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Recent biomedical applications of gold nanoparticles: A review

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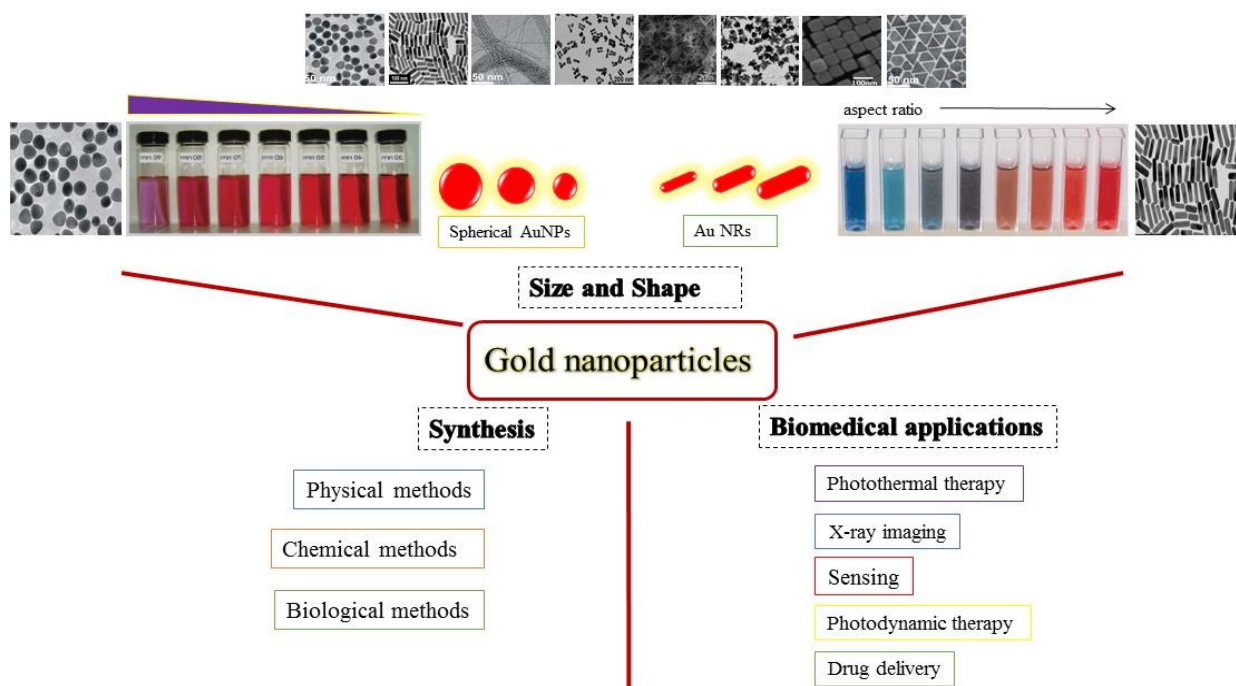
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ABSTRACT

Recent advances in nanotechnology are as a result of the development of engineered nanoparticles. Efficiently, metallic nanoparticles have been widely exploited for biomedical application and among them, gold nanoparticles (AuNPs) are highly remarkable. Consequent upon their significant nature, spherical and gold nanorods (Au NRs) nanoparticles attract extreme attention. Their intrinsic features such as optical, electronic, physicochemical and, surface plasmon resonance (SPR); which can be altered by changing the characterizations of particles such as shape, size, aspect ratio, or environment; ease of synthesis and functionalization properties have resulted to various applications in different fields of biomedicine such as sensing, targeted drug delivery, imaging, photothermal and photodynamic therapy as well as the modulation of two or three applications. This article reviewed the popular AuNPs synthesis methods and mentioned their established applications in various demands, especially in biological sensing.

Graphical abstract



Keywords: Gold nanoparticles, gold nanorods, properties, synthesis, biomedical applications, biosensor.

1. Introduction

Recent studies have shown the significant roles of nanomaterials in the progress of nanoscience and nanotechnology [1, 2]. In comparison to either small molecules or bulk materials, nanoscale structures express various physical and chemical properties. Nanoparticles possess particular intrinsic reactivity as a result of increased surface area, so an appropriate choice of materials for the manufacture of nanoparticle-based therapeutics would be made. The surface functionalities and depending on the particle size, shape and state of aggregation, the interaction will occur between nanomaterials and biological systems in various ways depending on the cell type, employing different uptake routes or targeting different organelles [3]. Among the different types of nanomaterials, metal nanoparticles especially AuNPs have attracted tremendous interests [4] from different fields of science, due to their particular features: high X-ray absorption coefficient, ease of synthetic manipulation, enabling precise control over the

particle's physicochemical properties [5], strong binding affinity to thiols, disulfides and amines [1], unique tunable optical [6] and distinct electronic properties [4]. Since the early twentieth century, various investigations have been conducted on the existence of anisotropic AuNPs. The structural, optical, electronic, magnetic, and catalytic properties of anisotropic AuNPs are different from, and most often superior to spherical gold nanoparticles [7].

Based on dimensions, AuNPs can be divided into three parts: a) one-dimensional AuNPs: nanorods, nanowires, nanotubes, nanobelts b) two dimensional AuNPs: gold nanoplates such as stars, pentagons, squares/rectangles, dimpled nanoplates, hexagons, truncated triangles and, c) three dimensional AuNPs: gold nanotadpoles, gold nanodumbbells (AuNDs), branched AuNPs such as nanopods, nanostars and gold nanodendrites. The anisotropy of these nonspherical, hollow, and nanoshell AuNP structures is the source of plasmon absorption in the visible region as well as in the near infrared (NIR) region, which is especially sensitive to the AuNP shape. This property has given rise to medical applications such as diagnostics and therapy [8]. Spherical or quasi-spherical gold nanoparticles have received the most attention because of the ease of synthesizing such structures [9]. Besides, Au NRs as a kind of elongated AuNPs, have attracted a great deal of scientific interest due to their anisotropic shape, display two separate SPR bands, as well as transverse (TSPR) and longitudinal plasmon bands (LSPR) [9]. The transversal band and the longitudinal band occur in the visible region and in the near infrared region (NIR) respectively [10]. The absorption in the NIR region produces maximal penetration of light in tissues. This makes the Au NRs suitable for *in vivo* applications. In addition, the converting of luminous energy into heat by localized SPR of Au NRs makes them an appropriate candidate for photothermal therapy [10]. Unique intrinsic features of these type of metals nanoparticles are fundamental to major biomedical applications. Optical, electronic, physicochemical properties and ease of synthesis of Au NRs provide an appropriate situation for nanotechnology improvement. Particularly, optical properties of Au NRs are tunable with their shape, leading to sensing, imaging, and biomedical therapeutic applications [11].

2. Properties of AuNPs

Explicit physical and chemical attributes make AuNPs prodigious scaffolds for various applications in therapeutics, detection and diagnostics, biolabeling, drug delivery [6, 12], chemical and biological sensing, imaging [13], nonlinear optics, photovoltaics, and catalysis fields [4, 14]. The dominant features of AuNPs include shape-related optoelectronic properties, large surface-to-volume ratio, excellent biocompatibility and low toxicity, these properties make them outstanding tools in biotechnology [15]. The surface plasmon resonance (SPR) band of AuNPs, as well as their important physical properties [15, 16], depend on their morphology (shape, solvent, surface ligand, core charge, temperature) and physiology [15, 17].

The intrinsic optical properties of AuNPs provide the opportunity of being composite therapeutic agents in the clinic. The morphology and physiology of AuNPs is associated with optical features. Changing the size of the nanoparticles, affects the color of colloidal AuNPs. This trait utilizes the fundamental of colorimetric detection forms [18]. In aqueous solution, spherical AuNPs demonstrate (present) a range of colors (orange, red to purple) as there is growth of core size from 1 to 100 nm with the exhibition of size relative absorption peak from 500 to 550 nm [15].

In recent times, researchers have focused on developing methods for the synthesis of different controllable particles shapes and sizes (Fig. 1). As the pictures indicate, a variety of

morphologies and sizes of the nanoparticles have specific optical property. It may turn in both the visible and near-infrared spectrum (Fig. 2.) [19].

3. Synthesis of AuNPs

To date, there are numerous preparative methods for the synthesis of AuNPs, containing top-down and bottom-up procedures (Fig. 3.). Recent efforts have been dedicated to controllable approaches involving size, shape, solubility, stability, and functionality [15].

Generally, procedures for the synthesis of Au NPs can be arranged into chemical, physical and biological methods.

(i) Methods such as the γ -irradiation method, microwave (MW) irradiation, sonochemical method, ultraviolet (UV) radiation, laser ablation, thermolytic process and photochemical process are categorized as physical procedures [17]. The γ -irradiation method renders controllable size and high purity of nanoparticles with a diameter of 5 - 40 nm. In the microwave irradiation method, heating or photochemical reduction method was utilized to form Au NPs [27]. The sonochemical method is capable of providing Au/Pd bimetallic particles and AuNPs within the pores of silica [28]. High temperature and ultraviolet (UV) radiation are the main reducing agents in thermolytic and ultraviolet (UV) radiation methods of AuNPs synthesis. By optimizing the synthesis condition, various sizes were achieved [29]. Photochemical process provides Au NRs. These one-dimensional nanoparticles possess extremely complex physical properties and self-assembly behaviors compared to those of spherical nanoparticles [30]. The laser ablation method can be used to produce AuNPs with tunable features. The reduction of gold (III) tetrachloroaurate is performed by the photo-induced effects of a 532 nm wavelength laser beam.

(ii) In chemical methods, chemical reactions are performed in an aqueous medium by a reduction agent. Citrate and sodium borohydride are the common reducing agents used. Among the conventional methods of chemical synthesis of colloidal gold, the Turkevitch procedure is highly utilized due to the simplicity and ease of synthesis, controllable size, and stability of colloidal nanoparticles [31]. In 1951, Hauser and Lynn 's preparation was modified by Turkevich. In the standardized procedure, sodium citrate reduced and stabilized the solution. It provides spherical AuNPs with a narrow size distribution [32]. The Brust–Schiffrin method expressed in 1994, utilized potent thiol–gold interactions to protect AuNPs with thiol ligands [14]. In the presence of alkanethiols, AuCl_4^- was reduced by sodium borohydride. This reaction was performed in two-phases (water-toluene) and nanoparticles with diameters in the range 1-3 nm were produced [33].

(iii) Biological synthesis known as favorable environment is another route of synthesis [34]. Reducing hazardous generated wastes and assisting “green chemistry” are the main objectives of biosynthesis. Solvent medium, reducing and stabilizer agents should be nontoxic and safe [35]. Plant-based compounds and derivatives, bacteria, fungi, algae, yeast, and viruses are employed as the common resources [31, 35-37]. As shown in Table 1, various shapes and sizes of nanoparticles would be obtained from these resources which have many effective applications in biomedical sciences.

4. Applications of AuNPs

4.1. Photodynamic therapy

Photodynamic therapy (PDT) is regarded as an important treatment for oncological diseases and certain skin or infectious diseases employ photosensitizers as light-sensitizing agents and a laser (the wavelength associated with a peak of dye absorption). Apoptosis or necrosis is induced in tumor cells by singlet oxygen and highly active free radicals generated via the energy of photosensitizers [79, 80].

Effective fluorescence quenching and surface plasmon resonance (SPR) absorption are the significant features of AuNPs which have been utilized in photodynamic therapy. Besides, gold conjugation facilitates intracellular penetration, due to its tendency to bind with thiols, disulfides, and amines [79].

4.2. Photothermal therapy

Photothermal therapy (PTT) known as thermal ablation [80] or optical hyperthermia [81] is a highly applied method in cancer therapy with minimal invasiveness [82-84] and it has attracted a great deal of attention in recent years [85]. AuNPs with maximum absorption in the visible or near IR region, receive light and generate heat. The heat causes the death of malignant tumors [79, 84]. Spherical solid AuNPs, with a diameter larger than 50 nm due to strong near-infrared (NIR) absorption is more common in PTT [82]. Figure 4 shows the schematic application of AuNPs in PTT and it was suggested to call this method plasmonic photothermal therapy (PPTT). Furthermore, AuNPs-antibody conjugates can be applied for both diagnosis and PPTT; the methods of what is known as theranostics. Like the PDT, binding properties of AuNPs play a key role in the process of intracellular transfer [79].

4.3. X-ray imaging

AuNPs have attracted the main attention as an x-ray contrast agent because it represents a high X-ray absorption coefficient, ease of synthetic manipulation, nontoxicity, surface functionalization for colloidal stability and targeted delivery. Common vascular contrast agents such as iodinated molecules have low-molecular weight. The water solubility of these iodinated aromatics are high and are indicative of low toxicity. Nevertheless, the blood circulation time is short and is rapidly eliminated through the kidneys. Hence, a short imaging window may require multiple injections with the risk of developing thyroid dysfunction. Au NPs have dominated the challenges [86]. The development of an imaging window due to longer vascular retention time compared with common agents is the consequence of significant properties of AuNPs [13, 86].

4.4. Drug delivery

As earlier described, considerable features of AuNPs such as unique optical, physicochemical properties, biocompatibility, functional flexibility, tunable monolayers, controlled dispersity, high surface area for loading the density of drugs, stability and nontoxicity make them an efficient nanocarrier in drug delivery systems (DDSs) [27, 31, 87]. These effective nanocarriers

are capable of transferring various drugs such as peptides [88], proteins [89, 90], plasmid DNAs (pDNAs), small interfering RNAs (siRNAs), and chemotherapeutic agents [91].

Besides the spherical nanoparticles, recent researchers have proposed stable colloidal gold nanorods as an appropriate agent for drug delivery. PEGylated Au NRs have provided a sufficient drug transfer by avoiding the reticular-endothelial system (RES) clearance. The other candidates are gold nanocages. By binding the cancer cell receptors to the nanocages surface conjugated with bioactive molecules such as antibodies, targeted drug delivery takes place. Verigene (FDA-approved) and Aurimmune (Phase-II) are gold based nanomaterials for the application of therapeutics [92].

4.5. Sensing

One of the major applications of AuNPs is in chemical and biological sensing. Utilizing the intrinsic features, Au NPs have been used as efficient sensors for the detection of different analytes such as metal ions, anions, and molecules like, saccharides, nucleotides [4], proteins [16, 93] and toxins [94]. Figure 5 shows various nanobiosensors based on AuNPs features. Regarding the intended detection and various properties of AuNPs, the sensor has been constructed. According to the sensing strategy, the AuNPs sensors can be colorimetric, fluorescence-based, electrical and electrochemical, surface plasmon resonance, surface enhanced Raman scattering (SERS)-based, quartz crystal microbalance-based and Bio-Barcode assay sensors [14]. Different types of nano-biosensors have employed special features of AuNPs.

The fundamentals of colorimetric sensing is based on visible color change due to the aggregation of AuNPs, fluorescence-based sensors: the fluorescence quenching feature of AuNPs, electrical and electrochemical sensors conductivity, high surface area and catalytic properties of AuNPs, Au NP-based surface plasmon resonance sensors: the optical properties (surface plasmon resonance, SPR) of AuNPs [16], surface-enhanced Raman scattering (SERS)-based sensors: the inelastic scattering of photons by AuNPs due to quantized vibrational level/signature, Au NPs in Quartz Crystal Microbalance-Based Sensing: the high surface area of AuNPs for enhancing detection sensitivity and Au NP-Based Bio-Barcode Assay: strong binding affinity of AuNPs to thiols and visible color change due to aggregation of AuNPs [14, 15, 95].

Among the different types of AuNPs, gold nanorods attract much attention for biosensing due to their substantial and unique optical properties. Optical absorbance and its changes are the main markers for the construction of sensitive biosensors [9].

5. Gold nanorods

Gold nanorods, as a kind of elongated AuNPs, and due to their anisotropic shape [9], possess two plasmon resonance absorptions in the visible and NIR regions, respectively, produced by transverse (TSPR), short axis, and longitudinal (LSPR), long axis, oscillation of electrons, but the spherical-shaped AuNPs have one absorption band in the visible region [10, 96, 97]. The absorption wavelength of the longitudinal bands is impressed by the aspect ratio of the nanorods and different colors are observed [10, 96]. The uptake, removal, and toxicity of Au NRs is associated with physiochemical properties such as surface groups and modifications, charge, size, shape and aspect ratio [98]. These attractable optical and unique physiochemical properties play a key role in various biomedical applications of Au NRs such as photothermal therapy, and near-infrared imaging, X-ray computed tomography, photoacoustic imaging, biosensing, drug/gene delivery, and thermal therapy of tumor [10, 96].

5.1. Synthesis of Au NRs

Well-defined size and shape synthesis of Au NRs has been considered in recent years due to the optical, electronic, physicochemical features of these nanomaterials. Various methods have been proposed for the production of Au NRs (Table 2). The first Au NRs were synthesized by the electrochemical method in 1997. Gold as the anode and platinum as the cathode were plunged in an electrolytic solution containing the cationic surfactant, hexadecyltrimethylammonium bromide (CTAB), and a cosurfactant. AuNPs were formed by turning the bulk gold metal (the anode form) into nanoparticles [10, 99].

The seeded growth synthesis of Au NRs is known as a convenient and versatile method and had been conducted between 1999 and 2004 [100]. Since then, various modifications have been performed on seeded growth syntheses [100-105], and it still has its own enthusiasts [102, 103]. Briefly, by adding the spherical ‘seed’ nanoparticles (~4 nm) to a growth solution such as gold salt, silver nitrate, ascorbic acid, and cetyltrimethylammonium bromide (CTAB), the rod like particles are formed [97].

5.2. Applications of Au NRs

Tunable surface plasmon and photothermal effects [106], the broadly accepted biocompatibility and functional versatility [107] of Au NRs are the basis of a wide range of biomedical sensing (Table 3), drug delivery [107-109], thermo-chemotherapeutic [106] and, PTT [106, 110-113] applications. Besides, these potential features, and its unparalleled nature establish promising multipurpose applications. The utilization of Au NRs as a suitable carrier, benefits their surface plasmon resonance for converting the incident light energy (NIR) into heat energy, is the foundation of major targeted drug delivery applications which combined with photothermal ablation for cancer therapy. Some multi-function applications of Au NRs include targeted cancer imaging and PTT [114, 115], cell imaging and drug delivery [116], therapeutic and imaging applications [117], drug delivery and therapeutic applications [118], drug delivery and PTT [96, 119-125], imaging and PTT [126-128], PDT and imaging [129-131], Near-Infrared Fluorescence imaging and photodynamic/photothermal therapy application [132], and PDT [133-138]. These are some multi-function application of Au NRs.

Biosensors have made a tremendous transformation in the detection of various substances especially medical diagnosis. In the meantime, AuNPs particularly Au NRs play a key role in the design of different types of nanobiosensors due to their inherent properties. Based on the transducer, piezoelectrical, optical, electrochemical and thermal/ calorimetric biosensors are designed. Among different AuNPs structures, Au NRs are appropriate candidates. The high surface area with efficient mass transport characteristics and eligible biocompatibility [139], extreme sensitive longitudinal SPR [140-142], high stability under high ionic strength conditions without addition of any stabilizing agent [143] are some significant features of Au NRs for progression of sensitive and selective biosensors.

6. Conclusions

In recent years, the conventional biomedical methods have been successfully replaced with modern nanotechnology methods for considerable accuracy, much sensitivity, efficiency and

high-speed measurement. Among the various applied types of metallic nanomaterials, AuNPs have attracted particular attention as a result of their intrinsic features. Obviously, there has been a great promotion in the synthesis, functionalization of AuNPs, which has resulted in advanced diagnostic and therapeutic procedure. Spherical and nanorods have been devoted a special place by optical, electronic, physicochemical, surface plasmon resonance (SPR) properties. Au NRs have been a suitable candidate for localized surface plasmon resonance technique. Besides, utilizing the different properties, plentiful technique assays can be designed and multipurpose applications have been attained which require more investigations.

References

- [1] Y. Zhou, C.Y. Wang, Y.R. Zhu, Z.Y. Chen, A novel ultraviolet irradiation technique for shape-controlled synthesis of gold nanoparticles at room temperature, *Chem. Mater.* 11 (1999) 2310–2312.
- [2] P. Zhao, N. Li, D. Astruc, State of the art in gold nanoparticle synthesis, *Coord. Chem. Rev.* 257 (2013) 638–665.
- [3] Z. Zhang, Z. Chen, F. Cheng, Y. Zhang, L. Chen, Highly sensitive on-site detection of glucose in human urine with naked eye based on enzymatic-like reaction mediated etching of gold nanorods, *Biosens. Bioelectron.* 89 (2017) 932–936.
- [4] Z. Zhang, J. Wang, X. Nie, T. Wen, Y. Ji, X. Wu, Y. Zhao, C. Chen, Near infrared laser-induced targeted cancer therapy using thermoresponsive polymer encapsulated gold nanorods, *J. Am. Chem. Soc.* 136 (2014) 7317–7326.
- [5] Y. Zhang, W. Chu, A.D. Foroushani, H. Wang, D. Li, J. Liu, C.J. Barrow, X. Wang, W. Yang, New gold nanostructures for sensor applications: a review, *Materials (Basel)*. 7 (2014) 5169–5201.
- [6] Y. Zhang, J. Qian, D. Wang, Y. Wang, S. He, Multifunctional gold nanorods with ultrahigh stability and tunability for in vivo fluorescence imaging, SERS detection, and photodynamic therapy, *Angew. Chemie Int. Ed.* 52 (2013) 1148–1151.
- [7] X. Zhang, Gold nanoparticles: recent advances in the biomedical applications, *Cell Biochem. Biophys.* 72 (2015) 771–775.
- [8] W. Zhang, F. Wang, Y. Wang, J. Wang, Y. Yu, S. Guo, R. Chen, D. Zhou, pH and near-infrared light dual-stimuli responsive drug delivery using DNA-conjugated gold nanorods for effective treatment of multidrug resistant cancer cells, *J. Control. Release.* 232 (2016) 9–19.
- [9] S. Zeng, K.-T. Yong, I. Roy, X.-Q. Dinh, X. Yu, F. Luan, A review on functionalized gold nanoparticles for biosensing applications, *Plasmonics*. 6 (2011) 491.
- [10] F. Yuan, H. Chen, J. Xu, Y. Zhang, Y. Wu, L. Wang, Aptamer-Based Luminescence Energy Transfer from Near-Infrared-to-Near-Infrared Upconverting Nanoparticles to Gold Nanorods and Its Application for the Detection of Thrombin, *Chem. Eur. J.* 20 (2014) 2888–2894.
- [11] Y.-Y. Yu, S.-S. Chang, C.-L. Lee, C.R.C. Wang, Gold nanorods: electrochemical synthesis and optical properties, *J. Phys. Chem. B.* 101 (1997) 6661–6664.
- [12] A.M. Youssef, M.S. Abdel-Aziz, S.M. El-Sayed, Chitosan nanocomposite films based on Ag-NP and Au-NP biosynthesis by *Bacillus Subtilis* as packaging materials, *Int. J. Biol. Macromol.* 69 (2014) 185–191.
- [13] D. Yin, X. Li, Y. Ma, Z. Liu, Targeted Cancer Imaging and Photothermal Therapy via Monosaccharide-Imprinted Gold Nanorods, *Chem. Commun.* (2017).
- [14] E.L.L. Yeo, U. Joshua, J. Cheah, D.J.H. Neo, W.I. Goh, P. Kanchanawong, K.C. Soo, P.S.P. Thong, J.C.Y. Kah, Exploiting the protein corona around gold nanorods for low-dose combined photothermal and photodynamic therapy, *J. Mater. Chem. B.* 5 (2017) 254–268.
- [15] Y.-C. Yeh, B. Creran, V.M. Rotello, Gold nanoparticles: preparation, properties, and applications in bionanotechnology, *Nanoscale*. 4 (2012) 1871–1880.
- [16] X. Ye, Y. Gao, J. Chen, D.C. Reifsnyder, C. Zheng, C.B. Murray, Seeded growth of monodisperse gold nanorods using bromide-free surfactant mixtures, *Nano Lett.* 13 (2013) 2163–2171.

- [17] Z. Yang, Z. Li, X. Lu, F. He, X. Zhu, Y. Ma, R. He, F. Gao, W. Ni, Y. Yi, Controllable Biosynthesis and Properties of Gold Nanoplates Using Yeast Extract, *Nano-Micro Lett.* 9 (2017) 5.
- [18] M. Yang, Y. Liu, W. Hou, X. Zhi, C. Zhang, X. Jiang, F. Pan, Y. Yang, J. Ni, D. Cui, Mitomycin C-treated human-induced pluripotent stem cells as a safe delivery system of gold nanorods for targeted photothermal therapy of gastric cancer, *Nanoscale.* 9 (2017) 334–340.
- [19] E. Yan, M. Cao, Y. Wang, X. Hao, S. Pei, J. Gao, Y. Wang, Z. Zhang, D. Zhang, Gold nanorods contained polyvinyl alcohol/chitosan nanofiber matrix for cell imaging and drug delivery, *Mater. Sci. Eng. C.* 58 (2016) 1090–1097.
- [20] G. Yamal, P. Sharmila, K.S. Rao, P. Pardha-Saradhi, Yeast Extract Mannitol medium and its constituents promote synthesis of Au nanoparticles, *Process Biochem.* 48 (2013) 532–538.
- [21] X. Xu, X. Liu, Y. Li, Y. Ying, A simple and rapid optical biosensor for detection of aflatoxin B1 based on competitive dispersion of gold nanorods, *Biosens. Bioelectron.* 47 (2013) 361–367.
- [22] Y. Xiao, H. Hong, V.Z. Matson, A. Javadi, W. Xu, Y. Yang, Y. Zhang, J.W. Engle, R.J. Nickles, W. Cai, Gold nanorods conjugated with doxorubicin and cRGD for combined anticancer drug delivery and PET imaging, *Theranostics.* 2 (2012) 757.
- [23] H.-L. Wu, H.-R. Tsai, Y.-T. Hung, K.-U. Lao, C.-W. Liao, P.-J. Chung, J.-S. Huang, I.-C. Chen, M.H. Huang, A comparative study of gold nanocubes, octahedra, and rhombic dodecahedra as highly sensitive SERS substrates, *Inorg. Chem.* 50 (2011) 8106–8111.
- [24] H.-Y. Wu, H.-C. Chu, T.-J. Kuo, C.-L. Kuo, M.H. Huang, Seed-mediated synthesis of high aspect ratio gold nanorods with nitric acid, *Chem. Mater.* 17 (2005) 6447–6451.
- [25] W. Wen, J.-Y. Huang, T. Bao, J. Zhou, H.-X. Xia, X.-H. Zhang, S.-F. Wang, Y.-D. Zhao, Increased electrocatalyzed performance through hairpin oligonucleotide aptamer-functionalized gold nanorods labels and graphene-streptavidin nanomatrix: Highly selective and sensitive electrochemical biosensor of carcinoembryonic antigen, *Biosens. Bioelectron.* 83 (2016) 142–148.
- [26] L. WeiáYap, Plasmonic caged gold nanorods for near-infrared light controlled drug delivery, *Nanoscale.* 6 (2014) 14388–14393.
- [27] Y. Wang, K.C.L. Black, H. Luehmann, W. Li, Y. Zhang, X. Cai, D. Wan, S.-Y. Liu, M. Li, P. Kim, Comparison study of gold nanohexapods, nanorods, and nanocages for photothermal cancer treatment, *ACS Nano.* 7 (2013) 2068–2077.
- [28] X. Wang, Y. Li, H. Wang, Q. Fu, J. Peng, Y. Wang, J. Du, Y. Zhou, L. Zhan, Gold nanorod-based localized surface plasmon resonance biosensor for sensitive detection of hepatitis B virus in buffer, blood serum and plasma, *Biosens. Bioelectron.* 26 (2010) 404–410.
- [29] S. Wang, X. Zhao, S. Wang, J. Qian, S. He, Biologically Inspired Polydopamine Capped Gold Nanorods for Drug Delivery and Light-Mediated Cancer Therapy, *ACS Appl. Mater. Interfaces.* 8 (2016) 24368–24384.
- [30] L. Wang, Y. Liu, W. Li, X. Jiang, Y. Ji, X. Wu, L. Xu, Y. Qiu, K. Zhao, T. Wei, Selective targeting of gold nanorods at the mitochondria of cancer cells: implications for cancer therapy, *Nano Lett.* 11 (2010) 772–780.
- [31] J. Wang, G. Zhu, M. You, E. Song, M.I. Shukoor, K. Zhang, M.B. Altman, Y. Chen, Z. Zhu, C.Z. Huang, Assembly of aptamer switch probes and photosensitizer on gold nanorods for targeted photothermal and photodynamic cancer therapy, *ACS Nano.* 6 (2012) 5070–5077.
- [32] B. Wang, J.-H. Wang, Q. Liu, H. Huang, M. Chen, K. Li, C. Li, X.-F. Yu, P.K. Chu, Rose-bengal-conjugated gold nanorods for in vivo photodynamic and photothermal oral cancer therapies, *Biomaterials.* 35 (2014) 1954–1966.
- [33] S.A. Wadhwani, U.U. Shedbalkar, R. Singh, M.S. Karve, B.A. Chopade, Novel polyhedral gold nanoparticles: green synthesis, optimization and characterization by environmental isolate of *Acinetobacter* sp. SW30, *World J. Microbiol. Biotechnol.* 30 (2014) 2723–2731.
- [34] L. Vigdeman, B.P. Khanal, E.R. Zubarev, Functional gold nanorods: synthesis, self-assembly, and sensing applications, *Adv. Mater.* 24 (2012) 4811–4841.

- [35] R. Venkatesan, A. Pichaimani, K. Hari, P.K. Balasubramanian, J. Kulandaivel, K. Premkumar, Doxorubicin conjugated gold nanorods: a sustained drug delivery carrier for improved anticancer therapy, *J. Mater. Chem. B*. 1 (2013) 1010–1018.
- [36] A.K. Vala, Exploration on green synthesis of gold nanoparticles by a marine-derived fungus *Aspergillus sydowii*, *Environ. Prog. Sustain. Energy*. 34 (2015) 194–197.
- [37] J. Turkevich, G. Garton, P.C. Stevenson, The color of colloidal gold, *J. Colloid Sci.* 9 (1954) 26–35.
- [38] S. Thatai, P. Khurana, S. Prasad, D. Kumar, A new way in nanosensors: gold nanorods for sensing of Fe (III) ions in aqueous media, *Microchem. J.* 113 (2014) 77–82.
- [39] A.S. Thakor, J. Jokerst, C. Zavaleta, T.F. Massoud, S.S. Gambhir, Gold nanoparticles: a revival in precious metal administration to patients, *Nano Lett.* 11 (2011) 4029–4036.
- [40] K.N. Thakkar, S.S. Mhatre, R.Y. Parikh, Biological synthesis of metallic nanoparticles, *Nanomedicine Nanotechnology, Biol. Med.* 6 (2010) 257–262.
- [41] G. Terentyuk, E. Panfilova, V. Khanadeev, D. Chumakov, E. Genina, A. Bashkatov, V. Tuchin, A. Bucharskaya, G. Maslyakova, N. Khlebtsov, Gold nanorods with a hematoporphyrin-loaded silica shell for dual-modality photodynamic and photothermal treatment of tumors in vivo, *Nano Res.* 7 (2014) 325–337.
- [42] S. Taranejoo, M. Janmaleki, M. Rafienia, M. Kamali, M. Mansouri, Chitosan microparticles loaded with exotoxin A subunit antigen for intranasal vaccination against *Pseudomonas aeruginosa*: an in vitro study, *Carbohydr. Polym.* 83 (2011) 1854–1861.
- [43] R.A. Taheri, A.H. Rezayan, F. Rahimi, J. Mohammadnejad, M. Kamali, Development of an immunosensor using oriented immobilized anti-OmpW for sensitive detection of *Vibrio cholerae* by surface plasmon resonance, *Biosens. Bioelectron.* 86 (2016) 484–488.
- [44] B. Sun, J. Wu, S. Cui, H. Zhu, W. An, Q. Fu, C. Shao, A. Yao, B. Chen, D. Shi, In situ synthesis of graphene oxide/gold nanorods theranostic hybrids for efficient tumor computed tomography imaging and photothermal therapy, *Nano Res.* 10 (2017) 37–48.
- [45] T.Y. Suman, S.R.R. Rajasree, R. Ramkumar, C. Rajthilak, P. Perumal, The Green synthesis of gold nanoparticles using an aqueous root extract of *Morinda citrifolia* L, *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* 118 (2014) 11–16.
- [46] K.S.U. Suganya, K. Govindaraju, V.G. Kumar, T.S. Dhas, V. Karthick, G. Singaravelu, M. Elanchezhian, Blue green alga mediated synthesis of gold nanoparticles and its antibacterial efficacy against Gram positive organisms, *Mater. Sci. Eng. C*. 47 (2015) 351–356.
- [47] J. Stone, S. Jackson, D. Wright, Biological applications of gold nanorods, *Wiley Interdiscip. Rev. Nanomedicine Nanobiotechnology*. 3 (2011) 100–109.
- [48] J.W. Stone, A.M. Alkilany, M.A. Hamaly, S. Canonico-May, Biomedical Applications of Anisotropic Gold Nanoparticles, in: *Anisotropic Shape-Selective Nanomater.*, Springer, 2017: pp. 399–426.
- [49] N. Stasyuk, G. Gayda, R. Serkiz, M. Gonchar, The green” synthesis of gold nanoparticles by the yeast *Hansenula polymorpha*, *Вісник Львівського Університету. Серія Біологічна*. (2016) 96–102.
- [50] S.K. Srivastava, R. Yamada, C. Ogino, A. Kondo, Biogenic synthesis and characterization of gold nanoparticles by *Escherichia coli* K12 and its heterogeneous catalysis in degradation of 4-nitrophenol, *Nanoscale Res. Lett.* 8 (2013) 70.
- [51] T.-T. Song, W. Wang, L.-L. Meng, Y. Liu, X.-B. Jia, X. Mao, Electrochemical detection of human ferritin based on gold nanorod reporter probe and cotton thread immunoassay device, *Chinese Chem. Lett.* 28 (2017) 226–230.
- [52] P. Singh, Y.J. Kim, C. Wang, R. Mathiyalagan, D.C. Yang, The development of a green approach for the biosynthesis of silver and gold nanoparticles by using *Panax ginseng* root extract, and their biological applications, *Artif. Cells, Nanomedicine, Biotechnol.* 44 (2016) 1150–1157.

- [53] M. Singh, D.C.C. Harris-Birtill, Y. Zhou, M.E. Gallina, A.E.G. Cass, G.B. Hanna, D.S. Elson, Application of gold nanorods for photothermal therapy in Ex Vivo human oesophagogastric adenocarcinoma, *J. Biomed. Nanotechnol.* 12 (2016) 481–490.
- [54] C. Singh, V. Sharma, P.K.R. Naik, V. KHandelwal, H. Singh, A green biogenic approach for synthesis of gold and silver nanoparticles using *Zingiber officinale*, *Dig. J. Nanomater. Biostructures.* 6 (2011) 535–542.
- [55] S.-H. Seo, B.-M. Kim, A. Joe, H.-W. Han, X. Chen, Z. Cheng, E.-S. Jang, NIR-light-induced surface-enhanced Raman scattering for detection and photothermal/photodynamic therapy of cancer cells using methylene blue-embedded gold nanorod@ SiO₂ nanocomposites, *Biomaterials.* 35 (2014) 3309–3318.
- [56] M. Sengani, A.M. Grumezescu, V.D. Rajeswari, Recent trends and methodologies in gold nanoparticle synthesis-a prospective review on drug delivery aspect, *OpenNano.* (2017).
- [57] S. Senapati, A. Syed, S. Moez, A. Kumar, A. Ahmad, Intracellular synthesis of gold nanoparticles using alga *Tetraselmis kochinensis*, *Mater. Lett.* 79 (2012) 116–118.
- [58] G. Sathishkumar, P.K. Jha, V. Vignesh, C. Rajkuberan, M. Jeyaraj, M. Selvakumar, R. Jha, S. Sivaramakrishnan, Cannonball fruit (*Couroupita guianensis*, Aubl.) extract mediated synthesis of gold nanoparticles and evaluation of its antioxidant activity, *J. Mol. Liq.* 215 (2016) 229–236.
- [59] A. Sasidharan, N.A. Monteiro-Riviere, Biomedical applications of gold nanomaterials: opportunities and challenges, *Wiley Interdiscip. Rev. Nanomedicine Nanobiotechnology.* 7 (2015) 779–796.
- [60] G.R. Salunke, S. Ghosh, R.J.S. Kumar, S. Khade, P. Vashisth, T. Kale, S. Chopade, V. Pruthi, G. Kundu, J.R. Bellare, Rapid efficient synthesis and characterization of silver, gold, and bimetallic nanoparticles from the medicinal plant *Plumbago zeylanica* and their application in biofilm control, *Int. J. Nanomedicine.* 9 (2014) 2635.
- [61] K. Saha, S.S. Agasti, C. Kim, X. Li, V.M. Rotello, Gold nanoparticles in chemical and biological sensing, *Chem. Rev.* 112 (2012) 2739–2779.
- [62] R.S. Riley, E.S. Day, Gold nanoparticle-mediated photothermal therapy: applications and opportunities for multimodal cancer treatment, *Wiley Interdiscip. Rev. Nanomedicine Nanobiotechnology.* (2017).
- [63] S. Rana, A. Bajaj, R. Mout, V.M. Rotello, Monolayer coated gold nanoparticles for delivery applications, *Adv. Drug Deliv. Rev.* 64 (2012) 200–216.
- [64] M. Ramakrishna, D.R. Babu, R.M. Gengan, S. Chandra, G.N. Rao, Green synthesis of gold nanoparticles using marine algae and evaluation of their catalytic activity, *J. Nanostructure Chem.* 6 (2016) 1–13.
- [65] F.A.A. Rajathi, C. Parthiban, V.G. Kumar, P. Anantharaman, Biosynthesis of antibacterial gold nanoparticles using brown alga, *Stoechospermum marginatum* (kützing), *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* 99 (2012) 166–173.
- [66] P. Rajasulochana, R. Dhamotharan, P. Murugakoothan, S. Murugesan, P. Krishnamoorthy, Biosynthesis and characterization of gold nanoparticles using the alga *Kappaphycus alvarezii*, *Int. J. Nanosci.* 9 (2010) 511–516.
- [67] Z. Qian, S.-J. Park, Silver seeds and aromatic surfactants facilitate the growth of anisotropic metal nanoparticles: gold triangular nanoprisms and ultrathin nanowires, *Chem. Mater.* 26 (2014) 6172–6177.
- [68] E. Priyadarshini, N. Pradhan, Gold nanoparticles as efficient sensors in colorimetric detection of toxic metal ions: a review, *Sensors Actuators B Chem.* 238 (2017) 888–902.
- [69] P. Prema, P.A. Iniya, G. Immanuel, Microbial mediated synthesis, characterization, antibacterial and synergistic effect of gold nanoparticles using *Klebsiella pneumoniae* (MTCC-4030), *RSC Adv.* 6 (2016) 4601–4607.
- [70] A. Pitchaimani, T.D.T. Nguyen, L. Maurmann, J. Key, S.H. Bossmann, S. Aryal, Gd³⁺ Tethered Gold Nanorods for Combined Magnetic Resonance Imaging and Photo-Thermal Therapy, *J. Biomed. Nanotechnol.* 13 (2017) 417–426.

- [71] D. Pissuwan, T. Niidome, M.B. Cortie, The forthcoming applications of gold nanoparticles in drug and gene delivery systems, *J. Control. Release.* 149 (2011) 65–71.
- [72] D. Philip, Green synthesis of gold and silver nanoparticles using *Hibiscus rosa sinensis*, *Phys. E Low-Dimensional Syst. Nanostructures.* 42 (2010) 1417–1424.
- [73] M. Pérez-Ortiz, C. Zapata-Urzúa, G.A. Acosta, A. Álvarez-Lueje, F. Albericio, M.J. Kogan, Gold nanoparticles as an efficient drug delivery system for GLP-1 peptides, *Colloids Surfaces B Biointerfaces.* 158 (2017) 25–32.
- [74] K. Park, M. Hsiao, Y.-J. Yi, S. Izor, H. Koerner, A. Jawaid, R.A. Vaia, Highly Concentrated Seed-Mediated Synthesis of Monodispersed Gold Nanorods, *ACS Appl. Mater. Interfaces.* 9 (2017) 26363–26371.
- [75] K. Park, L.F. Drummy, R.C. Wadams, H. Koerner, D. Nepal, L. Fabris, R.A. Vaia, Growth mechanism of gold nanorods, *Chem. Mater.* 25 (2013) 555–563.
- [76] S. Parida, C. Maiti, Y. Rajesh, K.K. Dey, I. Pal, A. Parekh, R. Patra, D. Dhara, P.K. Dutta, M. Mandal, Gold nanorod embedded reduction responsive block copolymer micelle-triggered drug delivery combined with photothermal ablation for targeted cancer therapy, *Biochim. Biophys. Acta (BBA)-General Subj.* 1861 (2017) 3039–3052.
- [77] H.J. Parab, C. Jung, J.-H. Lee, H.G. Park, A gold nanorod-based optical DNA biosensor for the diagnosis of pathogens, *Biosens. Bioelectron.* 26 (2010) 667–673.
- [78] M. Noruzi, D. Zare, K. Khoshnevisan, D. Davoodi, Rapid green synthesis of gold nanoparticles using *Rosa hybrida* petal extract at room temperature, *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* 79 (2011) 1461–1465.
- [79] J. Narang, N. Malhotra, G. Singh, C.S. Pundir, Electrochemical impedimetric detection of anti-HIV drug taking gold nanorods as a sensing interface, *Biosens. Bioelectron.* 66 (2015) 332–337.
- [80] C.J. Murphy, L.B. Thompson, A.M. Alkilany, P.N. Sisco, S.P. Boulos, S.T. Sivapalan, J.A. Yang, D.J. Chernak, J. Huang, The many faces of gold nanorods, *J. Phys. Chem. Lett.* 1 (2010) 2867–2875.
- [81] D. MubarakAli, N. Thajuddin, K. Jeganathan, M. Gunasekaran, Plant extract mediated synthesis of silver and gold nanoparticles and its antibacterial activity against clinically isolated pathogens, *Colloids Surfaces B Biointerfaces.* 85 (2011) 360–365.
- [82] P. Mishra, S. Ray, S. Sinha, B. Das, M.I. Khan, S.K. Behera, S.-I. Yun, S.K. Tripathy, A. Mishra, Facile bio-synthesis of gold nanoparticles by using extract of *Hibiscus sabdariffa* and evaluation of its cytotoxicity against U87 glioblastoma cells under hyperglycemic condition, *Biochem. Eng. J.* 105 (2016) 264–272.
- [83] V.S. Marangoni, J. Cancino-Bernardi, V. Zucolotto, Synthesis, Physico-Chemical Properties, and Biomedical Applications of Gold Nanorods—A Review, *J. Biomed. Nanotechnol.* 12 (2016) 1136–1158.
- [84] H.M. Manjunath, C.G. Joshi, N.G. Raju, Biofabrication of gold nanoparticles using marine endophytic fungus—*Penicillium citrinum*, *IET Nanobiotechnology.* 11 (2016) 40–44.
- [85] C. Malarkodi, S. Rajeshkumar, M. Vanaja, K. Paulkumar, G. Gnanajobitha, G. Annadurai, Eco-friendly synthesis and characterization of gold nanoparticles using *Klebsiella pneumoniae*, *J. Nanostructure Chem.* 3 (2013) 30.
- [86] M.A. Mackey, M.R.K. Ali, L.A. Austin, R.D. Near, M.A. El-Sayed, The most effective gold nanorod size for plasmonic photothermal therapy: theory and in vitro experiments, *J. Phys. Chem. B.* 118 (2014) 1319–1326.
- [87] P. Luo, Y. Liu, Y. Xia, H. Xu, G. Xie, Aptamer biosensor for sensitive detection of toxin A of *Clostridium difficile* using gold nanoparticles synthesized by *Bacillus stearothermophilus*, *Biosens. Bioelectron.* 54 (2014) 217–221.
- [88] X. Lu, X. Dong, K. Zhang, X. Han, X. Fang, Y. Zhang, A gold nanorods-based fluorescent biosensor for the detection of hepatitis B virus DNA based on fluorescence resonance energy transfer, *Analyst.* 138 (2013) 642–650.

- [89] A.J. Love, V. V Makarov, O. V Sinitsyna, J. Shaw, I. V Yaminsky, N.O. Kalinina, M.E. Taliansky, A Genetically Modified Tobacco Mosaic Virus that can Produce Gold Nanoparticles from a Metal Salt Precursor, *Front. Plant Sci.* 6 (2015).
- [90] S.E. Lohse, C.J. Murphy, The quest for shape control: a history of gold nanorod synthesis, *Chem. Mater.* 25 (2013) 1250–1261.
- [91] J. Liu, C. Detrembleur, D. Pauw-Gillet, S. Mornet, C. Jérôme, E. Duguet, Gold Nanorods Coated with Mesoporous Silica Shell as Drug Delivery System for Remote Near Infrared Light-Activated Release and Potential Phototherapy, *Small.* 11 (2015) 2323–2332.
- [92] H. Liu, T. Lian, Y. Liu, Y. Hong, D. Sun, Q. Li, Plant-mediated synthesis of Au nanoparticles: Separation and identification of active biomolecule in the water extract of *Cacumen Platycladi*, *Ind. Eng. Chem. Res.* 56 (2017) 5262–5270.
- [93] J. Lin, S. Wang, P. Huang, Z. Wang, S. Chen, G. Niu, W. Li, J. He, D. Cui, G. Lu, Photosensitizer-loaded gold vesicles with strong plasmonic coupling effect for imaging-guided photothermal/photodynamic therapy, *ACS Nano.* 7 (2013) 5320–5329.
- [94] D.-Z. Lin, P.-C. Chuang, P.-C. Liao, J.-P. Chen, Y.-F. Chen, Increasing the spectral shifts in LSPR biosensing using DNA-functionalized gold nanorods in a competitive assay format for the detection of interferon- γ , *Biosens. Bioelectron.* 81 (2016) 221–228.
- [95] Y. Li, Y. Zhang, M. Zhao, Q. Zhou, L. Wang, H. Wang, X. Wang, L. Zhan, A simple aptamer-functionalized gold nanorods based biosensor for the sensitive detection of MCF-7 breast cancer cells, *Chem. Commun.* 52 (2016) 3959–3961.
- [96] N. Li, D. Niu, X. Jia, J. He, Y. Jiang, J. Gu, Z. Li, S. Xu, Y. Li, Multiple gold nanorods@hierarchically porous silica nanospheres for efficient multi-drug delivery and photothermal therapy, *J. Mater. Chem. B.* 5 (2017) 1642–1649.
- [97] N. Li, P. Zhao, D. Astruc, Anisotropic gold nanoparticles: synthesis, properties, applications, and toxicity, *Angew. Chemie Int. Ed.* 53 (2014) 1756–1789.
- [98] M.R. Langille, M.L. Personick, J. Zhang, C.A. Mirkin, Defining rules for the shape evolution of gold nanoparticles, *J. Am. Chem. Soc.* 134 (2012) 14542–14554.
- [99] W. Kuo, C. Chang, Y. Chang, M. Yang, Y. Chien, S. Chen, C. Yeh, Gold nanorods in photodynamic therapy, as hyperthermia agents, and in near-infrared optical imaging, *Angew. Chemie.* 122 (2010) 2771–2775.
- [100] V.G. Kumar, S.D. Gokavarapu, A. Rajeswari, T.S. Dhas, V. Karthick, Z. Kapadia, T. Shrestha, I.A. Barathy, A. Roy, S. Sinha, Facile green synthesis of gold nanoparticles using leaf extract of antidiabetic potent *Cassia auriculata*, *Colloids Surfaces B Biointerfaces.* 87 (2011) 159–163.
- [101] A. Kumar, X. Zhang, X.-J. Liang, Gold nanoparticles: emerging paradigm for targeted drug delivery system, *Biotechnol. Adv.* 31 (2013) 593–606.
- [102] M. Kitching, P. Choudhary, S. Inguva, Y. Guo, M. Ramani, S.K. Das, E. Marsili, Fungal surface protein mediated one-pot synthesis of stable and hemocompatible gold nanoparticles, *Enzyme Microb. Technol.* 95 (2016) 76–84.
- [103] F. Kim, J.H. Song, P. Yang, Photochemical synthesis of gold nanorods, *J. Am. Chem. Soc.* 124 (2002) 14316–14317.
- [104] E.J. Kim, E.B. Kim, S.W. Lee, S.A. Cheon, H.-J. Kim, J. Lee, M.-K. Lee, S. Ko, T.J. Park, An easy and sensitive sandwich assay for detection of *Mycobacterium tuberculosis* Ag85B antigen using quantum dots and gold nanorods, *Biosens. Bioelectron.* 87 (2017) 150–156.
- [105] M.S. Khan, G.D. Vishakante, H. Siddaramaiah, Gold nanoparticles: a paradigm shift in biomedical applications, *Adv. Colloid Interface Sci.* 199 (2013) 44–58.
- [106] A.K. Khan, R. Rashid, G. Murtaza, A. Zahra, Gold nanoparticles: synthesis and applications in drug delivery, *Trop. J. Pharm. Res.* 13 (2014) 1169–1177.
- [107] E.H. Jeong, G. Jung, C. Am Hong, H. Lee, Gold nanoparticle (AuNP)-based drug delivery and molecular imaging for biomedical applications, *Arch. Pharm. Res.* 37 (2014) 53–59.

- [108] C. Jayaseelan, R. Ramkumar, A.A. Rahuman, P. Perumal, Green synthesis of gold nanoparticles using seed aqueous extract of *Abelmoschus esculentus* and its antifungal activity, *Ind. Crops Prod.* 45 (2013) 423–429.
- [109] B. Jang, J.-Y. Park, C.-H. Tung, I.-H. Kim, Y. Choi, Gold nanorod– photosensitizer complex for near-infrared fluorescence imaging and photodynamic/photothermal therapy in vivo, *ACS Nano.* 5 (2011) 1086–1094.
- [110] S. Hwang, J. Nam, S. Jung, J. Song, H. Doh, S. Kim, Gold nanoparticle-mediated photothermal therapy: current status and future perspective, *Nanomedicine.* 9 (2014) 2003–2022.
- [111] N.I. Hulkoti, T.C. Taranath, Biosynthesis of nanoparticles using microbes—a review, *Colloids Surfaces B Biointerfaces.* 121 (2014) 474–483.
- [112] T. Huang, F. Meng, L. Qi, Controlled synthesis of dendritic gold nanostructures assisted by supramolecular complexes of surfactant with cyclodextrin, *Langmuir.* 26 (2009) 7582–7589.
- [113] P. Huang, L. Bao, C. Zhang, J. Lin, T. Luo, D. Yang, M. He, Z. Li, G. Gao, B. Gao, Folic acid-conjugated silica-modified gold nanorods for X-ray/CT imaging-guided dual-mode radiation and photo-thermal therapy, *Biomaterials.* 32 (2011) 9796–9809.
- [114] H. Huang, W. Bai, C. Dong, R. Guo, Z. Liu, An ultrasensitive electrochemical DNA biosensor based on graphene/Au nanorod/polythionine for human papillomavirus DNA detection, *Biosens. Bioelectron.* 68 (2015) 442–446.
- [115] H. Huang, X. Liu, T. Hu, P.K. Chu, Ultra-sensitive detection of cysteine by gold nanorod assembly, *Biosens. Bioelectron.* 25 (2010) 2078–2083.
- [116] S. Her, D.A. Jaffray, C. Allen, Gold nanoparticles for applications in cancer radiotherapy: Mechanisms and recent advancements, *Adv. Drug Deliv. Rev.* 109 (2017) 84–101.
- [117] Y. He, J. Tian, J. Zhang, S. Chen, Y. Jiang, K. Hu, Y. Zhao, S. Zhao, DNAzyme self-assembled gold nanorods-based FRET or polarization assay for ultrasensitive and selective detection of copper (II) ion, *Biosens. Bioelectron.* 55 (2014) 285–288.
- [118] X. Han, X. Fang, A. Shi, J. Wang, Y. Zhang, An electrochemical DNA biosensor based on gold nanorods decorated graphene oxide sheets for sensing platform, *Anal. Biochem.* 443 (2013) 117–123.
- [119] J.F. Hainfeld, H.M. Smilowitz, M.J. O'Connor, F.A. Dilmanian, D.N. Slatkin, Gold nanoparticle imaging and radiotherapy of brain tumors in mice, *Nanomedicine.* 8 (2013) 1601–1609.
- [120] M. Guo, W. Li, F. Yang, H. Liu, Controllable biosynthesis of gold nanoparticles from a *Eucommia ulmoides* bark aqueous extract, *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* 142 (2015) 73–79.
- [121] M. Grzelczak, A. Sánchez-Iglesias, H.H. Mezerji, S. Bals, J. Pérez-Juste, L.M. Liz-Marzán, Steric hindrance induces crosslike self-assembly of gold nanodumbbells, *Nano Lett.* 12 (2012) 4380–4384.
- [122] K. Gopinath, K.S. Venkatesh, R. Ilangoan, K. Sankaranarayanan, A. Arumugam, Green synthesis of gold nanoparticles from leaf extract of *Terminalia arjuna*, for the enhanced mitotic cell division and pollen germination activity, *Ind. Crops Prod.* 50 (2013) 737–742.
- [123] G.P. Goodrich, L. Bao, K. Gill-Sharp, K.L. Sang, J. Wang, J.D. Payne, Photothermal therapy in a murine colon cancer model using near-infrared absorbing gold nanorods, *J. Biomed. Opt.* 15 (2010) 18001–18008.
- [124] N. Gonzalez-Ballesteros, S. Prado-López, J.B. Rodriguez-Gonzalez, M. Lastra, M.C. Rodríguez-Argüelles, Green synthesis of gold nanoparticles using brown algae *Cystoseira baccata*: Its activity in colon cancer cells, *Colloids Surfaces B Biointerfaces.* 153 (2017) 190–198.
- [125] C. Gonnelli, C. Giordano, U. Fontani, M.C. Salvatici, S. Ristori, Green Synthesis of Gold Nanoparticles from Extracts of *Cucurbita pepo* L. Leaves: Insights on the Role of Plant Ageing, in: *Adv. Bionanomaterials*, Springer, 2018: pp. 155–164.
- [126] P. Ghosh, G. Han, M. De, C.K. Kim, V.M. Rotello, Gold nanoparticles in delivery applications, *Adv. Drug Deliv. Rev.* 60 (2008) 1307–1315.

- [127] X. Fu, X. Fu, Q. Wang, L. Sheng, X. Huang, M. Ma, Z. Cai, Fluorescence switch biosensor based on quantum dots and gold nanoparticles for discriminative detection of lysozyme, *Int. J. Biol. Macromol.* (2017).
- [128] C. Fernández-López, L. Polavarapu, D.M. Solís, J.M. Taboada, F. Obelleiro, R. Contreras-Cáceres, I. Pastoriza-Santos, J. Pérez-Juste, Gold nanorod–pnipam hybrids with reversible plasmon coupling: Synthesis, modeling, and sers properties, *ACS Appl. Mater. Interfaces*. 7 (2015) 12530–12538.
- [129] N.-S. Eum, S.-H. Yeom, D.-H. Kwon, H.-R. Kim, S.-W. Kang, Enhancement of sensitivity using gold nanorods—Antibody conjugator for detection of *E. coli* O157: H7, *Sensors Actuators B Chem.* 143 (2010) 784–788.
- [130] L. Dykman, N. Khlebtsov, Gold nanoparticles in biomedical applications: recent advances and perspectives, *Chem. Soc. Rev.* 41 (2012) 2256–2282.
- [131] B. Duncan, C. Kim, V.M. Rotello, Gold nanoparticle platforms as drug and biomacromolecule delivery systems, *J. Control. Release*. 148 (2010) 122–127.
- [132] C. Du, A. Wang, J. Fei, J. Zhao, J. Li, Polypyrrole-stabilized gold nanorods with enhanced photothermal effect towards two-photon photothermal therapy, *J. Mater. Chem. B*. 3 (2015) 4539–4545.
- [133] E.C. Dreaden, A.M. Alkilany, X. Huang, C.J. Murphy, M.A. El-Sayed, The golden age: gold nanoparticles for biomedicine, *Chem. Soc. Rev.* 41 (2012) 2740–2779.
- [134] T.S. Dhas, V.G. Kumar, V. Karthick, K. Govindaraju, T.S. Narayana, Biosynthesis of gold nanoparticles using *Sargassum swartzii* and its cytotoxicity effect on HeLa cells, *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* 133 (2014) 102–106.
- [135] N.N. Dhanasekar, G.R. Rahul, K.B. Narayanan, G. Raman, N. Sakthivel, Green chemistry approach for the synthesis of gold nanoparticles using the fungus *Alternaria* sp, *J Microbiol Biotechnol.* 25 (2015) 1129–1135.
- [136] S.K. Das, A.R. Das, A.K. Guha, Microbial synthesis of multishaped gold nanostructures, *Small*. 6 (2010) 1012–1021.
- [137] M.-C. Daniel, D. Astruc, Gold nanoparticles: assembly, supramolecular chemistry, quantum-size-related properties, and applications toward biology, catalysis, and nanotechnology, *Chem. Rev.* 104 (2004) 293–346.
- [138] D.N. Correa-Llantén, S.A. Muñoz-Ibacache, M.E. Castro, P.A. Muñoz, J.M. Blamey, Gold nanoparticles synthesized by *Geobacillus* sp. strain ID17 a thermophilic bacterium isolated from Deception Island, Antarctica, *Microb. Cell Fact.* 12 (2013) 75.
- [139] L.E. Cole, R.D. Ross, J.M.R. Tilley, T. Vargo-Gogola, R.K. Roeder, Gold nanoparticles as contrast agents in x-ray imaging and computed tomography, *Nanomedicine*. 10 (2015) 321–341.
- [140] W. Il Choi, A. Sahu, Y.H. Kim, G. Tae, Photothermal cancer therapy and imaging based on gold nanorods, *Ann. Biomed. Eng.* 40 (2012) 534–546.
- [141] X. Cheng, R. Sun, L. Yin, Z. Chai, H. Shi, M. Gao, Light-Triggered Assembly of Gold Nanoparticles for Photothermal Therapy and Photoacoustic Imaging of Tumors In Vivo, *Adv. Mater.* 29 (2017).
- [142] K. Cheng, J. Zhang, L. Zhang, L. Wang, H. Chen, Aptamer biosensor for *Salmonella typhimurium* detection based on luminescence energy transfer from Mn²⁺-doped NaYF₄: Yb, Tm upconverting nanoparticles to gold nanorods, *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* 171 (2017) 168–173.
- [143] E. Castro-Longoria, A.R. Vilchis-Nestor, M. Avalos-Borja, Biosynthesis of silver, gold and bimetallic nanoparticles using the filamentous fungus *Neurospora crassa*, *Colloids Surfaces B Biointerfaces*. 83 (2011) 42–48.
- [144] E.T. Castellana, R.C. Gamez, D.H. Russell, Label-free biosensing with lipid-functionalized gold nanorods, *J. Am. Chem. Soc.* 133 (2011) 4182–4185.
- [145] J. Cao, T. Sun, K.T. V Grattan, Gold nanorod-based localized surface plasmon resonance biosensors: A review, *Sensors Actuators B Chem.* 195 (2014) 332–351.

- [146] H.-H. Cai, D. Lin, J. Wang, P.-H. Yang, J. Cai, Controlled side-by-side assembly of gold nanorods: A strategy for lead detection, *Sensors Actuators B Chem.* 196 (2014) 252–259.
- [147] M. Brust, M. Walker, D. Bethell, D.J. Schiffrin, R. Whyman, Synthesis of thiol-derivatised gold nanoparticles in a two-phase liquid–liquid system, *J. Chem. Soc. Chem. Commun.* (1994) 801–802.
- [148] S. Bhana, R. O'Connor, J. Johnson, J.D. Ziebarth, L. Henderson, X. Huang, Photosensitizer-loaded gold nanorods for near infrared photodynamic and photothermal cancer therapy, *J. Colloid Interface Sci.* 469 (2016) 8–16.
- [149] S. Barbosa, A. Agrawal, L. Rodríguez-Lorenzo, I. Pastoriza-Santos, R.A. Alvarez-Puebla, A. Kornowski, H. Weller, L.M. Liz-Marzán, Tuning size and sensing properties in colloidal gold nanostars, *Langmuir.* 26 (2010) 14943–14950.
- [150] S.K. Balasubramanian, L. Yang, L.-Y.L. Yung, C.-N. Ong, W.-Y. Ong, E.Y. Liya, Characterization, purification, and stability of gold nanoparticles, *Biomaterials.* 31 (2010) 9023–9030.
- [151] S. Bagdeli, A.H. Rezayan, R.A. Taheri, M. Kamali, M. Hosseini, FRET-based immunoassay using CdTe and AuNPs for the detection of OmpW antigen of *Vibrio cholerae*, *J. Lumin.* (2017).
- [152] Y.A. Attia, Y.E. Farag, Y.M.A. Mohamed, A.T. Hussien, T. Youssef, Photo-extracellular synthesis of gold nanoparticles using Baker's yeast and their anticancer evaluation against Ehrlich ascites carcinoma cells, *New J. Chem.* 40 (2016) 9395–9402.
- [153] P. Arunkumar, B. Raju, R. Vasantharaja, S. Vijayaraghavan, B.P. Kumar, K. Jeganathan, K. Premkumar, Near infra-red laser mediated photothermal and antitumor efficacy of doxorubicin conjugated gold nanorods with reduced cardiotoxicity in swiss albino mice, *Nanomedicine Nanotechnology, Biol. Med.* 11 (2015) 1435–1444.
- [154] S.A. Aromal, D. Philip, Green synthesis of gold nanoparticles using *Trigonella foenum-graecum* and its size-dependent catalytic activity, *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* 97 (2012) 1–5.
- [155] A. Annamalai, V.L.P. Christina, D. Sudha, M. Kalpana, P.T. V Lakshmi, Green synthesis, characterization and antimicrobial activity of Au NPs using *Euphorbia hirta* L. leaf extract, *Colloids Surfaces B Biointerfaces.* 108 (2013) 60–65.
- [156] B. Amini, M. Kamali, M. Salouti, P. Yaghmaei, Fluorescence bio-barcode DNA assay based on gold and magnetic nanoparticles for detection of Exotoxin A gene sequence, *Biosens. Bioelectron.* 92 (2017) 679–686.
- [157] A.M. Alkilany, L.B. Thompson, S.P. Boulos, P.N. Sisco, C.J. Murphy, Gold nanorods: their potential for photothermal therapeutics and drug delivery, tempered by the complexity of their biological interactions, *Adv. Drug Deliv. Rev.* 64 (2012) 190–199.
- [158] M.R.K. Ali, Y. Wu, Y. Tang, H. Xiao, K. Chen, T. Han, N. Fang, R. Wu, M.A. El-Sayed, Targeting cancer cell integrins using gold nanorods in photothermal therapy inhibits migration through affecting cytoskeletal proteins, *Proc. Natl. Acad. Sci.* (2017) 201703151.
- [159] M.R.K. Ali, M.A. Rahman, Y. Wu, T. Han, X. Peng, M.A. Mackey, D. Wang, H.J. Shin, Z.G. Chen, H. Xiao, Efficacy, long-term toxicity, and mechanistic studies of gold nanorods photothermal therapy of cancer in xenograft mice, *Proc. Natl. Acad. Sci.* (2017) 201619302.
- [160] S. Alex, A. Tiwari, Functionalized gold nanoparticles: synthesis, properties and applications—a review, *J. Nanosci. Nanotechnol.* 15 (2015) 1869–1894.
- [161] M. Alagiri, P. Rameshkumar, A. Pandikumar, Gold nanorod-based electrochemical sensing of small biomolecules: A review, *Microchim. Acta.* (2017) 1–24.
- [162] M. Aioub, S.R. Panikkanvalappil, M.A. El-Sayed, Platinum-Coated Gold Nanorods: Efficient Reactive Oxygen Scavengers That Prevent Oxidative Damage Toward Healthy, Untreated Cells during Plasmonic Photothermal Therapy, *ACS Nano.* 11 (2017) 579–586.

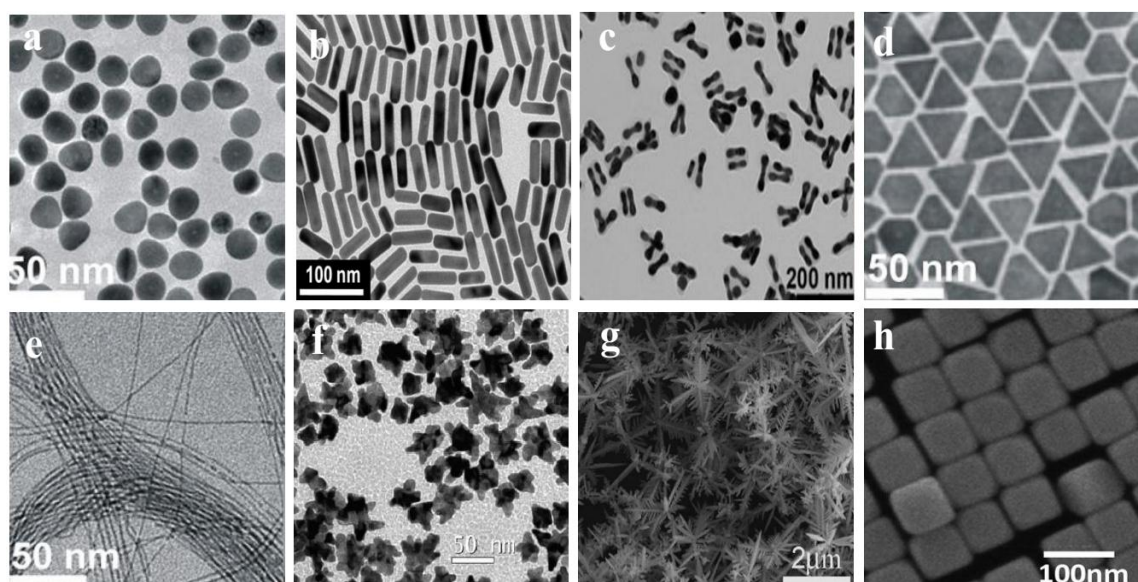


Fig. 1. (a-f) TEM images of AuNPs: a) quasi-spheres [20] b) nanorods [21] c) Nanodumbbells [22] d) triangular nanoprisms [20] e) Ultrathin nanowires [20] f) nanostars [23]; (g-h) SEM images of AuNPs: g) nanodendrites [24] h) nanocubes [25]. Copyright © American Chemical Society.

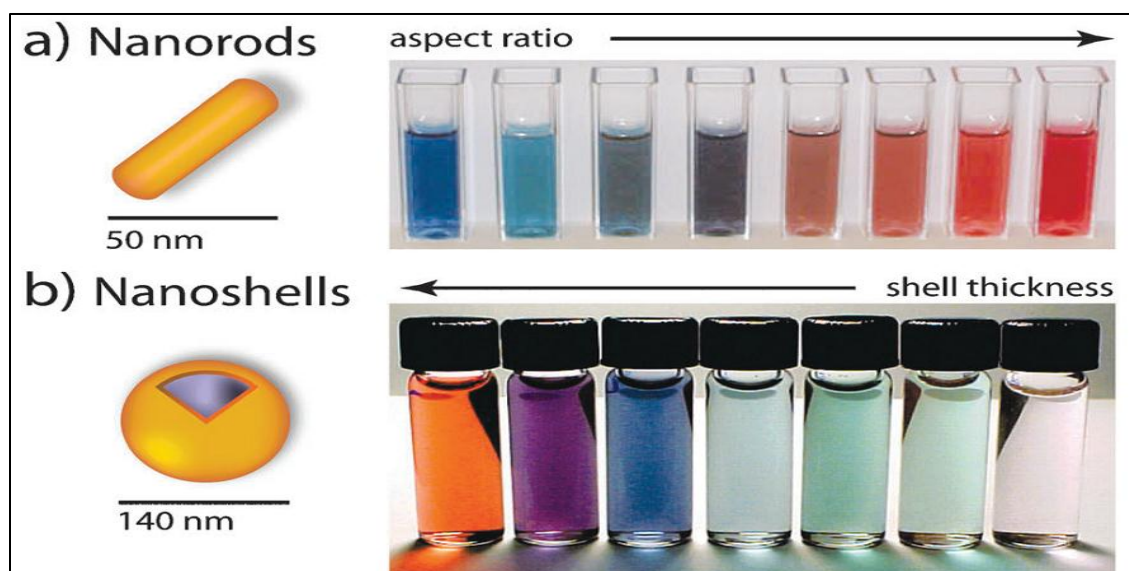


Fig. 2. Association of different morphologies and sizes of AuNPs with optical properties. (A) Gold nanorods, (B) Silica– gold core–shell nanoparticles [26].

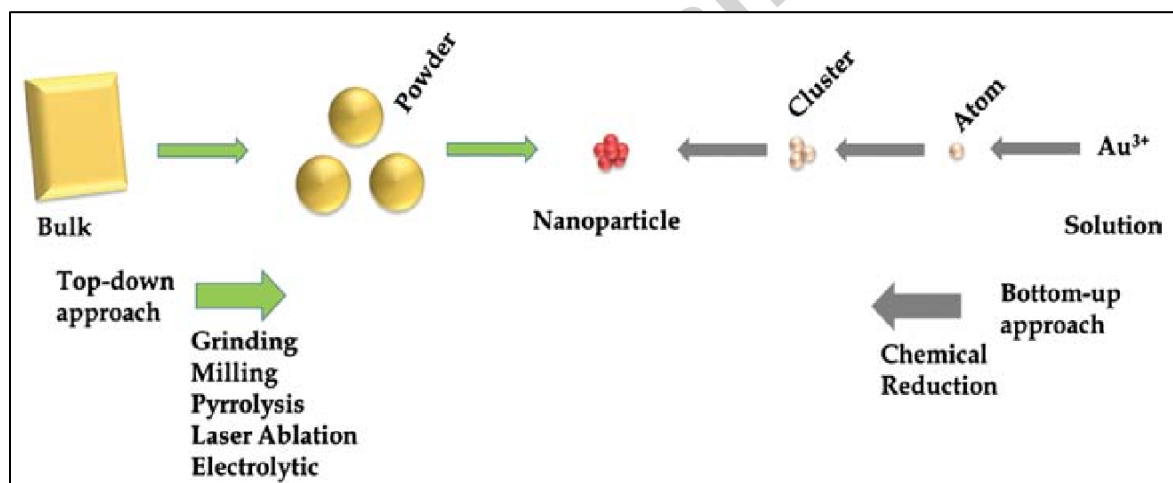


Fig. 3. Basic approaches for nanoparticle preparation [4].

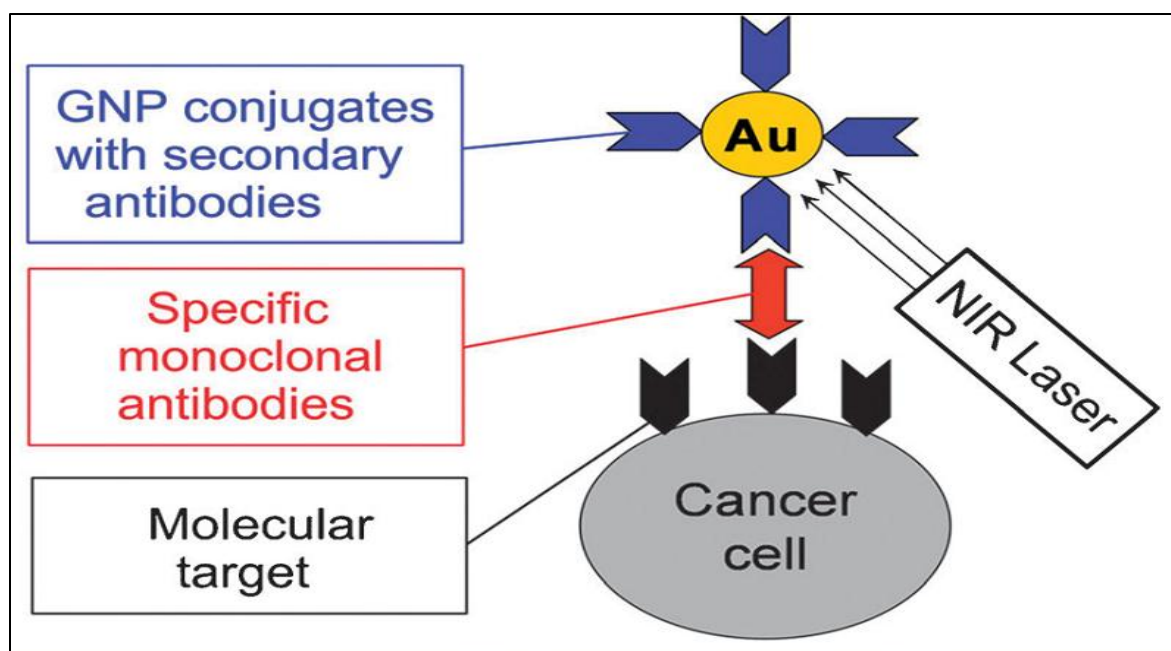


Fig. 4. Scheme for a PTT employing active delivery of AuNPs to cancer cells [79].

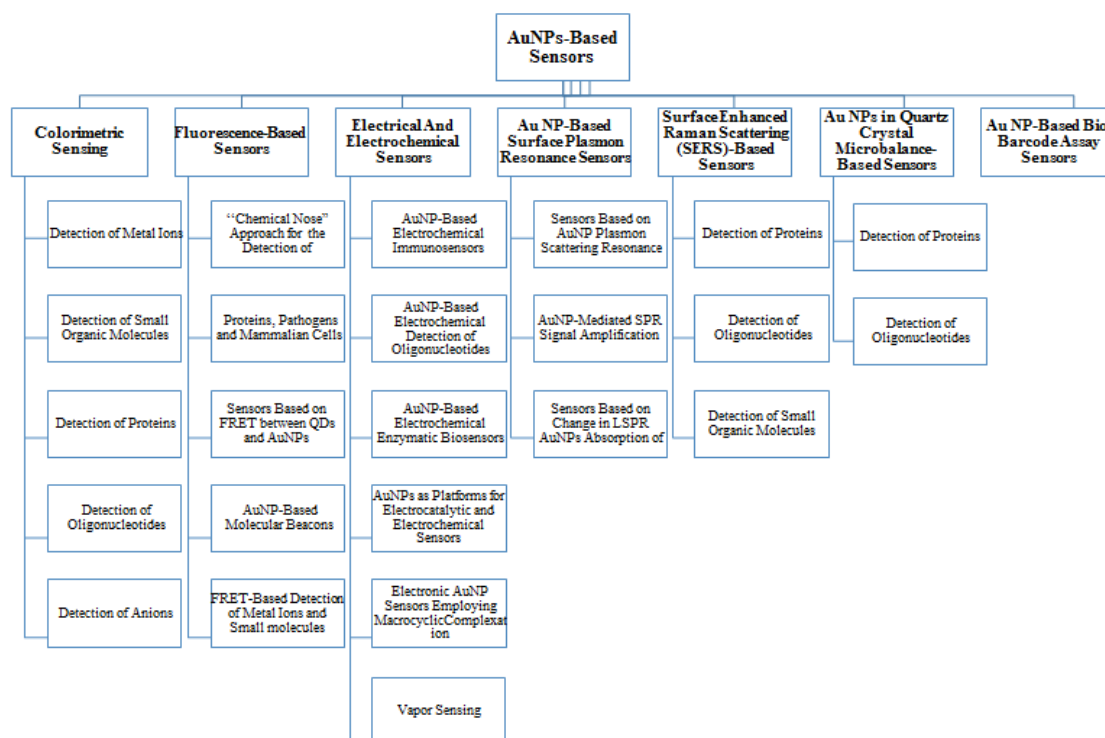


Fig. 5. Summary of different types of gold nanoparticles biosensors; utilizing the explicit physicochemical properties of this nanoparticles, various sensors can be designed.

Table 1 Biological resources for AuNPs synthesis.

Microorganism	Specious Name	Size range (nm)	Shape of AuNPs	Application	Ref
Plant-based compounds and derivatives	<i>Panax ginseng</i>	10 – 40	Spherical		[38]
	<i>Cacumen platycladi</i>	20-50	Spherical, triangular/hexagonal		[39]
	<i>Plumbago zeylanica</i>	>100	Spherical, triangles, hexagons	Antibacterial activity against Gram-negative and Gram-positive bacteria	[40]
	<i>Abelmoschus esculentus</i>	45–75	Spherical	Antifungal activity against <i>Puccinia graminis tritici</i> , <i>Aspergillus flavus</i> , <i>Aspergillus niger</i> and <i>Candida albicans</i>	[41]
	<i>Hibiscus sabdariffa</i>	25-30	Spherical	Selective toxicity towards U87 glioblastoma multiforme cell line under normal and hyperglycemic condition	[42]
	<i>Cucurbita pepo</i> L. leaves	10–15	Spherical		[43]
	<i>Morinda citrifolia</i>	12.17-38.26	Spherical	High potential for use in the treatment of anticancer, antidiabetic	[44]
	<i>Eucommia ulmoides</i>	~ 20	Spherical	Excellent catalytic activity	[45]
	<i>Trigonella foenum-graecum</i>	15-20	Spherical	Good catalytic activity for the reduction of 4-nitrophenol to 4-aminophenol by excess NaBH ₄	[46]
	<i>Zingiber officinale</i>	10	Spherical		[47]
	<i>Hibiscus rosa sinensis</i>	~14	Spherical, triangular		[48]
	<i>Rosa hybrida</i>	10	Cubic	High potential for use in biological applications	[49]
	<i>Euphorbia hirta</i> L.	6-71	Spherical	Antibacterial activity of the green synthesized Au NPs against bacterial strains of <i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> and <i>Klebsiella pneumonia</i>	[50]
	<i>Abelmoschus esculentus</i>	62	Spherical	Antifungal activity	[41]
	<i>Cassia auriculata</i>	15–25	Spherical, hexagonal, triangular	May promote anti-hyperglycemic	[51]
	<i>Couroupita guianensis</i> , Aubl.	26 ± 11	Cubic	antioxidant activity	[52]
	<i>Terminalia arjuna</i> .	20-50	Spherical	Enhance the mitotic cell division and pollen germination activity	[53]
	<i>Mentha piperita</i>	>100	Spherical	Antibacterial activity against clinically isolated pathogens	[54]
Bacteria	<i>Klebsiella pneumoniae</i> (MTCC- 4030)	10 - 15	Spherical	Antibacterial activity against <i>Escherichia coli</i>	[55]
	<i>Klebsiella pneumoniae</i>	35-65	Spherical	Medical and pharmaceutical applications	[56]
	<i>Geobacillus</i> sp. strain ID17	5–50	Quasi-hexagonal	Potential application of this microorganism in bioremediation of gold-bearing waste	[57]
	<i>Klebsiella pneumoniae</i>	35-65	Spherical		[56]
	<i>Escherichia coli</i> K12	50	Spherical	Catalytic activity	[58]

Table 1 (continued)

Fungi	<i>Acinetobacter sp. SW 30</i>	20 ± 10	Triangle, rod, spherical	[59]
	<i>Bacillus stearothermophilus</i>	5-30	Spherical, triangular	Detect toxin A (TOA) of <i>Clostridium difficile</i> based on an aptamer [60]
	<i>Bacillus Subtilis</i>	10-15	Cubic	Antimicrobial agents in packaging applications [61]
	<i>Aspergillus sydowii</i>	8.7–15.6	Spherical	[62]
	<i>Rhizopus oryzae</i>	43 ± 19	Flower-like	Stable in different physiological buffers and hemocompatible [63]
	<i>Penicillium citrinum</i>	60–80	Spherical	Antioxidant activity in pharmaceutical and cosmetic industries [64]
	<i>Alternaria sp.</i>	7–13 and 15–18 and 69-93	Quasi-spherical, Spherical, square, rod, pentagonal, hexagonal	[65]
	<i>Neurospora crassa</i>	32	spherical	[66]
	<i>Spirulina platensis</i>	~5	Spherical	Antibacterial activity against Gram positive organisms [67]
	<i>Turbinaria conoides and Sargassum tenerrimum</i>	27–35	Spherical	Catalytic activator [68]
Algae	<i>Sargassum swartzii</i>	35	Spherical	Anticancer for HeLa cells [69]
	<i>Stoechospermum marginatum</i>	18.7-93.7	Hexagonal, triangle	Antibacterial activator [70]
	<i>Cystoseira baccata (CB)</i>	8.4 ± 2.2	Spherical	Activity in colon cancer cells (cytotoxic effect against colon cancer cells) [71]
	<i>Tetraselmis kochinensis</i>	5–35	Spherical	Non toxic [72]
	<i>Kappaphycus alvarezii</i>	10-40	Spherical	[73]
	<i>Instant high-sugar dry yeasts</i>	Dependent on PH	Triangle, truncated triangle, hexagon Nano plates	Plasmonic Properties [74]
Yeast	<i>Yeast extract mannitol (YEM)</i>	4-20	Spherical	[75]
	<i>Baker's yeast (Saccharomyces cerevisiae)</i>	~5.0 ± 2.0	Spherical	Anticancer evaluation against Ehrlich ascites carcinoma cells [76]
	<i>Hansenula polymorpha</i>	20 – 40	Spherical	[77]
	<i>tobacco mosaic virus (TMV)</i>	10–40		[78]
Viruses				

Table 2 Various methods were for Au NRs Synthesis [9].

synthetic techniques		Type of Au NRs	
I	Aqueous wet-chemical CTAB-mediated synthetic procedures	crystalline nanorods	single-crystalline structure
			pentahedrally-twinned structure
II	Reduction of gold inside a sort template	polycrystalline structures	

III Synthesis in organic solvents

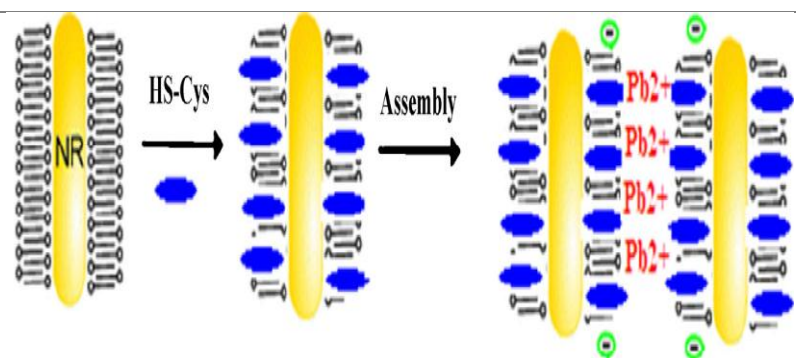
ultrathin rods and wires

Table 3 Au NRs properties and their applications in biosensing.

Properties	Agent or pathogen	Schematic representation of Au NRs biosensor	Ref
Localized surface plasmon resonance (LSPR)	Hepatitis B virus	The AU NRs biosensor based on the localized surface plasmon resonance (LSPR) behavior was designed to detect the hepatitis B surface antigen (HBsAg) which indicates active viral replication of hepatitis B virus (HBV).	[142]

Table 3 (continued)

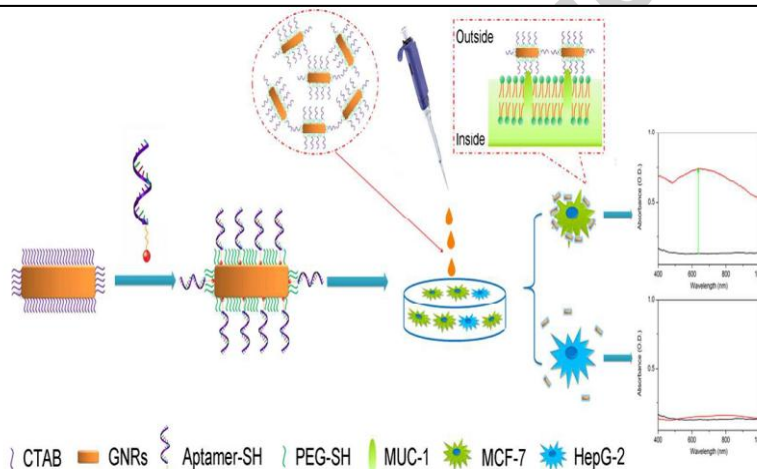
Lead



[144]

Aggregation-based Au NRs probe has been developed for the detection of Pb²⁺ in aqueous solution, based on the fact that Pb²⁺ ions induce assembly mediated signal enhancement of cysteine-functionalized Au NRs.

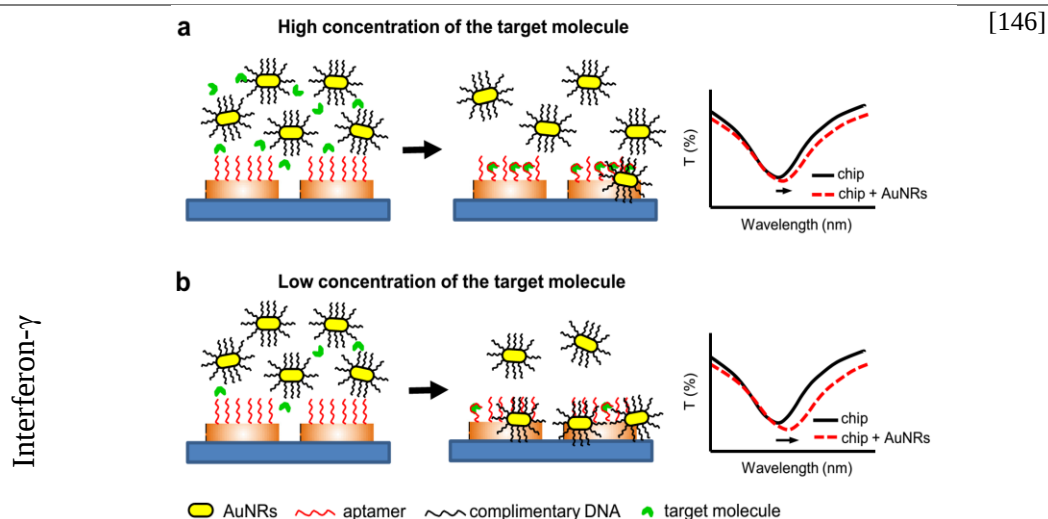
MCF-7 breast cancer cells



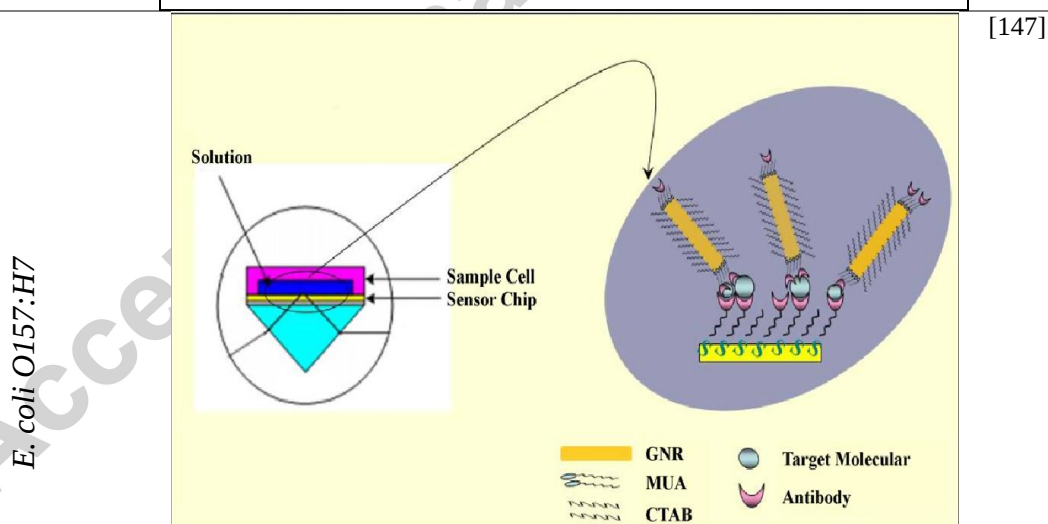
[145]

After the modification by MUC-1 aptamers, the special reaction between functionalized AU NRs and target cells were detected by LSPR spectra. Because of the anisotropy of AU NRs, the longitudinal LSPR band was demonstrated to be more sensitive in reflecting the dispersion state of nanoparticles than the transverse one both experimentally and theoretically.

Table 3 (continued)



Using the DNA-functionalized Au NRs to increase the spectral shift in LSPR sensing with the aptamer-functionalized LSPR chip; (a) When the concentration of the target molecule is high, the number of the Au NRs immobilized on the chip surface through DNA hybridization is reduced, and the LSPR shift induced by the Au NRs is small. (b) When the concentration of

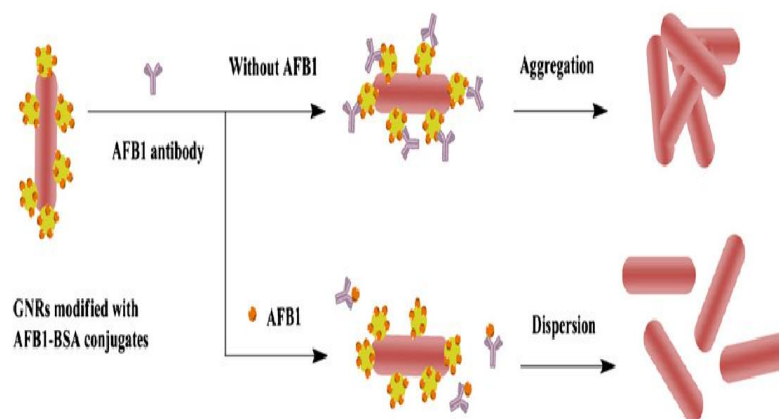


The diagram of combined target molecular to antibody with AU NRs in SPR sensor; AU NRs were used to enhance the sensitivity of the biosensor for *Escherichia coli* O157:H7 in surface plasmon resonance (SPR) system.

Table 3 (continued)

Optical property

Aflatoxin B1 (mycotoxins produced by *Aspergillus flavus* and *Aspergillus parasiticus*)

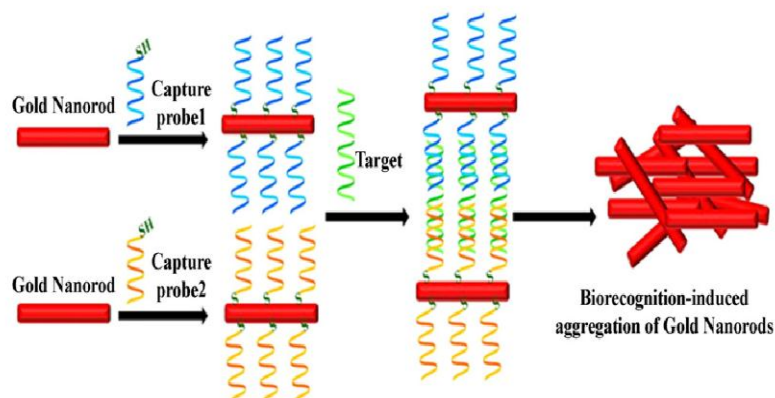


[143]

AFB1 sensing based on competitive dispersion of AU NRs; AU NR-AFB1-BSA (bovine serum albumin) conjugates aggregated after mixing with free antibodies, resulting in significant changes in absorption intensity; At the same time the existence of AFB1 molecules in samples caused dispersion of nanorods, as a result of competitive immune-reaction with antibodies.

[141]

Chlamydia trachomatis



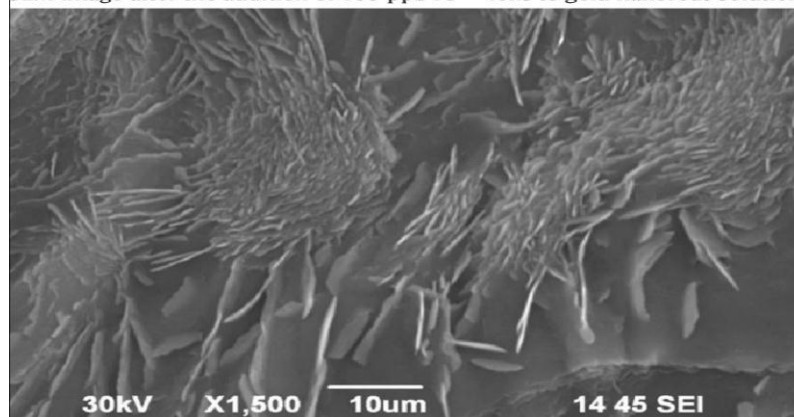
Schematic representation of DNA hybridization detection with Au NRs. Specific target recognition induces aggregation of gold nanorods.

Table 3 (continued)

Fe(III) ions

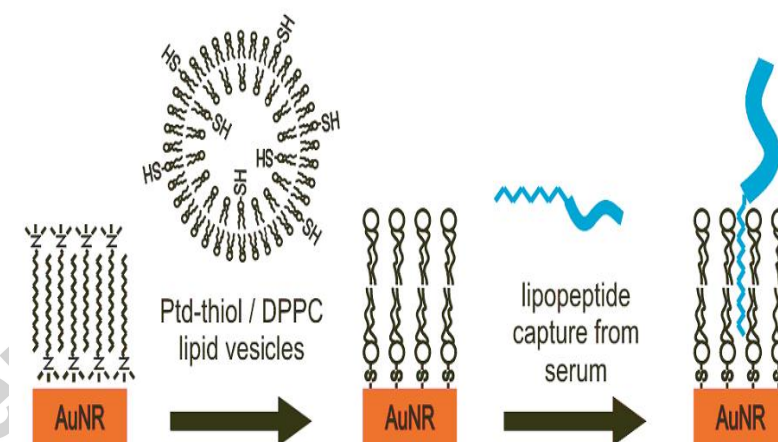
SEM image after the addition of 100 ppb Fe^{3+} ions to gold nanorods solution

[148]



SEM image after the addition of 100 ppb Fe^{3+} ions to gold nanorods solution. A novel and systematic method for sensitive detection of Fe^{3+} ions in aqueous media using Au NRs as nano sensors is presented. The interaction of Fe^{3+} ions with Au NRs was investigated by surface enhanced Raman scattering (SERS) using crystal violet (CV) molecule.

lipophilic analytes



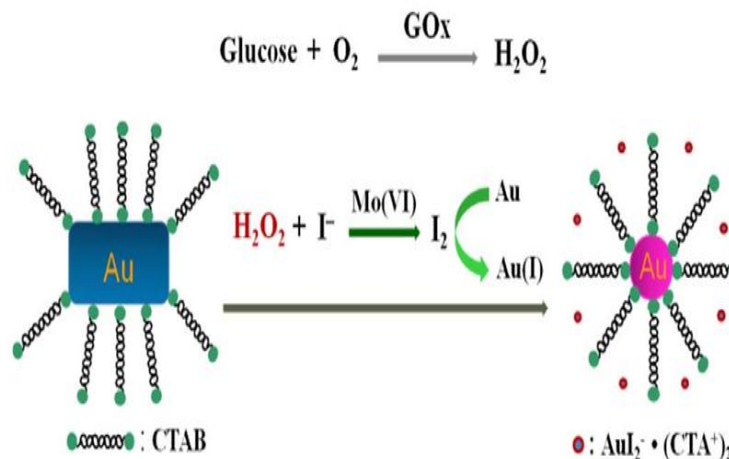
[149]

The production of lipid-capped Au NRs was carried out by replacing the CTAB bilayer with a lipid layer using small unilamellar vesicles containing lipids with a thiol head group. The lipid-capped Au NRs can be used for the capture of lipophilic analytes from complex solutions for label-free biosensing applications.

Table 3 (continued)

[150]

Glucose

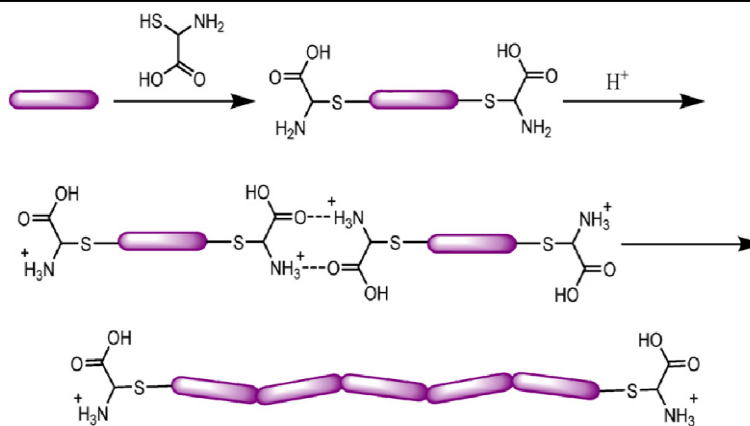


The mechanism of the AU NR-etching-based glucose sensor; AU NRs are etched by I₂ along the longitudinal direction due to less surface passivation and/or the higher reaction activities at the tips of AU NRs. The enzymatic-like etching of GNRs result in the shortening of AU NRs with increase in glucose concentration. The shortening of AU NRs leads to a blue-shift of the LSPR of AU NRs, accompanied by a color change from blue to red

[151]

self-assembly

Cysteine

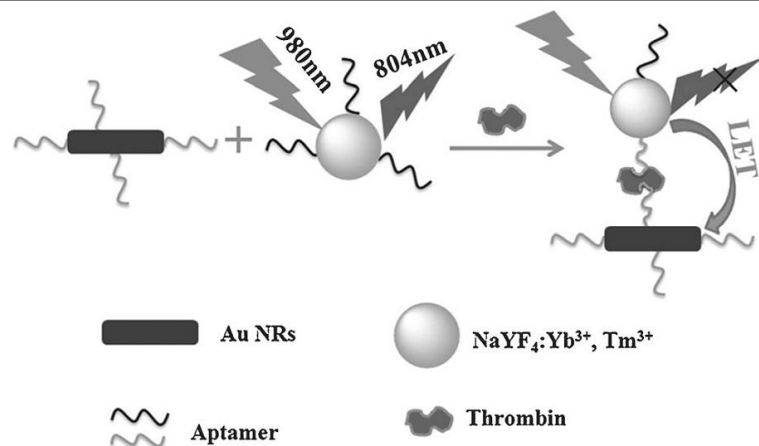


Detection of cysteine using self-assembly of gold nanorods in the acidic medium; This methodology is based on the end-to-end assembly of gold nanorods and it is very sensitive due to the useful formation of the gold nanorods chain in the presence of cysteine under acidic aqueous solution.

Table 3 (continued)

luminescence energy transfer (LET)

Thrombin

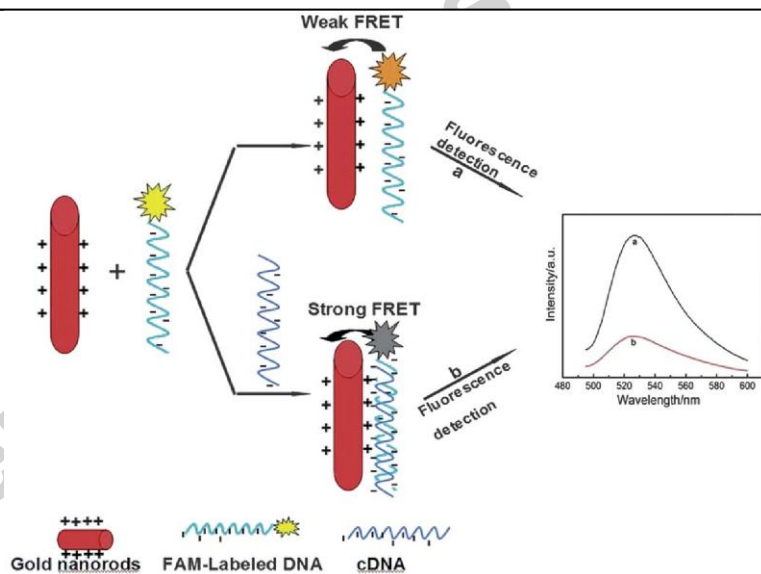


[152]

Two probes (UCNPs-TBA15 and Au NRs-TBA29) were obtained by functionalization of the poly (acrylic acid) (PAA)-modified NaYF₄: Yb³⁺, Tm³⁺ UCNPs and the CTAB-stabilized Au NRs with thrombin aptamers.

Fluorescence resonance energy transfer

Hepatitis B virus DNA

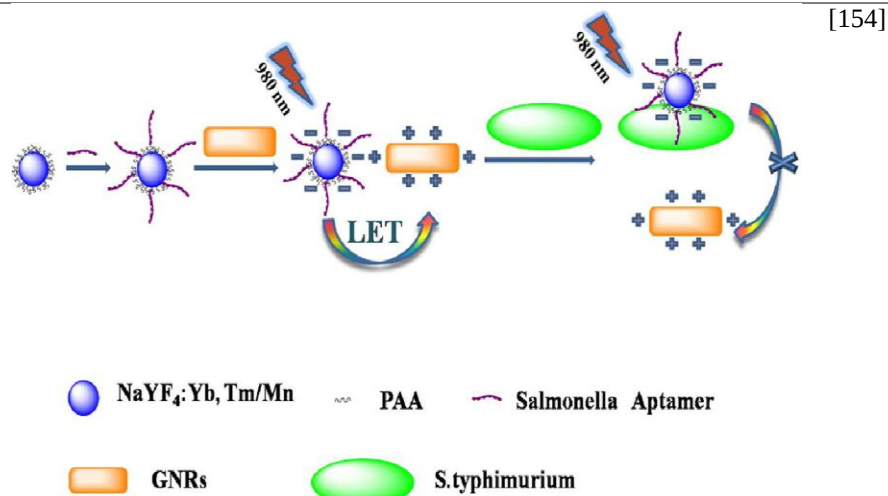


[153]

Schematic representation of the strategy for DNA hybridization detection; The Au NRs-based fluorescent biosensor was designed for the detection of HBV sequences, and FAM-ss DNA was selected as the probe DNA.

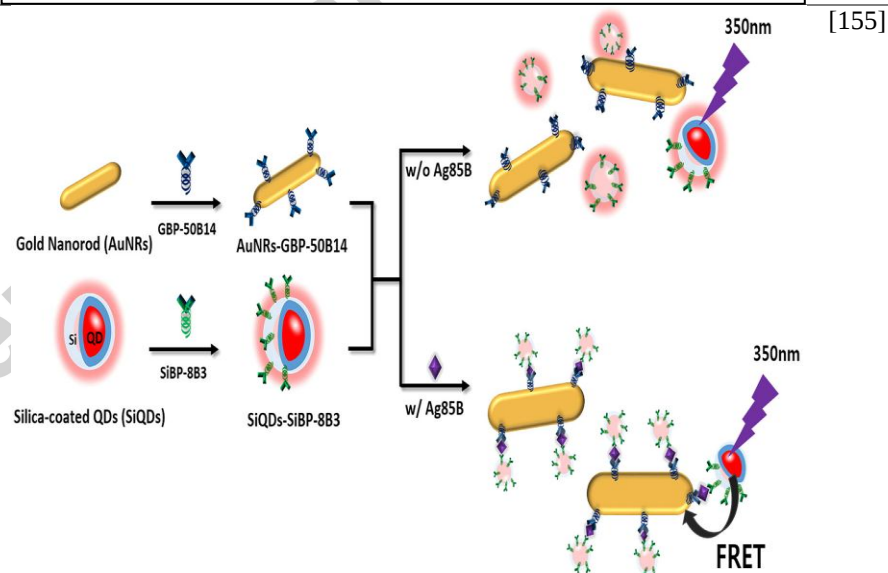
Table 3 (continued)

Salmonella typhimurium



The fabrication process of bio functionalized nanoparticles and principle of the performed bioassay; Au NRs served as the acceptor (positively charged). On the basis of the electrostatic interactions, the donor and the acceptor were brought into close proximity, which resulted in luminescence quenching. Subsequently, *S. typhimurium* was added to the system.

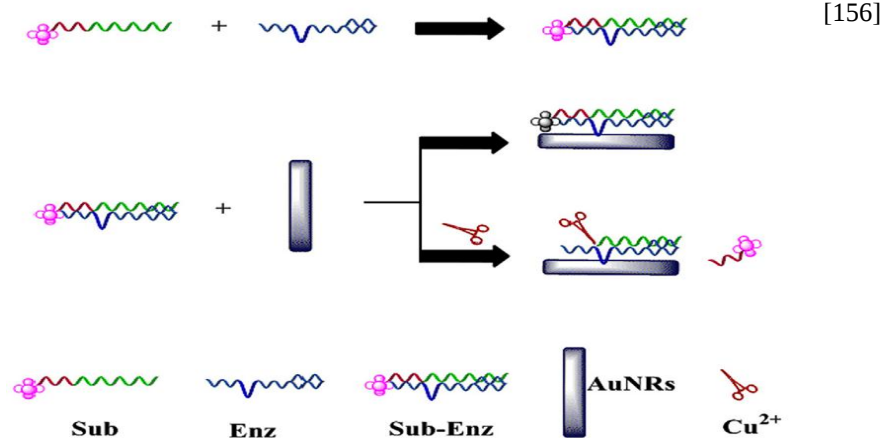
Mycobacterium tuberculosis Ag85B



The fluorescent analysis method for the detection of secreted antigen from *M. tuberculosis* based on sandwich assay via antigen-antibody interaction using Au NRs and SiQD; The secreted antigen Ag85B and fused recombinant antibodies GBP-50B14 and SiBP-8B3 were employed for the detection of the active *M. tuberculosis*.

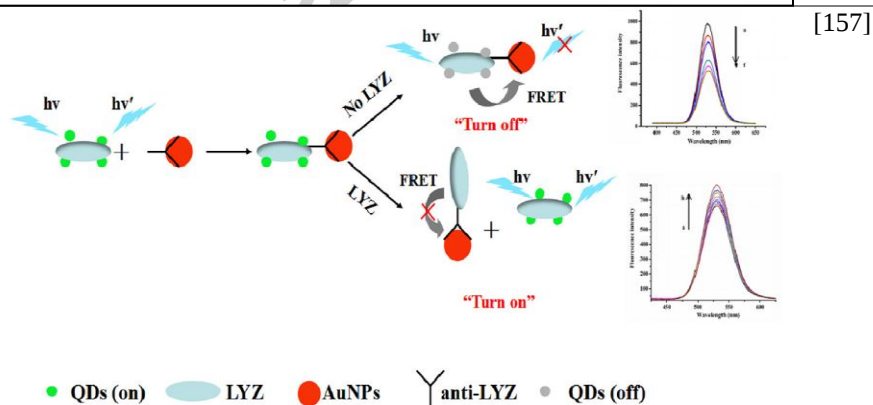
Table 3 (continued)

Copper(II)



The strategy of Cu^{2+} ions detection using Cu^{2+} dependent DNA-cleaving DNAzyme via Au NRs-based fluorescence resonance energy transfer assay; In the absence of Cu^{2+} , the fluorescence of TAMRA was significantly quenched. Upon the presence of Cu^{2+} , the DNA was cleaved at the guanine base site into two parts. The cleaved short TAMRA-labeled DNA fragment has a lower charge density than the Sub-Enz duplex, which weakened the electrostatic interaction between DNA and Au NRs and the fluorescence intensity of the system recovered.

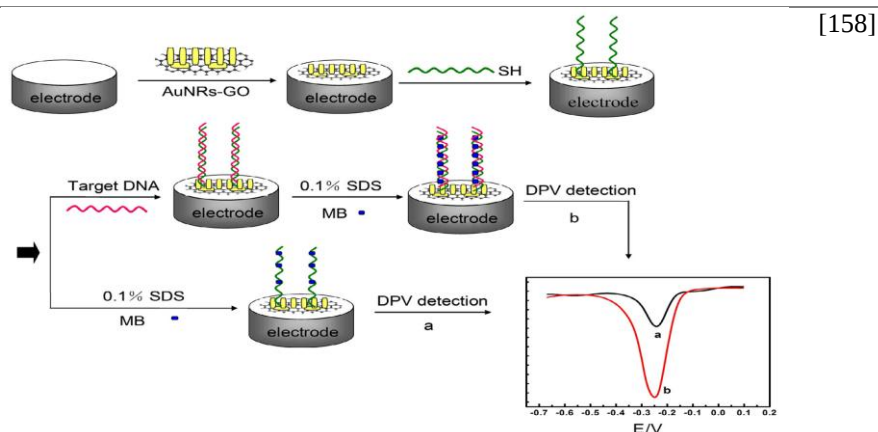
Lysozyme



Principle of the fluorescence switch control system. the strong fluorescence of LYZ-modified Quantum Dots (QDs-LYZ) could be effectively quenched by gold nanoparticle (Au NPs) modified with antibodies against LYZ(anti-LYZ) due to fluorescence resonance energy transfer (FRET) between QDs-LYZ and anti-LYZ-Au NPs. The fluorescence can be reversibly recovered by LYZ (on state) owing to specific competitive interactions between LYZ, QDs-LYZ and anti-LYZ-Au NPs. The interaction of QDs-LYZ with anti-LYZ-Au NPs was studied by absorption, fluorescence spectroscopy.

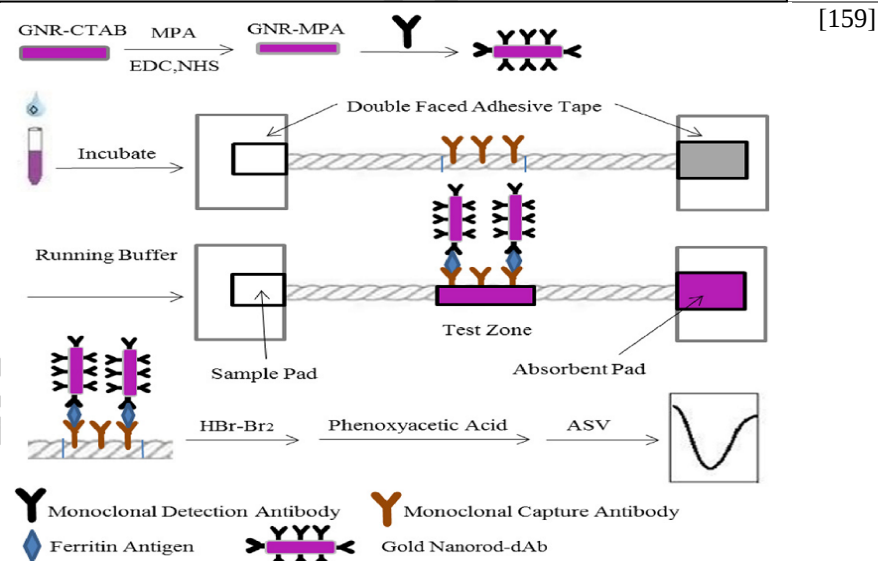
Table 3 (continued)

DNA



The DNA biosensor fabrication procedure; MB, methylene blue; SDS, sodium dodecyl sulfate; DPV, differential pulse voltammetry. An electrochemical sensor for sensitive and selective DNA detection was constructed based on Au NRs decorated graphene oxide (GO) sheets.

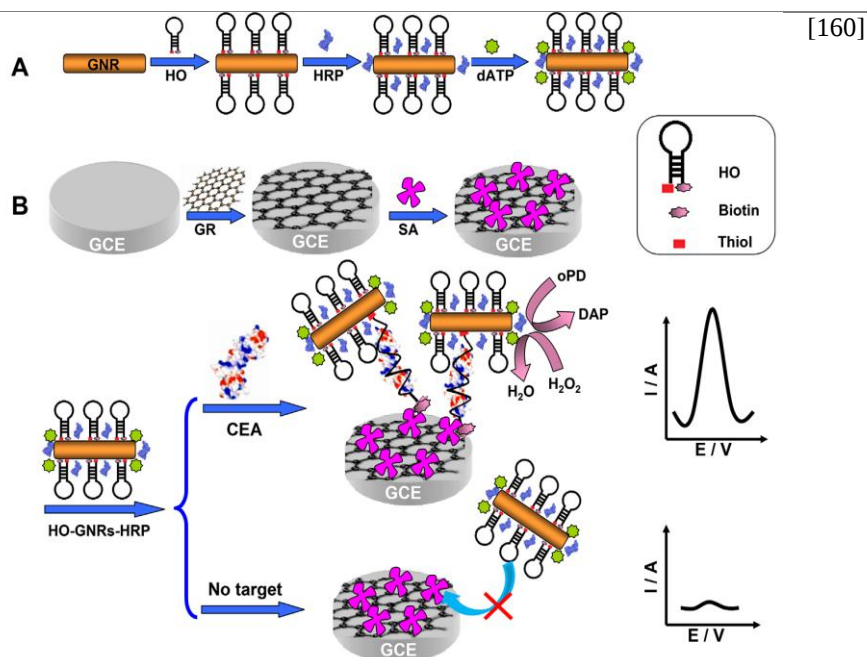
Human ferritin



The principle of the nature cotton thread based immunoassay device for electrochemical detection of ferritin. The solution of GNR-dAb-Ferritin complexes was applied to the sample pad, and immediately migrated along the cotton thread to cAb, which was pre-immobilized as the test zone of the cotton thread, where the ferritin antigen is captured to form sandwich AU NR-dAb-Ferritin cAb complexes. Because of the accumulation of AU NR complex, a purple band would appear on the test zone.

Table 3 (continued)

Electrical property



(A) Modification procedure of HO-GNRs-HRP conjugate. (B) Schematic illustration of the assembly process of SA-CS/GR/GCE and electrochemical sensing strategy for the detection of CEA: In the presence of target CEA, the binding reactions of CEA with the loop portions of the HO groups caused HO's loop-stem structure opened and exposed the biotins, and then HRP-AU NRs-HO conjugates were captured on graphene and streptavidin modified electrodes via the reaction between the exposed biotins and preimmobilized streptavidins. The accumulation of HRP effectively catalyzed the hydrogen peroxide-mediated oxidation of o-phenylenediamine to generate an electrochemical reduction current for CEA detection.

Table 3 (continued)

- Au NRs have attracted many interests because they possess certain specific features.
- Different controllable particles shapes and sizes of AuNPs have been synthesized by chemical, physical and biological methods.
- Various biomedical applications could be done by applying the physicochemical properties of AuNPs.

Accepted manuscript