



CdTe QDs based fluorescent sensor for the determination of gallic acid in tea

Xuanping Tan^a, Qin Li^b, Jidong Yang^{a, c, *}^a Chongqing Three Gorges University, Chongqing 404000, China^b Chongqing Medical and Health school, Fuling, Chongqing 408100, China^c School of Chemistry and Chemical Engineering, Southwest University, Chongqing 400715, China

ARTICLE INFO

Article history:

Received 8 March 2019

Received in revised form 3 July 2019

Accepted 7 July 2019

Available online 16 July 2019

Keywords:

Fluorescence

Light switch

KMnO₄

Gallic acid

ABSTRACT

A new fluorescent light switch method, which based on *N*-acetyl-L-cysteine capped CdTe QDs (NALC-CdTe QDs), was developed for the detection of gallic acid (GA). The QDs possess a fluorescence emission wavelength at 520 nm and with symmetric fluorescence. When KMnO₄ is added, the high fluorescence of QDs could be effectively quenched for the electron transfer process between KMnO₄ and QDs. But with the addition of GA, the fluorescence of KMnO₄-QDs system could recover for the reason that redox reaction of GA and KMnO₄. Therefore, a fluorescent light switch method could be used for GA with a detection range of 0.6–12.6 μg·mL⁻¹ and a detection limit of 0.56 ng·mL⁻¹. Furthermore, the feasibility of the proposed fluorescence biosensor in tea was also studied and satisfactory results were obtained.

© 2019 Elsevier B.V. All rights reserved.

1. Introduction

Gallic acid (GA: 3,4,5-trihydroxybenzoic acid (C₆H₂(OH)₃COOH)) is an organic acid, which exist in gallnuts, grapes, sumac, witch hazel, tea leaves, hops and oak bark [1,2]. As a strong antioxidant, GA has large numbers of biological properties include anti-mutagenic, anticarcinogenic, antioxidant activities and other potential health effects [3,4]. Therefore, it is necessary to establish selectivity and sensitivity methods for the determination of GA in the food substances, and it is also important for understanding their health benefits.

Up to know, a wide variety of methods have been developed to detect GA, for example, High Performance Liquid Chromatography (HPLC) [5], flow injection analysis [6,7], resonance light scattering [8], thin layer chromatography [9] and electrochemical methods [10–16]. Although these techniques are capable of absolute identification and have low detection limits for GA, they are rather costly, sophisticated, time-consuming, and inappropriate for on-site field analysis.

As a novel fluorescent nanomaterial, Quantum dots (QDs) have many special optical properties, including high quantum yield, high molar extinction coefficient, wide range of absorption spectrum, narrow and symmetrical fluorescence spectrum, large stokes shift and high photochemical stability [17]. Based on the above special

optical properties, much research has been devoted to the study of the synthesis and applications of quantum dots (QDs) [18–25], especially in fluorescent labeling, biological imaging and sensing system [26–28]. Among them, light switch QDs-based sensing systems have appeared as a novel type of sensors and have provided a broader platform for QDs in recent years [29–32]. In this type of sensing system, the fluorescence of the QDs is initially quenched (turned off), and recovered (turned on) in the next sequence step with analytes. It has the advantages of versatility and high selectivity in the light switch process. In this study, a new fluorescent sensor for the determination of GA based on the reversible “off-on” fluorescence change of *N*-acetyl-L-cysteine (NALC) capped CdTe QDs was build. KMnO₄ was utilized as it could efficiently quenched the fluorescence intensity of QDs (as shown in Fig. 1).

2. Experimental

2.1. Materials

All chemicals are analytically pure or above. Te power, *N*-acetyl-L-cysteine and gallic acid were produced by Sinopharm Chemical Reagent Co., Shanghai, China. CdCl₂·2.5H₂O was purchased from Shanghai Chemicals Reagent Co., Shanghai, China. NaBH₄ comes from Tianjin Huanwei Fine Chemical Co., Tianjin, China. Tris-HCl buffer solutions with different pH were prepared by mixing 0.2 mol·L⁻¹ Tris and 0.1 mol·L⁻¹ HCl in different proportions. The water used in all experiments had a resistivity greater than 18 MΩ·cm⁻¹.

* Corresponding author at: Chongqing Three Gorges University, Wanzhou, Chongqing 404000, China.

E-mail address: 1107875655@qq.com (J. Yang).

2.2. Instruments

Hitachi F-2500 spectrofluorophotometer (Hitachi Company, Tokyo, Japan), UV-2450 spectrophotometer (Tianmei Corporation, Shanghai, China). Hitachi-600 transmission electron microscopy (TEM, Hitachi Company, Japan), PHS-3C pH meter (Leici, Shanghai, China).

2.3. Synthesis of QDs

According to previous literature [33], CdTe QDs (capped by N-acetyl-L-cysteine) were synthesized by hydrothermal method with modification. In a 50 mL three-neck flask, N₂ was used as a protective gas, add in sequence with Te power (0.0255 g), which was magnetically stirred to dissolve in 10 mL of distilled water, and an excessive amount of NaBH₄ with slow speed. After a period of time, the black Te power disappeared and the reactant became a clear and transparent solution.

150 mL distilled water was added to a 250 mL three-neck flask containing 0.0488 g CdCl₂·2.5H₂O. Then 0.0574 g NALC was added with the protection of N₂ and magnetic stirring, at the same time adjusted the pH to 11.25 with NaOH solution (1 mol·L⁻¹). Under the appropriate stirring speed, add excessive H₂SO₄ (0.5 mol·L⁻¹) to NaHTe solution drop by drop, and generated H₂Te gas was loaded into the CdCl₂ solution through N₂. Orange liquid was obtained after the reflux reaction at 369 K for 1 h. The final concentration of NALC-capped CdTe QDs, as measured by the Cd²⁺ concentration, was 1.425×10^{-3} mol·L⁻¹.

2.4. Detection process

In a 5 mL colorimetric tube, 0.5 mL of prepared quantum dots, 0.5 mL of Tris-HCl buffer solution with a pH of 7.4 and KMnO₄ solution with different concentrations were added, diluted with water and shaken for 10 min, and the fluorescence of the system was determined.

Then research the effect of GA on the fluorescent restoration of the QDs-KMnO₄ system. 0.5 mL of CdTe QDs solution, 0.5 mL Tris-HCl buffer solution (pH 7.4), and a certain amount of KMnO₄, different amount of GA were sequentially added to a 5.0 mL colorimetric tube, thoroughly shaken and equilibrated for 10 min until fully mixed. The fluorescence spectra were recorded in the 480–660 nm emission wavelength range, and the fluorescence intensity of the maximum emission peak was used for quantitative analytical.

3. Results and discussion

3.1. Feasibility of the GA assay system

According to Fig. 1, the fluorescence spectrum of CdTe QDs fluorescence was narrow, and with nearly monodisperse and homogeneous, which could also be displayed by the TEM image and the absorption spectra of the QDs, as shown in Fig. S1. With the addition of KMnO₄, the photoluminescence intensity of CdTe QDs decreased obviously. For the fluorescence quenching of QDs by metal ions, it is a complicated process, and there are two main explanations [34–37]: (1) electron transfer from the photoexcited QDs to the cations bound at its surface; and (2) the formation of new nonradiative surface channels for electron annihilation within the QDs. In this work, the fluorescence quenching of QDs in the presence of KMnO₄, might be due to the nonradiative electron-hole recombination annihilation through an effective electron transfer process.

3.2. Fluorescence quenching effect of KMnO₄ on CdTe QDs

Fig. 2 shows the fluorescence emission spectra of quantum dots in the presence or absence of KMnO₄. As the concentration of

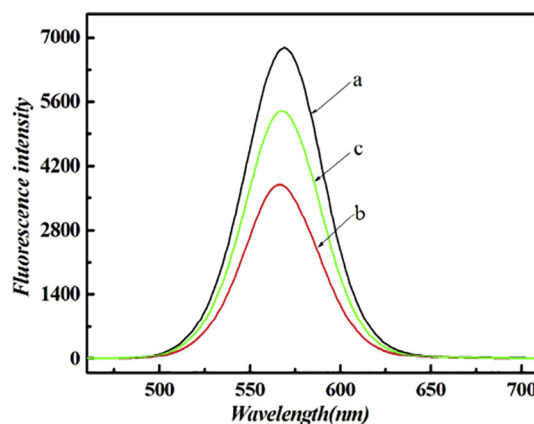


Fig. 1. The fluorescence spectra of NALC-capped CdTe QDs (curve a), NALC-capped CdTe QDs-KMnO₄ system (curve b) and NALC-capped CdTe QDs-KMnO₄ system in the presence of GA (curve c).

KMnO₄ increasing, the fluorescence intensity of CdTe QDs gradually decreased, which deduced that the QDs fluorescence quenching was mainly concentration dependent. But according to the inset of Fig. 2, with the KMnO₄ concentration increasing, the fluorescence intensity of QDs decreasing, when KMnO₄ concentration was 1.6×10^{-4} mol·L⁻¹, the degree of quenching achieve the maximum value. Therefore, 1.6×10^{-4} mol·L⁻¹ of KMnO₄ for further investigation and the GA was added to induce the fluorescence restoration of the CdTe QDs-KMnO₄ system.

3.3. Fluorescence recovery of CdTe QDs-KMnO₄ system induced by GA

GA with its strong reducing property, which could react with KMnO₄, as a result, the fluorescence of QDs was switch on after the addition of GA into the QDs-KMnO₄ system. Fig. 3A shows the fluorescence spectra of QDs-KMnO₄ complex upon the addition of various concentrations of GA. With the GA concentration increase, the fluorescence intensity of QDs-KMnO₄ complex gradually increased and could remained almost the same strength with a further increase of GA concentration. The Fig. 3B reveals a good linear correlation between $\Delta F = F_1 - F$ (F is the fluorescence intensity of QDs-KMnO₄ system, and F_1 is the fluorescence intensity of QDs-KMnO₄ system with the addition of various concentration of

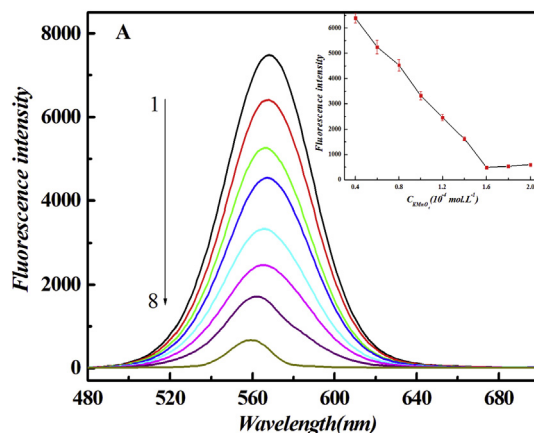


Fig. 2. (A) Fluorescence spectra of NALC-capped CdTe QDs in the presence of KMnO₄ in 0.5 mL Tris-HCl buffer solution at pH = 7.4; the inset shows the linear curve of the quenched fluorescence intensity of QDs against the concentration of KMnO₄. (NALC-capped CdTe QDs: 1.425×10^{-4} mol·L⁻¹; 0.5 mL Tris-HCl buffer solution (7.4)).

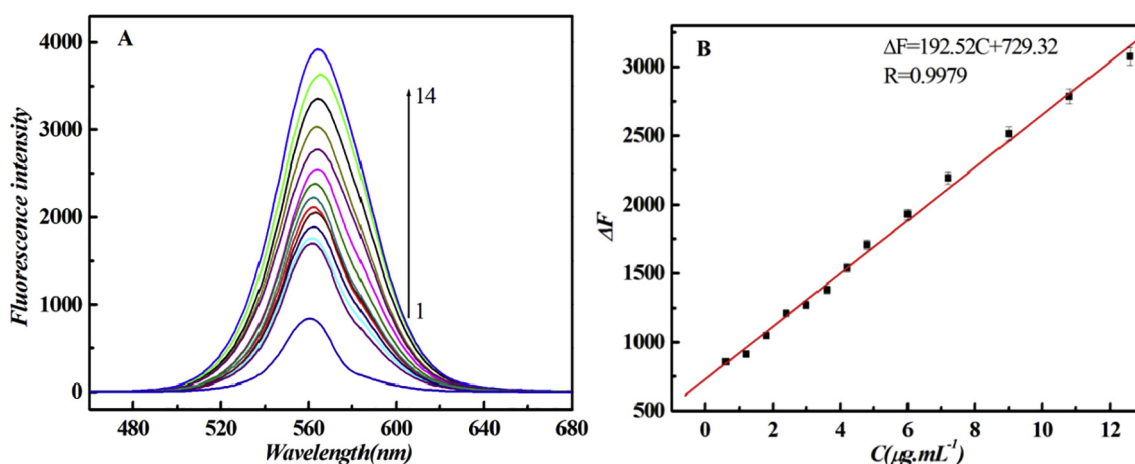


Fig. 3. (A) Fluorescence spectra of NALC-capped CdTe QDs with the addition of $1.6 \times 10^{-4} \text{ mol} \cdot \text{L}^{-1}$ of KMnO_4 recover due to introducing GA in the concentration range of $0.6\text{--}12.6 \mu\text{g} \cdot \text{mL}^{-1}$ (0.6, 1.2, 1.8, 2.4, 3.0, 3.6, 4.2, 4.8, 6.0, 7.2, 9.0, 10.8, $12.6 \mu\text{g} \cdot \text{mL}^{-1}$ respectively); (B) The linear calibration plot of the fluorescence intensity against the concentration of GA. (Concentration of NALC-capped CdTe QDs: $1.425 \times 10^{-4} \text{ mol} \cdot \text{L}^{-1}$).

GA) and GA concentration ranging from 0.6 to $12.6 \mu\text{g} \cdot \text{mL}^{-1}$. The regression equation is $\Delta F = 729.32 + 192.52C$, with the regression coefficient for 0.9979 , and the detection limit (LOD) for $0.56 \text{ ng} \cdot \text{mL}^{-1}$. As shown in Table 1, compared with the other methods before, the method in this work has lower detection limit and wider detection range.

3.4. Optimization of the method

3.4.1. Influence of buffer solution

Usually, the acidity environment is an important factor for the reaction system and the fluorescence intensity of QDs itself. By measuring the influence of buffer solution with pH value between 7.0 and 8.0 in the QDs- KMnO_4 system, it is concluded that KMnO_4 can quench the fluorescence of QDs to the maximum extent in buffer solution with pH value of 7.4 , as shown in Fig. S2A. Therefore, the optical pH of QDs- KMnO_4 system was 7.4 . And from Fig. S2B, the maximum fluorescence recovery of the QDs- KMnO_4 system after the addition of GA was obtained at pH 7.4 , which fitted well with the optimum pH (7.4) of the QDs- KMnO_4 system. Hence, the buffer solution chose Tris-HCl (7.4) in this experiment.

3.4.2. Effect of reaction time

Reaction time is an indispensable factor in the reaction system. At room temperature, the effects of different reaction times on the system were studied. By recording the experimental phenomena, it was found that the whole system reacted completely after about 5 min , and the fluorescence intensity of the system remained almost unchanged in the following 90 min , which indicating that the QDs- KMnO_4 system exhibited good stability. Thus, the fluorescence intensity of QDs- KMnO_4 system was recorded after the addition of KMnO_4 for 5 min .

The effect of the reaction time on the fluorescence recovery of QDs- KMnO_4 system after the addition of GA was also discussed.

Table 1

Comparison of the reported methods for GA.

Method	Detection limit ($\mu\text{g}/\text{mL}$)	Detection range ($\mu\text{g}/\text{mL}$)	Reference
HPLC	Not mention	$0.052\text{--}0.26$	[38]
Flow injection analysis	0.58	$0.70\text{--}100$	[39]
Chromatography	2.50	$2.50\text{--}120$	[40]
Polarography	0.28	$0.85\text{--}204$	[41]
Electrochemical methods	0.07	$0.10\text{--}10$	[42]
Fluorescence method	5.6×10^{-4}	$0.60\text{--}12.6$	This work

And the results indicated that the fluorescence recovery of QDs by GA was completed in 10 min and lasted for more than 120 min .

3.4.3. Effect of KMnO_4 concentration

For the ternary system, the addition of intermediate substances directly affects the sensitivity of the whole detection system. Theoretically, in order to provide an optimal “off” state of the biosensor, a higher concentration of quencher should be applied to completely turn off the fluorescence of CdTe QDs. However, a significant amount of quencher in the system will hamper the response sensitivity of subsequent GA detection. If the amount of KMnO_4 in the solution were excessive, some GA would react with free KMnO_4 , therefore, the reminder GA reacting with KMnO_4 , which induced the fluorescence quenching of CdTe QDs would decrease. According to the preliminary experimental results (as shown in Fig. S3), when the concentration of KMnO_4 was $1.6 \times 10^{-4} \text{ mol} \cdot \text{L}^{-1}$, the recovered fluorescence intensity induced by GA reached the maximum value. In addition, the inset of Fig. 2 shown that when the KMnO_4 was at a lower concentration, the fluorescence intensity decrease greatly, but with the increase of the concentration of KMnO_4 , the extent of the decrease in fluorescence intensity diminished. When KMnO_4 concentration was $1.6 \times 10^{-4} \text{ mol} \cdot \text{L}^{-1}$, the fluorescence intensity variation did not enhanced. Thus, $1.6 \times 10^{-4} \text{ mol} \cdot \text{L}^{-1}$ KMnO_4 was employed to be the quencher of the CdTe QDs in the biosensor preparation.

3.5. Possible reaction mechanism of the fluorescent light switch method

The fluorescence of QDs is caused by the combination of electron and hole pairs, that fluorescence quenching can be observed when the electron-hole pair recombination is blocked. This phenomenon may be due to the fact that atoms on the surface of quantum dots are cross-linked with molecules or atoms in the medium. The changes of the surface or environment of QDs would affect the efficiency of electron-hole recombination and consequently alter the optical properties of these particles. About the fluorescence quenching mechanism usually includes the dynamic quenching and static quenching [43]. Generally, in the dynamic quenching, fluorescence is subsequently quenched when the electron acceptor collides with the excited fluorophore, and charge transfer often occurred. Therefore, no changes in the absorption spectra are expected. But on the contrary, in static quenching, because of the ground state complex formation during the state

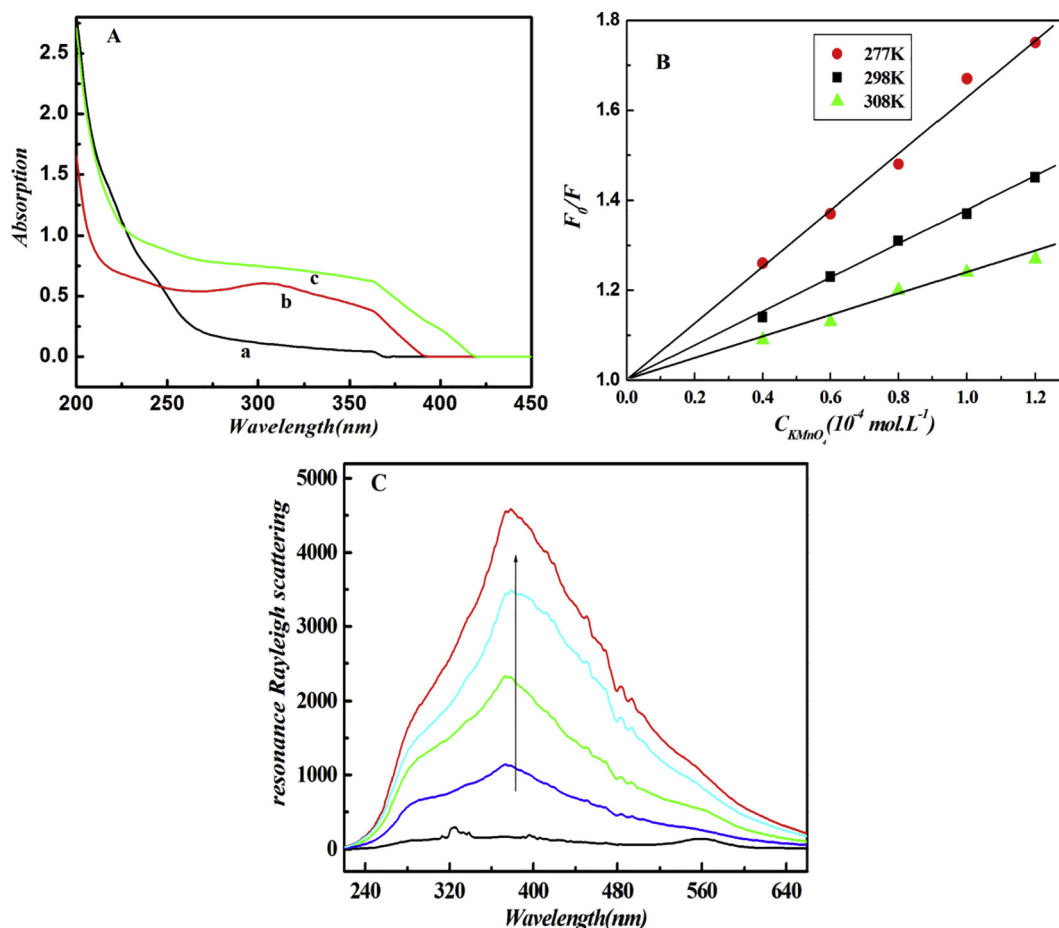


Fig. 4. (A) UV-vis absorption spectra of (a) NALC-CdTe QDs, (b) KMnO₄ (with distilled water as the reference), (c) mixture solution system (NALC-CdTe QDs and KMnO₄); (B) Stern-Volmer curves for the QDs-KMnO₄ solution system at three different temperatures (277 K, 298 K, 308 K); (C) RRS spectra of NALC-capped CdTe QDs in the presence of KMnO₄ in pH 7.4 Tris-HCl buffer solution.

quenching, the absorption spectra of the fluorophore will be perturbed. In order to precisely reveal the fluorescence quenching mechanism, the UV-vis absorption spectra of KMnO₄, QDs and KMnO₄-QDs system were recorded. As shown in Fig. 4A, the UV-vis absorption spectrum of QDs in the presence of KMnO₄ was higher, a bit flatter and with an observed red shift when compared with the single UV-vis spectrum of QDs, which indicated that the changes in size of surface properties of the QDs. Namely, the quenching type is static quenching. On the other hand, the well-known Stern-Volmer equation ($F_0/F = 1 + K_{sv}[Q]$) is also a way to distinguish static quenching and dynamic quenching. In the equation, F_0 and F represent the fluorescence intensity of QDs in the absence and presence of quencher (KMnO₄), respectively. $[Q]$ is the concentration of quencher. K_{sv} is the Stern-Volmer quenching constant. According to Fig. 4B, the linear plots for the QDs-KMnO₄ solution system at three different temperatures (277 K, 298 K, 308 K) were investigated. It can be seen that the quenching constants decreased with rise of temperature, which indicates that the probable quenching mechanism is static quenching.

The RRS scattering intensity of both KMnO₄ solution and QDs solution is very weak, but when the two are mixed together, the RRS scattering of the whole reaction system is significantly enhanced, and its maximum scattering peak is at 370 nm. The influence of solution ionic strength on the RRS strength of the whole reaction system was observed by adding NaCl solution (ion enhancer) to the reaction system. The results indicated that with the ionic strength enhanced, the RRS intensity of the system increased. Therefore, the electrostatic attraction played an

important role in the reaction of KMnO₄ and NALC-CdTe QDs, which further prove the quenching type is static quenching.

On account of electron transfer of KMnO₄ to the surface of QDs, the QDs are then switched to the fluorescence “off” status. But the fluorescence of QDs could switch “on” again attribute to the redox and coordination reaction of KMnO₄ with GA. As shown in Fig. S4A,

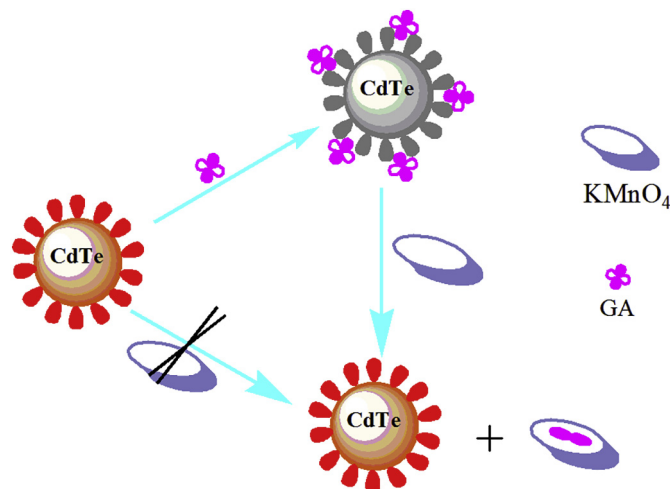


Fig. 5. The mechanism of the proposed “off-on” fluorescence biosensor.

Table 2

Determination of GA in green tea and black tea.

Sample	Found ^a (mg/g)	Added (μg/mL)	Found (μg/mL)	Recovery (%)	RSD (%)
Green tea	8.71	2.00	2.87, 2.89, 2.84, 2.79, 2.82	97.34	1.40
	8.56	4.00	4.76, 4.90, 4.97, 4.94, 4.72	99.19	2.28
	8.68	6.00	6.98, 6.73, 6.85, 6.80, 6.78	95.72	1.40
Back tea	5.25	2.00	2.48, 2.42, 2.53, 2.45, 2.49	93.87	1.68
	4.92	4.00	4.59, 4.53, 4.43, 4.47, 4.35	93.86	2.01
	4.98	6.00	6.42, 6.61, 6.38, 6.57, 6.60	102.18	1.65

^a The content of GA in every gram tea.

the fluorescence emission spectrum of QDs was not influenced by the addition of GA, which indicated that GA had no obvious influence on the fluorescence intensity of CdTe QDs, and further illustrated that the recovered fluorescence of the “turn-off on” system was caused by the interaction of KMnO₄ and GA. In addition, the absorption spectra of QDs (a), KMnO₄-QDs (b) and QDs-KMnO₄-GA system (c) (shown in Fig. S4B) were recorded. With the result that the addition of GA caused the decrease of KMnO₄-QDs absorption intensity, which also verify that the fluorescence recovery of the “turn-off on” system was caused by the interaction of KMnO₄ with GA through redox reaction. The worked principle of the biosensor was shown in Fig. 5.

3.6. Selectivity of the light switch fluorescence biosensor

Selectivity is crucial in measuring a test method. The effects of some common substances, such as amino acid, anion and cation, on fluorescence were investigated, and the results are shown in Fig. S5. Assuming the fluorescence intensity of NALC-CdTe QDs was 100% and KMnO₄-CdTe QDs is zero, GA exhibited a significant fluorescence enhancing effect on the proposed light switch biosensor. Cysteine and tryptophan could also recover the fluorescence of KMnO₄-CdTe QDs system, but compared with GA, the recovered efficiency of cysteine and tryptophan are strikingly lower. For the other kinds of amino acids, anion and cation, the recovered efficiencies are very low. Therefore, this proposed method had high selectivity of GA and can be used for the determination of GA.

3.7. Application of the proposed light switch biosensor

The developed fluorescent light switch method was used to determination of the content of GA in black and green tea to evaluate the feasibility. Weighed 1.0 g green and black tea accurately, then soaked them with 20 mL of boiling water three times, and finally filtered and transferred the samples to a 100 mL flask. The extract was diluted to 100 mL as the stock sample solution. For the determination with the proposed fluorescent light switch method, 0.5 mL of the sample solution was taken. The standard addition method was used for the determination of GA in the black and green tea real samples. As the results shown in Table 2, which indicated that the amount of GA in the green tea was higher than that of black tea. In addition, according to adding different amount of GA standards in the real samples, the recovery test had been performed. Based on the results shown in Table 2, the new light switch fluorescence biosensor was sensitive and accurate for the GA detection.

4. Conclusion

Summarily, a new fluorescent light switch method was fabricated in Tris-HCl buffer solution for the determination of GA. NALC-CdTe QDs with obviously fluorescence were synthesized successfully. After addition of KMnO₄, the fluorescence of QDs was

quenched, but surprisingly when added GA, the quenched fluorescence could be recovered in the system. Therefore, a novel and simple fluorescent light switch was developed to detect GA in tea. The new method presented a high performance, high sensitivity and selectivity in a wide range of concentration.

Acknowledgements

This work was supported by the youth project of Chongqing Chongqing Three Gorges University (18QN03), the National Natural Science Foundation of China (No. 21475014) and all authors here express their deep thanks.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.saa.2019.117356>.

References

- [1] D.M. Stanković, M. Ognjanović, F. Martin, L. Švorc, J.F.M.L. Mariano, B. Antić, Design of titanium nitride- and wolfram carbide-doped RGO/GC electrodes for determination of gallic acid, *Anal. Biochem.* 539 (2017) 104.
- [2] Y.Y. Wang, X.Y. Zhu, F. Ding, Y.Q. Liu, L. Yang, P. Zou, Q.B. Zhao, X.X. Wang, H.B. Rao, Colorimetric detection of gallic acid based on the enhanced oxidase-like activity of floral-like magnetic Fe₃O₄@MnO₂, *Luminescence* 34 (2019) 55.
- [3] I. Mudnic, D. Modun, V. Rastija, J. Vukovic, I. Brizic, V. Katalinic, B. Kozina, M. Medic-Saric, M. Boban, *Food Chem.* 119 (2010) 1205.
- [4] X.X. Wang, J.W. Wang, N.J. Yang, Flow injection chemiluminescent detection of gallic acid in olive fruits, *Food Chem.* 105 (2007) 340.
- [5] Y.G. Zuo, H. Chen, Y.W. Deng, Simultaneous determination of catechins, caffeine and gallic acids in green, Oolong, black and pu-erh teas using HPLC with a photodiode array detector, *Talanta* 57 (2002) 307.
- [6] S.F. Li, H.M. Sun, D. Wang, L. Qian, Y. Zhu, S.J. Tao, Determination of gallic acid by flow injection analysis based on luminol-AgNO₃-Ag NPs chemiluminescence system, *Chin. J. Chem.* 30 (2012) 837.
- [7] X.Q. Lin, F. Li, Y.Q. Pang, H. Cui, Flow injection analysis of gallic acid with inhibited electrochemiluminescence detection, *Anal. Bioanal. Chem.* 378 (2004) 2028.
- [8] A. Andreu-Navarro, J.M. Fernández-Romero, A. Gómez-Hens, Determination of antioxidant additives in foodstuffs by direct measurement of gold nanoparticle formation using resonance light scattering detection, *Anal. Chim. Acta* 695 (2011) 11.
- [9] K. Dhalwal, V.M. Shinde, Y.S. Biradar, K.R. Mahadik, Simultaneous quantification of bergenia, catechin, and gallic acid from *Bergenia ciliata* and *Bergenia ligulata* by using thin-layer chromatography, *J. Food Compos. Anal.* 21 (2008) 496.
- [10] L.P. Souza, F. Calegari, A.J.G. Zarbin, L.H.M. Junior, M.F. Bergamini, Voltammetric determination of the antioxidant capacity in wine samples using a carbon nanotube modified electrode, *J. Agric. Food Chem.* 59 (2011) 7620.
- [11] J.H. Luo, B.L. Li, N.B. Li, H.Q. Luo, Sensitive detection of gallic acid based on polyethyleneimine-functionalized graphene modified glassy carbon electrode, *Sensors Actuators B Chem.* 186 (2013) 84.
- [12] R. Abdel-Hamid, F. Emad Newair, Adsorptive stripping voltammetric determination of gallic acid using an electrochemical sensor based on polyepinephrine/glassy carbon electrode and its determination in black tea sample, *J. Electroanal. Chem.* 704 (2013) 32.
- [13] V. Carralero Sanz, M. Luz Mena, A. Gonzalez-Cortes, P. Yanez-Sedeno, J.M. Pingarron, Development of a tyrosinase biosensor based on gold nanoparticles-modified glassy carbon electrodes: application to the measurement of a bioelectrochemical polyphenols index in wines, *Anal. Chim. Acta* 528 (2005) 1.
- [14] S.M. Ghoreishi, M. Behpour, M. Khayatkhani, M.H. Motaghefifard, Simultaneous determination of ellagic and gallic acid in *Punica granatum*, *Myrtus communis* and *Itriphala* formulation by an electrochemical sensor based on a carbon paste electrode modified with multi-walled carbon nanotubes, *Anal. Methods* 3 (2011) 636.
- [15] N.S. Sangeetha, S. Sriman Narayanan, A novel bimediate amperometric sensor for electrocatalytic oxidation of gallic acid and reduction of hydrogen peroxide, *Anal. Chim. Acta* 828 (2014) 34.
- [16] J. Tashkhourian, S.F. Nami-Ana, S. Hashemnia, M.R. Hormozi-Nezhad, Construction of a modified carbon paste electrode based on TiO₂ nanoparticles for the determination of gallic acid, *J. Solid State Electrochem.* 17 (2013) 157.
- [17] C.J. Murphy, Peer reviewed: optical sensing with quantum dots, *Anal. Chem.* 74 (2002) 520A.
- [18] I.L. Medintz, H. Mattoussi, Quantum dot-based resonance energy transfer and its growing application in biology, *Phys. Chem. Chem. Phys.* 11 (2009) 17–45.
- [19] S. Kundu, S. Bhattacharyya, A. Patra, Photoinduced energy transfer in dye encapsulated polymer nanoparticle-CdTe quantum dot light harvesting assemblies, *Mater. Horiz.* 2 (2015) 60–67.

- [20] S. Saha, D. Majhi, K. Bhattacharyya, N. Preeyanka, A. Dattab, M. Sarkar, Evidence of homo-FRET in quantum dot–dye heterostructured assembly, *Phys. Chem. Chem. Phys.* 20 (2018) 9523–9535.
- [21] A. Ray, S. Bhattacharya, A. Bauri, Exhibition of Förster resonance energy transfer from CdSe/ZnS quantum dots to zinc porphyrin studied in solution, *J. Mol. Liq.* 276 (2019) 770–778.
- [22] S. Silvi, and A. Credi, Luminescent sensors based on quantum dot-molecule conjugates, *Chem. Soc. Rev.* 2015, 44, 4275–4289.
- [23] A. Ray, A. Bauri, S. Bhattacharya, Study of chemical physics on energy transfer phenomenon between quantum dots and a designed diporphyrin in solution, *J. Mol. Liq.* 263 (2018) 64–71.
- [24] A. Ray, A. De, S. Bhattacharya, Study of energy transfer phenomenon between quantum dots and zinc porphyrin in solution, *J. Mol. Liq.* 246 (2017) 17–24.
- [25] Y.R. Jiang, S.G. Yong Cho, M. Shim, Light-emitting diodes of colloidal quantum dots and nanorod heterostructures for future emissive displays, *J. Mater. Chem. C* 6 (2018) 2618–2634.
- [26] J.K. Jaiswal, S.M. Simon, Potentials and pitfalls of fluorescent quantum dots for biological imaging, *Trends Cell Biol.* 14 (2004) 497.
- [27] U. Resch-Genger, M. Grabolle, S. Cavaliere-Jaricot, R. Nitschke, T. Nann, Quantum dots versus organic dyes as fluorescent labels, *Nat. Methods* 2008, 5, 763.
- [28] Q. Ma, X.G. Su, Recent advances and applications in QDs-based sensors, *Analyst* 136 (2011) 4883.
- [29] Y. Liu, C.M. Deng, L. Tang, A.J. Qin, R.R. Hu, J.Z. Sun, B.Z. Tang, Specific detection of D-glucose by a tetraphenylethene-based fluorescent sensor, *J. Am. Chem. Soc.* 133 (2011) 660.
- [30] P. Wu, X. Yan, Ni²⁺-modulated homocysteine-capped CdTe quantum dots as a turn-on photoluminescent sensor for detecting histidine in biological fluids, *Biosens. Bioelectron.* 26 (2010) 485.
- [31] X.P. Tan, Q. Li, X.N. Zhang, Y.Z. Shen, J.D. Yang, A novel and sensitive turn-on fluorescent biosensor for the determination of thioctic acid based on Cu²⁺-modulated N-acetyl-L-cysteine capped CdTe quantum dots, *RSC Adv.* 5 (2015), 44173.
- [32] X.P. Tan, J.D. Yang, Q. Li, Q. Yang, Detection of glutathione with an “off–on” fluorescent biosensor based on N-acetyl-L-cysteine capped CdTe quantum dots, *Analyst* 140 (2015) 6748.
- [33] Q. Xiao, S. Huang, W. Su, W.H. Chan, Y. Liu, Facile synthesis and characterization of highly fluorescent and biocompatible N-acetyl-L-cysteine capped CdTe/CdS/ZnS core/shell/shell quantum dots in aqueous phase, *Nanotechnology* 23 (2012), 495717.
- [34] Y.F. Chen, Z. Rosenzweig, Luminescent CdS quantum dots as selective ion probes, *Anal. Chem.* 74 (2002) 5132.
- [35] P. Wu, Y. Li, X.P. Yan, CdTe quantum dots (QDs) based kinetic discrimination of Fe²⁺ and Fe³⁺, and CdTe QDs-Fenton hybrid system for sensitive photoluminescent detection of Fe²⁺, *Anal. Chem.* 81 (2009) 6252.
- [36] A.V. Isarov, Optical and photochemical properties of nonstoichiometric cadmium sulfide nanoparticles: surface modification with copper(II) ions, *J. Chryschoos, Langmuir.* 13 (1997) 3142.
- [37] L. Zhou, Y. Lin, Z. Huang, J. Ren, X. Qu, Carbon nanodots as fluorescence probes for rapid, sensitive, and label-free detection of Hg²⁺ and biothiols in complex matrices, *Chem. Commun.* 48 (2012) 1147 (C. Bo, Z. Ping, *Anal. Bioanal. Chem.* 2005, 381, 986).
- [38] D.S. Zhou, L.H. Zhang, W.T. Liao, W.J. Ni, C.Y. Guo, J. Long, Y.J. Liong, Y. Zeng, Determination of gallic acid and ellagic acid in platycarya strobilacea insect tea by HPLC, *Journal of Guangdong Pharmaceutical University* (2019), <https://doi.org/10.16809/j.cnki.2096-3653.2018100103>.
- [39] T.S. Ma, J.L. Zhang, H.T. Zhu, T.F. Wang, T. Wang, Y.M. Zhou, Determination of gallic acid with H₂O₂-fluorescence chemiluminescence system, *Journal of Henan University(Natural Science)* 6 (2016) 712–716.
- [40] J.P. Aucamp, Y. Hara, Z. Apostolides, Simultaneous analysis of tea catechins, caffeine, gallic acid, theanine and ascorbic acid by micellar electrokinetic capillary chromatography, *J. Chromatogr. A* 876 (2000) 235–242.
- [41] N.S. Sangeetha, S.S. Narayanan, A novel bimediant amperometric sensor for electrocatalytic oxidation of gallic acid and reduction of hydrogen peroxide, *Anal. Chim. Acta* 828 (2014) 34–45.
- [42] J.H. Luo, B.L. Li, N.B. Li, H.Q. Luo, Sensitive detection of gallic acid based on polyethyleneimine-functionalized graphene modified glassy carbon electrode, *Sensors Actuators B* 186 (2013) 84–89.
- [43] Y. Chen, Z. Rosenzweig, Luminescent CdS quantum dots as selective ion probes, *Anal. Chem.* 74 (2002) 5132.