive glucose detection.

Table 1

Compared with reported silicon nanomaterials for glucose detection.

Method	Detection	Monitoring system	Linear range	Detect limit	Reference
Silica sol-gel	Oxygen sensing/GOx	Fluorescence Enhancement	0.1–5 mM	0.06 mM	[37]
SiQDs	GOx	Fluorescence Quenching	5–650 μM	0.68 μM	[38]
Mesoporous Si NWs	GOx	Fluorescence Enhancement	0.1–50 mM	1.06 μM	[39]
Si NPs	GOx	Fluorescence Quenching	0.01–7 mM	3.5 μM	[40]
Si-dots	GOx	Colorimetric	0.17–200 μM	0.05 μM	[41]
NH ₂ @SiQDs	Direct Detection	Fluorescence Quenching	1–90 μM	1 μM	This work

Table 2Adding standard recovery test to detect glucose in human serum ($n = 3$).

	Added (μM)	Detected (μM)	Recovery rate (%)	RSD (%)
1 ^a	0	45.8		
2	10	53.4	95.7	5.41
		52.4	93.9	
		55.1	98.7	
3	20	62.1	94.4	6.94
		61.1	92.8	
		66.0	96.5	
4	30	73.3	96.7	5.94
		70.1	92.5	
		73.7	97.2	

^a Hospital test result was 4.6 mmol/L as measured by commercial blood glucose meter.

this glucose sensor is suitable and promising for the determination of glucose in human serum samples.

4. Conclusion

In summary, we prepared amino-functionalized silicon quantum dots by microemulsion method. ($\text{NH}_2\text{@SiQDs}$). The as-prepared $\text{NH}_2\text{@SiQDs}$ possesses high water solubility, high fluorescence quantum yield and photostability. Additionally, $\text{NH}_2\text{@SiQDs}$ was demonstrated to be capable of sensing blood glucose without enzyme. The detection range of this method is 1.0×10^{-6} – 9.0×10^{-5} mol/L with a detection limit of 0.3 $\mu\text{mol/L}$. Compared to the existing electrochemical non-enzymatic glucose sensors, its superior selectivity and sensitivity are well elucidated. It provides insights for the further applications of such silicon quantum dots/electrochemical nanomaterials in non-enzymatic glucose biosensing.

Acknowledgments

This work was financially supported by Shanxi Hundred Talent Program (2015), research project supported by Shanxi Scholarship Council of China (2016–8) and the National Natural Science Foundation of China (21707082 and 21575083). We also thank the financial support of the Hong Kong Baptist University (FRG2/17–18/059).

References

- [1] S.K. Jung, J.R. Trimarchi, R.H. Sanger, et al., Development and application of a self-referencing glucose microsensor for the measurement of glucose consumption by pancreatic β -cells, *Anal. Chem.* 73 (2001) 3759–3767.
- [2] T. Pillinger, K. Beck, C. Gobjila, et al., Impaired glucose homeostasis in first-episode schizophrenia: a systematic review and meta-analysis, *JAMA psychiatry* 74 (2017) 261–269.
- [3] M.T. Gailliot, R.F. Baumeister, The physiology of willpower: linking blood glucose to self-control, *Self-Regulation and Self-Control* (2018) 137–180.
- [4] M. Pozo, M. Claret, Hypothalamic control of systemic glucose homeostasis: the pancreas connection, *Trends Endocrinol. Metab.* 29 (2018) 581–594.
- [5] K.J. Cash, H.A. Clark, Nanosensors and nanomaterials for monitoring glucose in diabetes, *Trends Mol. Med.* 16 (2010) 584–593.
- [6] E. Toschi, H. Wolpert, Utility of continuous glucose monitoring in type 1 and type 2 diabetes, *Endocrinol. Metab. Clin.* 45 (2016) 895–904.
- [7] E. Lauretti, J.G. Li, A. Di Meco, et al., Glucose deficit triggers tau pathology and synaptic dysfunction in a tauopathy mouse model, *Transl. Psychiatry* 7 (2017) e1020, <https://doi.org/10.1038/tp.2016.296>.
- [8] T. Tsujimoto, R. Yamamoto-Honda, H. Kajio, et al., Prediction of 90-day mortality in patients without diabetes by severe hypoglycemia: blood glucose level as a novel marker of severity of underlying disease, *Acta Diabetol.* 52 (2015) 307–314.
- [9] O. Courjean, N. Mano, Recombinant glucose oxidase from *Penicillium amagasakense* for efficient bioelectrochemical applications in physiological conditions, *J. Biotechnol.* 151 (2011) 122–129.
- [10] C.G. Fanelli, F. Porcellati, S. Pampanelli, et al., Insulin therapy and hypoglycaemia: the size of the problem, *Diabetes Metab. Res. Rev.* 20 (2004) S32–S42.
- [11] S.A. Zaidi, J.H. Shin, Recent developments in nanostructure based electrochemical glucose sensors, *Talanta* 149 (2016) 30–42.
- [12] O. Parlak, A. Incel, L. Uzun, et al., Structuring au nanoparticles on two-dimensional MoS_2 nanosheets for electrochemical glucose biosensors, *Biosens. Bioelectron.* 89 (2017) 545–550.
- [13] F. Xie, X. Cao, F. Qu, et al., Cobalt nitride nanowire array as an efficient electrochemical sensor for glucose and H_2O_2 detection, *Sensors Actuators B Chem.* 255 (2018) 1254–1261.
- [14] D.W. Hwang, S. Lee, M. Seo, et al., Recent advances in electrochemical non-enzymatic glucose sensors—a review, *Anal. Chim. Acta* (2018) 1–34.
- [15] S. Ramasahayam, S.H. Koppuravufi, L. Arora, S.R. Chowdhury, Noninvasive blood glucose sensing using near infra-red spectroscopy and artificial neural networks based on inverse delayed function model of neuron, *J. Med. Syst.* 39 (2014) 166.
- [16] W.-C. Shih, K.L. Bechtel, M.V. Rebec, Noninvasive glucose sensing by transcutaneous Raman spectroscopy, *J. Biomed. Opt.* 20 (2015) 51036.
- [17] R. Wu, K. Zhu, X. Zhang, et al., Nonimmobilized biomaterial capillary electrophoresis for screening drugs targeting human glucose transporter 1, *Anal. Chem.* 89 (2017) 12951–12959.
- [18] A.L. Galant, R.C. Kaufman, J.D. Wilson, Glucose: detection and analysis, *Food Chem.* 188 (2015) 149–160.
- [19] K. Tian, S. Alex, G. Siegel, et al., Enzymatic glucose sensor based on au nanoparticle and plant-like ZnO film modified electrode, *Mater. Sci. Eng. C* 46 (2015) 548–552.
- [20] A. Liu, Q. Lang, B. Liang, et al., Sensitive detection of maltose and glucose based on dual enzyme-displayed bacteria electrochemical biosensor, *Biosens. Bioelectron.* 87 (2017) 25–30.
- [21] C.A. Cordeiro, A. Sias, T. Koster, et al., In vivo “real-time” monitoring of glucose in the brain with an amperometric enzyme-based biosensor based on gold coated tungsten (W-au) microelectrodes, *Sensors Actuators B Chem.* 263 (2018) 605–613.
- [22] T. Zhou, R.T. Anderson, H. Li, et al., Bandgap tuning of silicon quantum dots by surface functionalization with conjugated organic groups, *Nano Lett.* 15 (2015) 3657–3663.
- [23] L. Wang, Q. Li, H.Y. Wang, et al., Ultrafast optical spectroscopy of surface-modified silicon quantum dots: unraveling the underlying mechanism of the ultrabright and color-tunable photoluminescence, *Light: Science & Applications*. 4 (2015) e245, <https://doi.org/10.1038/lsa.2015.18>.
- [24] J.D. Schiffman, R.G. Balakrishna, Quantum dots as fluorescent probes: synthesis, surface chemistry, energy transfer mechanisms, and application, *Sensors and Actuators B: Chemical* 258 (2017) 1191–1214.
- [25] S. Zong, J. Zong, C. Chen, et al., Single molecule localization imaging of exosomes using blinking silicon quantum dots, *Nanotechnology* 29 (2018), 065705.
- [26] K.G. Reeves, A. Schleife, A.A. Correa, et al., Role of surface termination on hot electron relaxation in silicon quantum dots: a first-principles dynamics simulation study, *Nano Lett.* 15 (2015) 6429–6433.
- [27] G.H. Pan, A. Barras, L. Boussekey, et al., Silica cross-linked micelles loading with silicon nanoparticles: preparation and characterization, *ACS Appl. Mater. Interfaces* 5 (2013) 7042–7049.
- [28] K. Lokesh, G. Kavitha, E. Manikandan, et al., Effective ammonia detection using n-ZnO/p-NiO heterostructured nanofibers[J], *IEEE Sensors J.* 16 (2016) 2477–2483.
- [29] N. Geetha, S. Sivarajani, A. Ayeshamariam, et al., High performance photo-catalyst based on nanosized ZnO-TiO_2 nanoplatelets for removal of RhB under visible light irradiation[J], *J. Adv. Microsc. Res.* 13 (2018) 12–19.
- [30] E. Manikandan, J. Kennedy, G. Kavitha, et al., Hybrid nanostructured thin-films by PLD for enhanced field emission performance for radiation micro-nano dosimetry applications[J], *J. Alloys Compd.* 647 (2015) 141–145.
- [31] A.M. Amanulla, S.K.J. Shahina, R. Sundaram, et al., Antibacterial, magnetic, optical and humidity sensor studies of $\beta\text{-CoMoO}_4\text{-Co}_3\text{O}_4$ nanocomposites and its synthesis and characterization[J], *J. Photochem. Photobiol. B Biol.* 183 (2018) 233–241.
- [32] M.V. Arularasu, R. Sundaram, C.M. Magdalane, et al., Synthesis, humidity sensing, photocatalytic and antimicrobial properties of thin film nanoporous $\text{PbWO}_4\text{-WO}_3$ nanocomposites[J], *Journal of Nanostructures* 7 (2017) 47–56.
- [33] S. Poovaragan, R. Sundaram, C.M. Magdalane, et al., Photocatalytic activity and humidity sensor studies of magnetically reusable $\text{FeWO}_4\text{-WO}_3$ composite nanoparticles[J], *J. Nanosci. Nanotechnol.* 19 (2019) 859–866.
- [34] J.H. Warner, A. Hoshino, K. Yamamoto, et al., Water-soluble Photoluminescent silicon quantum dots, *Angew. Chem. Int. Ed.* 117 (2005) 4626–4630.
- [35] T.H. Le, H.D. Jeong, Characterization of band gaps of silicon quantum dots synthesized by etching silicon nanopowder with aqueous hydrofluoric acid and nitric acid, *Bull. Kor. Chem. Soc.* 35 (2014) 1523–1528.
- [36] Y. Zhuo, H. Miao, D. Zhong, et al., One-step synthesis of high quantum-yield and excitation-independent emission carbon dots for cell imaging, *Mater. Lett.* 139 (2015) 197–200.
- [37] G. Chang, Y. Tatsu, T. Goto, et al., Glucose concentration determination based on silica sol-gel encapsulated glucose oxidase optical biosensor arrays, *Talanta* 83 (2010) 61–65.
- [38] Y. Yi, J. Deng, Y. Zhang, et al., Label-free Si quantum dots as photoluminescence probes for glucose detection, *Chem. Commun.* 49 (2013) 612–614.

- [39] R. Ghosh, R. Das, P.K. Giri, Label-free glucose detection over a wide dynamic range by mesoporous Si nanowires based on anomalous photoluminescence enhancement, *Sensors Actuators B Chem.* 260 (2018) 693–704.
- [40] L. Ding, Z. Gong, M. Yan, et al., Determination of glucose by using fluorescent silicon nanoparticles and an inner filter caused by peroxidase-induced oxidation of o-phenylenediamine by hydrogen peroxide, *Microchim. Acta* 184 (2017) 4531–4536.
- [41] Q. Chen, M. Liu, J. Zhao, et al., Water-dispersible silicon dots as a peroxidase mimetic for the highly-sensitive colorimetric detection of glucose, *Chem. Commun.* 50 (2014) 6771–6774.