

Versatile Carbon Nanofiber-Based Sensors

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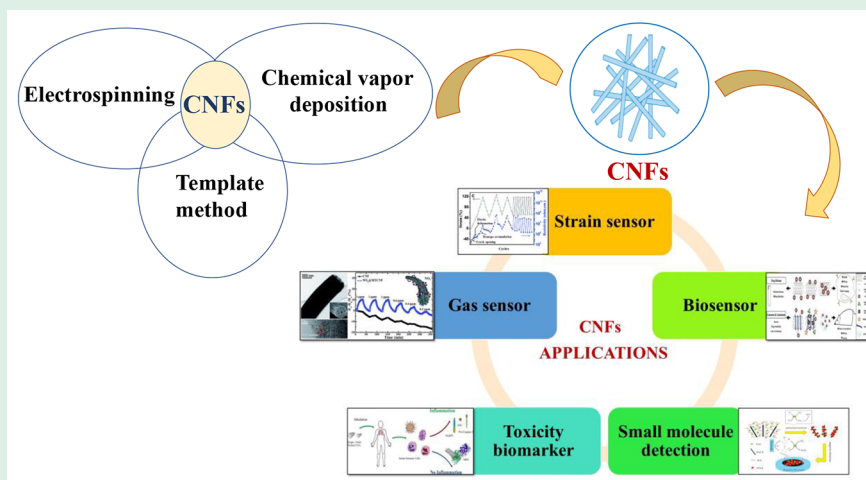


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ABSTRACT: Carbon nanofibers (CNFs) display colossal potential in different fields like energy, catalysis, biomedicine, sensing, and environmental science. CNFs have revealed extensive uses in various sensing platforms due to their distinctive structure, properties, function, and accessible surface functionalization capabilities. This review presents insight into various fabrication methods for CNFs like electrospinning, chemical vapor deposition, and template methods with merits and demerits of each technique. Also, we give a brief overview of CNF functionalization. Their unique physical and chemical properties make them promising candidates for the sensor applications. This review offers detailed discussion of sensing applications (strain sensor, biosensor, small molecule detection, food preservative detection, toxicity biomarker detection, and gas sensor). Various sensing applications of CNF like human motion monitoring and energy storage and conversion are discussed in brief. The challenges and obstacles associated with CNFs for futuristic applications are discussed. This review will be helpful for readers to understand the different fabrication methods and explore various applications of the versatile CNFs.

KEYWORDS: CNFs, sensors, biosensors, toxicity, detection limit

1. INTRODUCTION

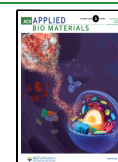
Carbon-based nanomaterials are shining in the field of sensors owing to their unique properties and chemical flexibility. They have prodigious potential in numerous applications, electrochemical supercapacitors,¹ water splitting,² photocatalytic applications,^{3–5} hydrogen production,^{6–8} drug detection,^{9–11} pollutant degradation,^{5,12–14} sensing,^{15–19} optical sensors,¹⁹ battery technology,^{20,21} etc. Carbon nanofibers (CNFs), 1D nanomaterials, have a nanometer range diameter.²² Fishbone, platelet, and parallel²³ structures are the common forms of CNFs based on the angle between graphene layers and the growth axis²⁴ (Figure 1). In herringbone or fishbone ($0^\circ < \text{angle} < 90^\circ$) structures, graphene layers are stacked obliquely with regard to the fiber axis; in platelet (angle = 90°) structures, graphene layers and the fiber axis are perpendicular;

and in parallel (angle = 0°) structures, the growth axis and graphene layers are parallel to each other.²⁴ Parallel CNFs always exhibit a hollow core (see Figure 1). CNFs show hexagonal carbon arrangements with three sp^2 hybridized carbons having 1.415 Å bond length and a fourth carbon attached via intermolecular stacking with an interlayer spacing of 3.35 Å.²⁵ The important properties of CNFs include low thermal expansion, light weight, high tensile strength, chemical

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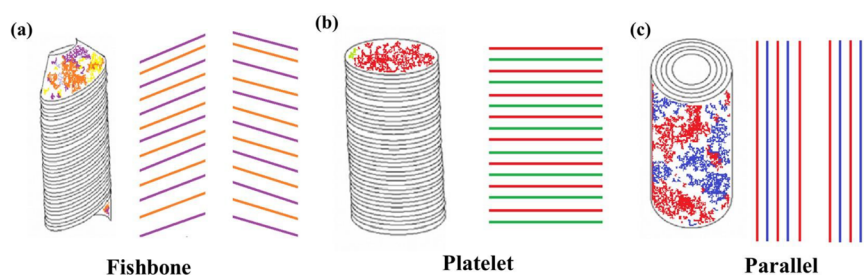


Figure 1. Different forms of CNFs: (a) fishbone, (b) platelet, and (c) parallel.

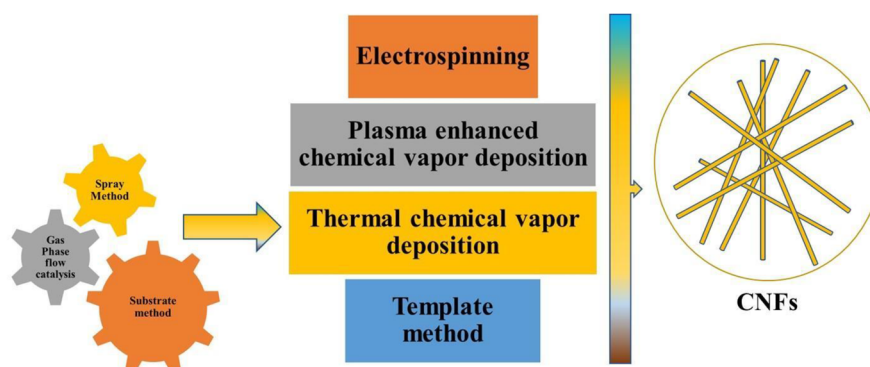


Figure 2. Various techniques for the fabrication of CNFs.

resistance, stiffness, excellent thermal conductivity, and high temperature tolerance.²⁶ CNFs can be used as catalyst supports because of their favorable characteristics such as large surface area, stability under different reaction conditions, and porosity. They can also be used in fixed-bed applications as CNFs exhibit bulk crushing strength²⁷ of 1 MPa. Electric conductivity is significant in various applications ranging from electronics to composites. CNFs are organized as sp^2 -discontinuous linear filaments with an aspect ratio²⁸ greater than 100:1, which provides excellent surface area to volume ratio. CNFs can be treated chemically to enhance specific surface area and surface oxygen moieties. Functionalization in an acidic oxidizing medium leads to surface chemistry variation, thus increasing the hydrophilicity in CNFs, which is effective for excellent dispersion in an aqueous medium for carbon particles.²³ There are various routes for the synthesis of CNFs, which include chemical vapor deposition, electrospinning, drawing, templating, and phase separation.²⁹

CNFs have edge planes and graphite planes showing great potential for surface modifications to build functional hybrids that find applications as nanodevices,³⁰ in sensing,³¹ in biomedicine, in energy storage,³² as batteries,³³ as supercapacitors,³⁴ in energy applications,³⁵ in fuel cells or solar cells,^{36,37} in drug delivery,³⁷ and in environmental science areas.³⁸

This review is organized as follows: **Section 1** gives a general introduction to CNFs and their different forms and properties. **Section 2** gives insight into various synthesis methods for CNFs, like electrospinning, plasma-enhanced chemical vapor deposition, thermal chemical vapor deposition, and template methods, with the merits and demerits of each technique. **Section 3** provides a comprehensive discussion on various sensing applications of CNFs, including strain sensors, biosensors, small molecule detection, food preservative detection, toxicity biomarker detection, disease detection, and gas sensors, and other applications of CNFs like human

motion monitoring and energy conversion. Lastly, the challenges and obstacles related to CNFs are given. The detailed review will aid understanding of the broad range of possible usage of CNFs. This review will provide the detailed bibliographic data published by various researchers on sensing applications of CNFs under one roof, which can help readers to understand better.

2. SYNTHESIS OF CARBON NANOFIBERS

CNFs can be synthesized using electrospinning, thermal chemical vapor deposition (spray method, gas-phase flow catalysis, and substrate method), plasma-enhanced chemical vapor deposition, and template methods as shown in **Figure 2**.

2.1. Electrospinning Technique. Electrospinning is an efficacious method to make polymeric nanofibers because it allows modification of chemical composition, fiber morphology, fibrous architecture, and functionality.^{39,40} This technique produces nanofibers ranging in size from a few tens of nanometers to a few micrometers, such as in nonwoven fabrics or mats.⁴¹ The method is simple, less-expensive,⁴² and biocompatible⁴³ and can produce continuous CNFs.^{44–46} In this method, the polymer solution is first charged using static electricity (thousands to tens of volts) and in an electric field. Then at the spinning port, the charged polymer tends to form a Taylor cone. The cone is accelerated and drafted as the electric force approaches the spinning mixture tension. The movable jet is thinned and drafted progressively, and fibers are eventually deposited on a collecting plate, creating a fibrous mat close to nonwoven cloth. The fiber mat is then preoxidized in the atmosphere; after that carbonization in a nitrogen atmosphere produces the CNFs. Parameters like concentration, spinning voltage, working distance, and solution feeding rate of the polymer solution are the critical electrospinning processing parameters that can modify the morphology of the fibers formed.⁴⁷ For instance, as the

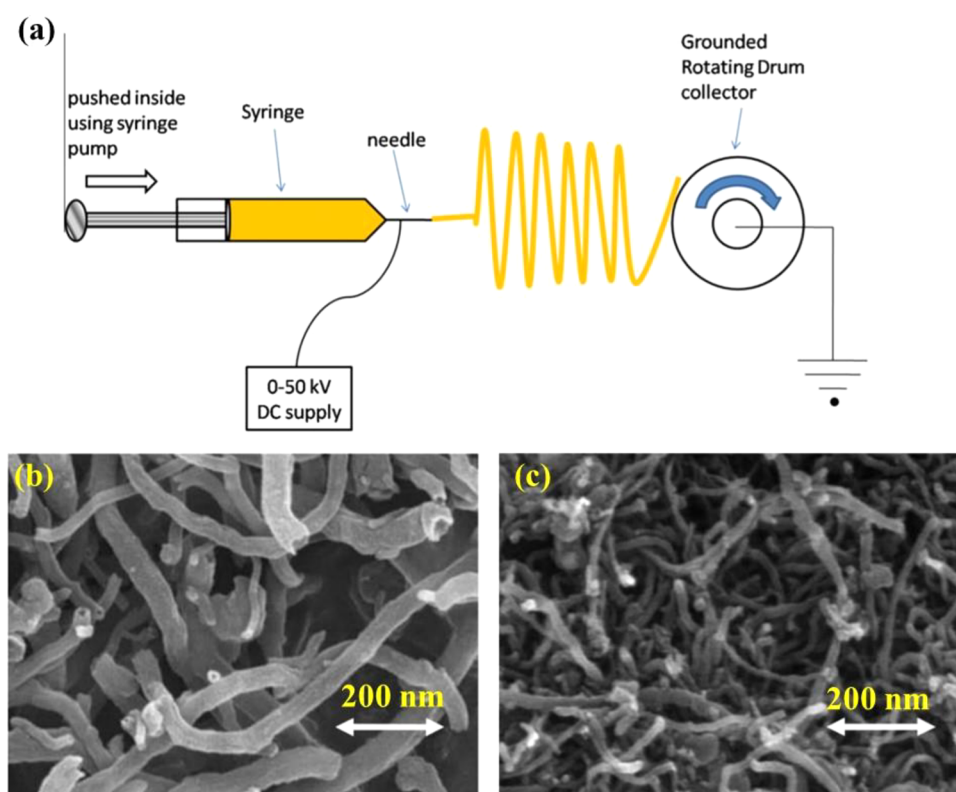


Figure 3. (a) Illustration showing the setup for the electrospinning technique. Reprinted with permission from ref 49. Copyright 2012 Elsevier. (b, c) SEM micrographs of CNFs synthesized by the chemical vapor deposition method. Reproduced from ref 52. Copyright 2008 American Chemical Society.

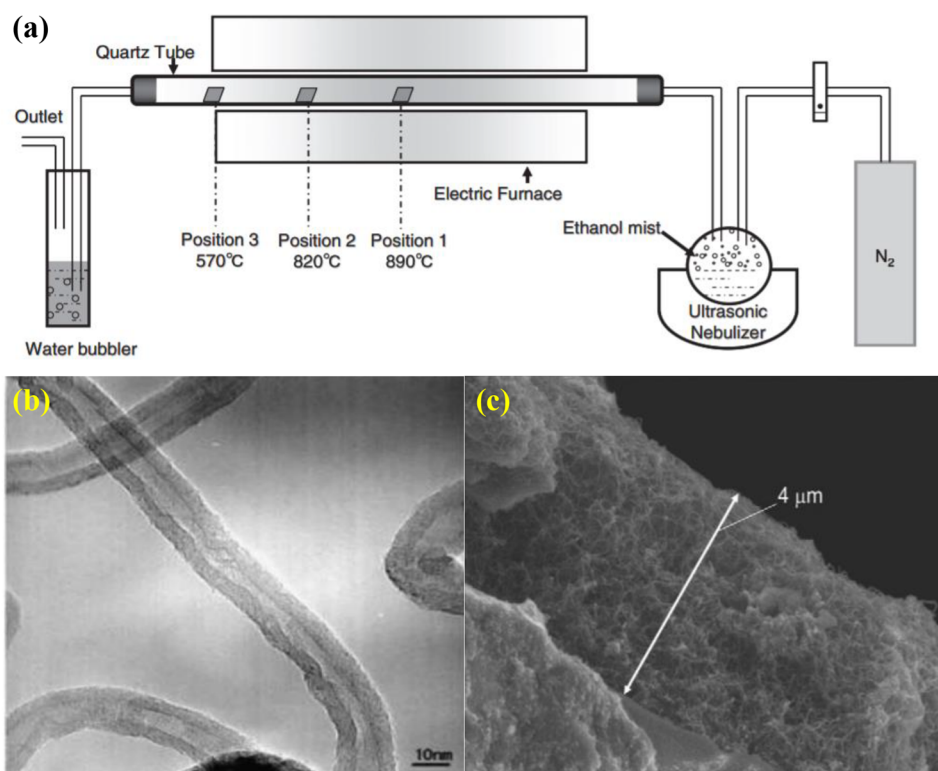


Figure 4. (a) Spray method setup used to produce CNFs, (b) TEM micrograph of CNFs synthesized by gas-phase flow catalysis, and (c) SEM micrograph of CNFs produced on cordierite substrate. Reprinted with permission from refs 55–57. Copyright 2011, 2004, and 2007, respectively, Elsevier).

carbonization temperature rises, the fiber size shrinks.⁴⁸ This method is most effective in synthesizing CNFs (10 nm to 10 μm) with a persistent framework. See the layout of this technique⁴⁹ in Figure 3a. The electrospinning method has various benefits; for example, the nanofiber nonwoven fabric can be produced directly, spinning can be done at room temperature, and a wide range of raw materials can be used.²⁹

2.2. Thermal Chemical Vapor Deposition. Chemical vapor deposition (CVD) involves thermally decomposing cheap a hydrocarbon composite using a metal catalyst within a temperature range of 500–1000 °C to produce CNFs.^{50,51} Hulicova-Jurcakova et al.⁵² synthesized graphitic CNFs using the chemical vapor deposition method, the SEM images of which are illustrated in Figure 3b,c. This technique can produce microscale CNFs having a higher aspect ratio. Further classification of this method into the spray method, gas-phase flow catalysis, and the substrate method is done based on the catalyst added. Parameters like high temperature, a vacuum environment, and volatile precursors are required for CVD. Ideal alignment, growth rate, size, and diameter can be achieved using this method.⁵³

2.2.1. Spray Method. For the spray method to synthesize CNFs, a catalyst is sprayed in benzene, and the mixture containing the catalyst is then sprayed into a high-temperature reaction chamber. Spray-growing carbon fiber allows continual injection of the catalyst, which creates favorable conditions for the industrial continuous processing.⁵⁴ During spraying, the catalytic particles are unevenly distributed and hard to monitor, leading to minute proportions of CNFs, as well as the formation of some carbon black particles. Bao et al.⁵⁵ synthesized CNFs using a spray pyrolysis method by placing ethanol in a medication cup. The schematic of the spray method used to synthesize CNFs is shown in Figure 4a.

2.2.2. Gas-Phase Flow Catalysis. This technique explicitly allows catalyst precursor heating and in the form of gas adds it into the chamber with hydrocarbon gas. Hydrocarbon gas and catalyst are decomposed and eventually agglomerated into nanoparticles, which are coated with CNFs.⁵⁸ Lim et al.⁵⁶ synthesized tubular CNFs (Figure 4b) over a Co–Mo catalyst as support using CO as a carbon source by the gas phase flow catalysis method, leading to continuous production and high yield per unit time as the amount of catalyst volatilization can be regulated in this method.

2.2.3. The Substrate Technique. This process involves SiO₂ fibers or ceramic as a substrate to spread the nanocatalytic particles (transition metals, Ni, Fe, and Co) uniformly on the surface. Hydrocarbon gas pyrolysis onto the catalyst's surface was done, and carbon was deposited to synthesize CNFs. Garcia-Bordejé et al.⁵⁷ synthesized high purity CNFs utilizing C₂H₆/H₂ and CH₄/H₂ as the carbon source and catalyst (Ni) on a ceramic substrate at 600 °C. An SEM image of a CNF coating layer produced at 650 °C using cordierite substrate is shown in Figure 4c. The authors also investigated the effect of different temperatures, carbon sources, CNF porosity, uniformity, and layers thickness.⁵⁷

2.3. Plasma-Enhanced CVD. Plasma provides activation energy for CVD as it constitutes a huge number of high-energy electrons (100–300 eV). This method is performed at a lower temperature than the normal CVD method. The collision of a gas-phase molecule with electrons facilitates the gas molecules' decomposition, excitation, compounding, and ionization, thereby producing high-activity chemical groups.^{59–61} Saidin et al.⁵⁹ synthesized CNFs utilizing the plasma-enhanced CVD

(PECVD) method by employing a Ni/Cr-glass thin film catalyst for CNF growth. The influence of the different thicknesses of catalyst and acetylene to gaseous ammonia ratios were investigated, and they concluded that the optimum condition for the synthesis of CNFs is a 1:3 acetylene to NH₃ gas ratio with the width of 15 nm for the Ni/Cr-catalyst. Shoukat and Khan⁶⁰ synthesized vertically aligned CNFs by inductively coupled plasma-enhanced CVD using a temperature of ~650 °C and toluene as a carbon source. Gupta and Sharma⁶¹ studied the effect of N doping on CNF growth by plasma-enhanced CVD and determined that N-doped CNFs had a lower tip diameter, smaller height, and improved field emission characteristics compared to undoped CNFs. Low substrate temperature and high deposition rates are required in this method.⁶² Figure 5 shows the experimental setup for the

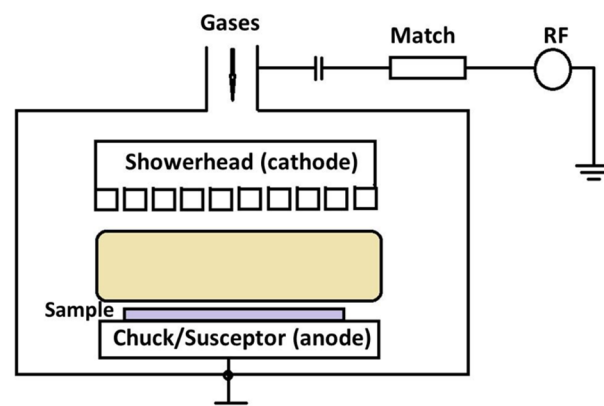


Figure 5. Experimental setup of plasma-enhanced CVD technique.

plasma-enhanced CVD method in which reactant gases go into the process chamber via the shower head. Potential is applied to shower head for the generation of plasma.

Aligned CNFs can be produced using plasma-enhanced CVD, but this is an expensive technique having low production efficiency. Parameters influencing the fabrication in this method are RF power, gas flow rates, chamber pressure, temperature, and spacing between the shower head and susceptor. A comparison of synthesis methods for CNFs with merits and demerits is summarized in Table 1.

Table 1. Advantages and Disadvantages of Synthesis Methods for CNFs

Methods	Advantages	Disadvantages
Electrospinning	facile, easy to build, requires high voltage, useful for porous CNFs, effective	requirement for post-treating, carbonization, and high voltage
CVD	high-quality CNFs, excellent conductivity, and efficiency	high cost and high energy consumption
Template synthesis	easy and simple process	removal of template required and expensive
Catalytic hydrogenation and hydro-thermal synthesis	high yield	hard to control, high cost, and high energy consumption

2.4. Template Method. This technique is somewhat less prevalent in producing CNFs. The basis for template synthesis of CNFs in this approach is the thermal breakdown of the carbon source in nanosized channels of an anodic aluminum oxide (AAO) film. The film is cleaned to dissolve the AAO leaving the CNFs (Figure 6). Wang et al.⁶³ synthesized CNFs

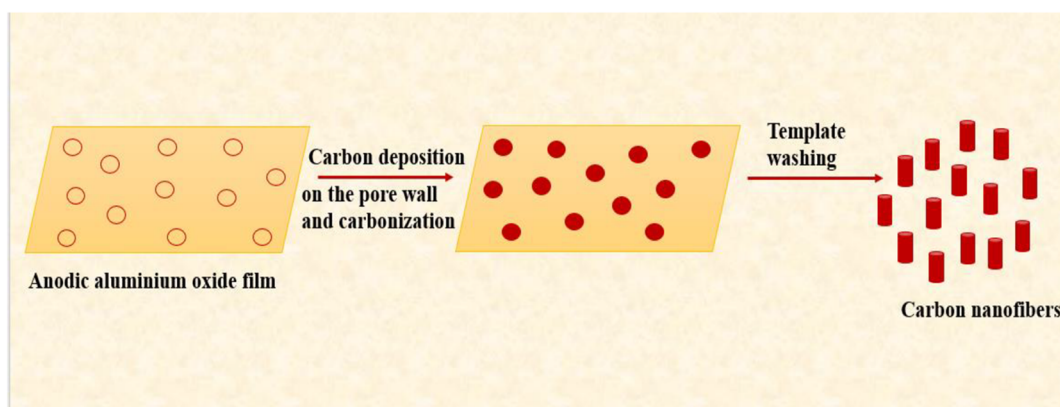


Figure 6. Formation of CNFs by the template method.²³

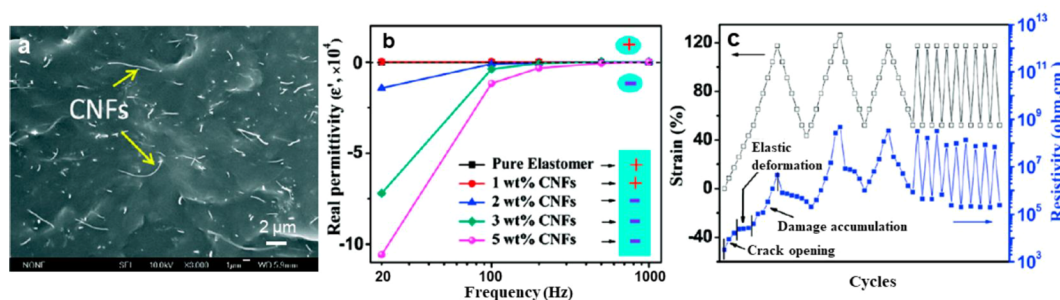


Figure 7. (a) SEM microstructure of the synthesized nanocomposite, (b) real permittivity in 20–1000 Hz frequency range, and (c) instantaneous resistivity response with a strain of 5 wt % CNFs/vistamaxx 6202FL nanocomposite toward cyclic strain. Reprinted with permission from ref 86. Copyright 2011 American Chemical Society.

using Pluronic F127/resol as a precursor in ethanol solvent at the carbonization temperatures between 350 and 700 °C. Thus, synthesized CNFs have nanofibers with linear mesocages having a pore size of ~ 18 nm. Xing and co-workers⁶⁴ synthesized the mesoporous CNFs using AAO and colloidal silica (porogen) as a film and phenolic resin as a precursor in ethanol at the carbonization temperature of 700 °C. The as-prepared CNFs are mesoporous hollow tubular fibers having a diameter of 21 nm. Processing parameters influencing the synthesis include template wall chemistry, pH scale, concentration, soaking times, and temperature, allowing the nanofibers to be controlled.⁶⁵

Unlike the electrospinning process, CNFs did not shrink during carbonization because of near contact between the nanofiber surface and the pore wall of AAO.⁶⁶

CNFs have also been prepared by other techniques like catalytic hydrogenation^{67,68} and hydrothermal synthesis.⁶⁹ Chen and co-workers⁶⁷ achieved carbon deposition from CO₂ through catalytic hydrogenation using Ni/Al₂O₃ samples containing various potassium (K) concentrations in which K as additive produced the CNFs or carbon deposition during a reverse water–gas shift reaction.⁶⁷ Ren et al.⁶⁸ used electrolytic conversion to synthesize the CNFs (with diameters of ~ 200 –300 nm) from atmospheric CO₂ dissolved in the molten carbonates. The authors found that catalytic hydrogenation is a cost-effective method to synthesize CNFs. Gupta et al.⁶⁹ synthesized CNFs by the microwave-assisted hydrothermal route using a reaction temperature of 400 °C for 2 h.

2.5. Surface Modification of CNFs. Regulating the CNF surface chemistry is important for knowing their functionality. Both physical and chemical modifications lead to alteration in

hydrophobicity, chemical reactivity, and surface charge. Modification was done by surface coating methods (chemical vapor deposition of thin-film coatings, electro- or electroless plating) and chemical functionalization.⁷⁰ Surface coating methods enhance the chemical stability and mechanical strength of CNFs and add functionality like variable electrical isolation and conductivity. Chemical functionalization enhances the dispersibility, surface reactivity, and wettability of CNFs. The chemical vapor deposition of thin-film coatings can be done via in situ PECVD coating and surface modification in which catalytic preparation of the carbon nanostructure affects the graphitic structure and condition of the surface of nanofibers.⁷¹ Silicon oxide is the most utilized coating deposited on CNFs through CVD.^{72–74} Moon et al. used SiO_x to coat multiwalled carbon nanotubes (MWCNTs) and discovered that the thin layer prevents thermal degradation and high temperature oxidation. It also enhances the field emission properties.⁷⁴ Researchers have also coated CNFs and polymers with thin films through CVD^{75,76} by electro- or electroless plating. Guillorn et al. used Au electrodeposition for decorating CNFs, validating that the sidewalls and underlying interconnected structures are passivated, and the extreme tips of the nanofibers were electroactive.⁷⁷ The chemical functionalization of CNFs permits controlled surface chemistry to allow flexible wettability, both hydrophilicity and hydrophobicity.⁷⁸ This can be done by wet etch treatment, plasma treatment, photochemical functionalization, thermal treatment, and adding polymers and linkers.⁷⁰

3. CARBON NANOFIBER-BASED SENSOR APPLICATIONS

CNFs having large surface areas facilitate the adsorption of a vast number of target molecules. CNFs have a high ability for electron transfer contributing to their extensive range of applications in chemical sensing.^{79,80}

3.1. Pressure or Strain Sensors. Traditional pressure sensors like silicon capacitive and silicon piezoresistive pressure sensors have less power consumption and high measurement precision and are economical but are unable to perform well in high-intensity piezoresistive measurements. CNFs are extensively used due to their cheap cost, excellent electrical conductivity, and potentially improved mechanical properties like strain power and fracture toughness.^{81–83} CNFs showed exceptional mechanical properties having Young's modulus of 25–200 GPa and up to 12 GPa tensile strength.⁸⁴ Factors affecting the mechanical properties of CNFs are dispersion and distribution,⁸⁵ adhesion and interface, and aspect ratio.⁸⁵ Zhu et al.⁸⁶ used a solvent-assisted casting process to create a conductive polymer nanocomposite fortified with CNFs using Vistamaxx 6202FL (15 and 85 wt % of ethylene and propylene, respectively) as a hosting polymer matrix material. As shown in Figure 7a,b, CNFs have tremendous mechanical deformation for use as strain sensors. When CNFs are stretched to 120% strain, strain recovery to 40% and resistivity changes of 10^2 – 10^3 times are observed (Figure 7c).

Yan et al.⁸³ used carbon/graphene composite nanofiber yarns (CNYs) to synthesize flexible strain sensors that showed brilliant stability and sensitivity. Multineedle electrospinning was used to fabricate CNYs, which were then stabilized and carbonized. When tested under 2% applied strain, the average gauge factor was more than 1700 when the number of substrate and yarns thickness were 129 and 4, respectively. During stretching–relaxation cycles (300), the strain sensor maintained a higher degree of stability. Hu et al.⁸⁷ synthesized a resistance-type strain sensor from metal-coated carbon nanofiller/epoxy composites. Multiwalled CNTs and vapor grown carbon fibers (VGCFs) with Ni, Cu, and Ag were utilized as nanofillers. The gauge factor of these sensors having Ag coatings is ~ 80 times larger than the gauge factor of a metal-foil strain gauge.

Azhari and Banthia⁸⁸ prepared two diverse types of electrically conductive cement-based piezoresistive sensors, one with carbon fibers alone and the other with a hybrid of carbon nanotubes (1%) with CNFs (15%). The latter sensor was repeatable and more accurate than the traditional cement-based sensor having a gauge factor of 445 and a load amplitude of 30 kN.⁸⁸ Baeza et al. calculated the properties of strain sensing of the cement composites. The authors synthesized a CNF cementitious composite (CNFCC) and a carbon fiber cement composite (CFCC), which were used as sensors for a conventional reinforced concrete beam. Higher sensitivity was observed with CNFCC with a gauge factor ~ 190 when 2 wt % CNFs were added to the cement matrix, and in doing so, the composite sensing ability was increased as the sensor became thinner.⁸⁹ Wang et al.⁹⁰ used different contents of CNFs with epoxy-based composites to synthesize strain sensors and investigated the effects on different parameters such as gauge factor, linearity, hysteresis, and repeatability. These findings showed that sensors having 0.58 vol % of CNFs had better piezoresistive output than those containing 0.29 vol % CNFs

with a gauge factor of 37.1, hysteresis of 6.3%, linearity of 5.5%, and repeatability of 6.3%.⁹⁰

Wu et al.⁹¹ reported a novel and simplistic approach to fabricate a porous polydimethylsiloxane/CNF composite (p-PDMS/CNFs) and used it as a deformable strain sensor. The 3-D porous nanocomposites of CNFs incorporated in PDMS walls had a significantly higher failure strain ($\sim 94\%$) than PDMS alone ($\sim 48\%$). The gauge factor of the as-prepared sensor was further modified to offer values between 1.0 and 6.5. The composite exhibited constant piezoresistive output with a quick response time and excellent linearity up to $\sim 70\%$. These highly flexible composites have potential as a flexible strain sensor to monitor human joint movements and as conductors that are flexible for wearable electronics because of their tunable sensitivity and conductivity. Recently, Luo et al.⁹² developed PDMS/CNF nanocomposites for sensing applications wherein sugar-coated CNFs were used to create the templates, which were then used to fabricate the nanocomposite structures that were tested under cyclic loads at different functional maximum strains, which were used to investigate the piezoresistive sensing functions. Long-term sensor performance was assessed using 12-h durability tests.⁹² Chowdhury et al.⁹³ reported the piezoresistive sensing functions of two different shapes of polydimethylsiloxane (PDMS)/carbon nanofiber (CNF) composites. The compressive sensors of two different shapes, truncated cone and cylinder, were measured with compression strains as low as 3% and gauge factors of 6.3 and 18.3, respectively. The sensing capability of these materials was tested by independently applying strain showing them to be highly repeatable in 1000 cycles under compression. The synthesized nanocomposites were used as sensor arrays to detect pressure from 35 to 690 kPa. The flexibility and ease of the fabrication could overcome the current gap between the performance and applicability of the flexible sensors. Charara et al.⁹⁴ synthesized compressible and flexible piezoresistive nanocomposites of CNFs and PDMS for sensing applications. A sugar template was used to synthesize porous PDMS having 74.7% porosity. Due to the conductive CNFs network and the porous microstructures of the fabricated nanocomposites, extremely versatile pressure sensors displayed the piezoresistive characteristics of up to 70% compressive strains. CNFs were used as a conductive nanofillers in a polymer matrix as composites for sensor applications. The commonly used polymers for the matrix are conducting polymers, such as polyacetylene, polypyrrole, polyazulene, polyparaphylene, and polyaniline.⁹⁵

3.2. Biosensors. CNFs have been effectively used in various clinical biosensing applications²⁹ due to excellent sensitivity to target molecules, high surface area for adsorption, good electron transferability, and strong electrocatalytic oxidation of the small molecules like viruses, nucleic acids, and proteins. Wang et al.⁹⁶ fabricated TiO_2/C core–shell nanofiber arrays, which were further doped with Ni–Co nanoparticles, and synthesized composite electrodes that exhibit good stability and enhanced sensitivity ($975.3 \mu\text{A} \mu\text{M}^{-1} \text{cm}^{-2}$) for glucose determination in the linear range of 1 μM to 7658 μM with a 0.6 μM detection limit; these are a suitable choice as a glucose sensor.⁹⁶ Li and co-workers⁹⁷ doped carbon nanofibers with different bimetallic MCo (M = Fe, Cu, Mn, and Ni) nanomaterials via electrospinning and thermal treatment that were used to detect glucose non-enzymatically. The CuCo-CNFs showed excellent efficacy for the detection of glucose in human serum samples with good

reproducibility, excellent stability, high sensitivity ($507 \mu\text{A} \mu\text{M}^{-1} \text{cm}^{-2}$) and a linear range of 0.02 to 11 mM. These composite nanofiber electrodes are suitable replacements for traditional enzymatic glucose estimation tests because they are nonenzymatic and economical because of the presence of CNFs.⁹⁷ Similarly, Keerthi et al.⁹⁸ used urchin-like CuS and xanthan gum-derived CNFs to produce a sensor for enzyme-free sensing of glucose. The sensitivity of the synthesized composite was as high as $23.7 \mu\text{A} \mu\text{M}^{-1} \text{cm}^{-2}$. The sensor showed excellent repeatability, durability, and operational stability and a detection limit of $0.03 \mu\text{M}$. Glucose oxidase was immobilized onto a Prussian blue functionalized CNF/glassy carbon electrodes to build a glucose biosensor having good stability, repeatability, and selectivity as a glucose biosensor with a quick response of 5 s, linear range of 0.02 to 12 mM, detection limit of $0.5 \mu\text{M}$, and high sensitivity⁹⁹ of $35.94 \mu\text{A} \mu\text{M}^{-1} \text{cm}^{-2}$.

Guo and co-workers¹⁰⁰ developed TiC–CNF films via a one-step synthesis technique; these were used to evolve a biosensor for glucose detection that showed sensing performance in a linear range of 0.013–10.5 mM with a lower detection limit of $3.7 \mu\text{M}$. TiC–CNFs served as promising substrates for fabricating electrochemical biosensors due to their simple synthetic method and excellent electrochemical efficiency.¹⁰⁰ Liu et al.¹⁰¹ constructed a highly efficacious nonenzymatic glucose sensor by doping NiCo_2O_4 nanoneedle onto electrospun CNFs, which performed over an extensive linear range (0.005–19.175 mM) with a detection limit of $1.5 \mu\text{M}$ and high sensitivity ($1947 \mu\text{A} \mu\text{M}^{-1} \text{cm}^{-2}$).¹⁰¹ Recently, Ding et al.¹⁰² used CuCo_2O_4 decorated CNFs for enzyme-free glucose detection, and the composite was further treated with electro-polymerization of thiophene-3-boronic acid (TBA) and thiophene. The sensor thus obtained showed higher selectivity of 2932 and $708 \mu\text{A} \mu\text{M}^{-1} \text{cm}^{-2}$ with excellent output with linear range of 0.01–0.5 mM and 0.5–1.5 mM, respectively; in addition, it had excellent stability and repeatability.¹⁰²

CNFs were also used to estimate H_2O_2 as it is a vital biological molecule that has an essential role amid numerous physiological activities; it is a reactive oxygen species, so it plays a role in diabetes, cancer, and neurodegenerative diseases.^{103,104} Zhang and co-workers¹⁰⁵ developed a non-enzymatic hydrogen peroxide (H_2O_2) sensor made by carbonization of electrospun polypyrrole nanoparticles embedded in polyacrylonitrile composite nanofibers (NFs). When polypyrrole nanoparticle precursor content was 12 wt %, a significant number of N-doped carbon nanoparticle-embedded carbon nanofibers (NCNPs) were incorporated in the NCNF substrate without any aggregation. The prepared NCNPF composite had a small diameter, high pyrrolic-N content, and higher defective sites, resulting in H_2O_2 reduction with higher electrocatalytic output. NCNPF-12 wt % based H_2O_2 sensor¹⁰⁵ had a quick amperometric response of less than 2 s with $383.9 \text{ A M}^{-1} \text{cm}^{-2}$ sensitivity. Recently, a novel sensor was designed by Riaz et al.¹⁰⁶ for efficient sensing of H_2O_2 on multiple sensor platforms by carbonizing electrospun PAN fibers having a framework of zeolitic imidazole particles (ZIF-67), which were further doped with carbon encompassing cobalt nanoparticles. When the composite was tested using glassy carbon electrodes (GCEs), it showed sensitivity of $300 \mu\text{A mM}^{-1} \text{cm}^{-2}$ with a limit of detection ($10 \mu\text{M}$) that was lower than the approved residual limit of H_2O_2 allowed by the US FDA for food packaging applications; however, its linear sensing range was extended up to 50 mM when used as a free-standing film

electrode. In addition, when integrated with a portable potentiostat, it allowed mobile detection on screen-printed electrodes (SPEs).¹⁰⁶ Laurila et al.¹⁰⁷ have grown CNFs with Pt particles for the detection of H_2O_2 with a fast response time. CNFs from electrospun PAN were first functionalized using carboxylic acid and HNO_3 treatment before they were used to incorporate hydroxyapatite (HA) or Prussian blue (PB) into FCNF-HA and FCNF-PB composites. The FCNF-HA composite was further coated onto a Au electrode, then cytochrome c was immobilized on the synthesized electrode surface. The fabricated biosensor showed vigorous electrocatalytic activity with a quick response to H_2O_2 , having a detection limit of $0.3 \mu\text{M}$. The catalytic current was linear ($2.0 \mu\text{M}$ to 8.7 mM) to H_2O_2 concentration.¹⁰⁸

CNFs have been commonly used to quantify ascorbic acid, uric acid, and particularly levels of dopamine in the human body, which were utilized for an initial diagnosis of Parkinson's disease.^{109–111} Recent advances in this field give us a more accurate, rapid, and economical method of initial diagnosis of the disease. Yue et al.¹¹² fabricated molybdenum disulfide golf ball-like nanosheets deposited onto the CNFs, which were used for the excellent electrochemical dopamine detection in analysis of real urine samples. The as-prepared electrode had a lower detection limit of 36 nM at S/N = 3; their recoveries were shown at different levels of 10, 20, and 40 μM of DA, which was 101.6%, 99.8%, and 107.8% respectively. The synthesized material has outstanding electrochemical properties because of the simultaneous presence of two nanomaterials.¹¹²

Numerous recent studies have targeted the development of improved biosensors for practical applications in cancer treatment. Afzali et al.¹¹³ fabricated a novel voltammetric sensor for pemetrexed (an anticancer drug) detection utilizing a Pd/CNF/methyl trioctyl ammonium bis-(trifluoromethylsulfonyl) imide ionic liquid modified carbon paste electrode via electrospinning method. The authors demonstrated quasi-reversible behavior of the electrode sensor, and square wave voltammetric (SWV) capacities allowed detection of pemetrexed concentration within a linear range of 1.00–35.0 nM having a detection limit of 0.33 nM under the optimized settings. The designed sensor was used for pemetrexed quantification in human plasma from cancer patients and in pharmaceuticals.¹¹³

Several types of research have been pursued on the utilization of CNF-based electrodes for the detection of anticancer drugs. For example, a variety of CNF-based electrospun materials were used for detecting nilutamide in prostate cancer treatment.¹¹⁴ Rajendran et al.¹¹⁴ established an electrochemical sensor for detection of the anti-prostate-cancer drug nilutamide (NLM) in biological samples (spiked serum and urine samples). The authors prepared N-doped CNFs (N-CNFs) by electrospinning, which were further carbonized, and a solvothermal method was used to grow vanadium tetrasulfide nanorods onto the N-CNFs. A glassy carbon electrode (GCE) was modified with the synthesized material and showed a linear range of 0.001–760 μM with a detection limit of 90 pM, along with fast response and excellent stability.

Kokulnathan et al.¹¹⁵ used cerium vanadate doped CNFs (CeV/CNFs) for efficacious catalytic activity to detect nilutamide from spiked human urine. A pure glassy carbon electrode was altered using the synthesized composite and displayed an irreversible reduction peak because of nilutamide reduction to hydroxylamine. The material showed a lower

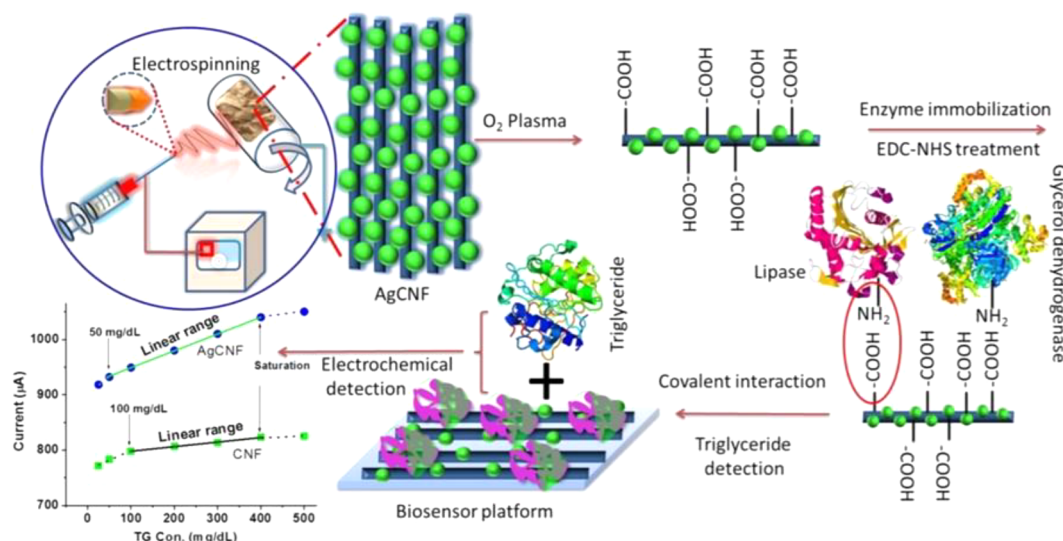


Figure 8. Schematic depiction of the fabrication of CNFs and biosensing for the detection of triglyceride. Reprinted with permission from ref 40. Copyright 2017 Elsevier.

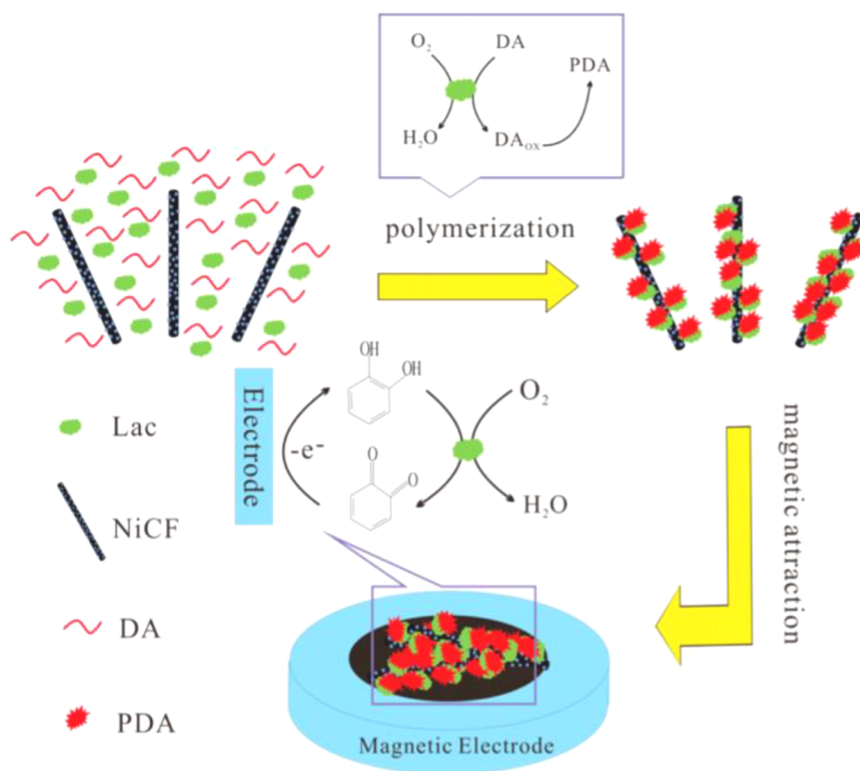


Figure 9. Synthesis scheme of polydopamine–laccase–nickel nanoparticles loaded onto CNFs and sensing application toward catechol. Reprinted with permission from ref 124. Copyright 2014 American Chemical Society.

detection limit of 2.0 nM and higher sensitivity ($1.36 \mu A \mu M^{-1} cm^{-2}$). This method by which drug concentration was precisely mapped in saliva, urine, and plasma is promising. Loaiza et al.¹¹⁶ fabricated a novel lactate biosensor via chemical reduction of Pt precursor onto a graphitized CNF surface. The developed lactate biosensor displayed a detection limit of $6.9 \mu M$ and excellent reproducibility of 4.9%. The biosensor's long-term stability was also assessed, and even after three months of storage at room temperature, 90% of the signal was retained, while retention was 95% after 18 months at $-20^\circ C$.

These findings have shown that the method can produce responsive electrochemical lactate biosensors with long-term enzymatic activity stability.¹¹⁶ Recently, Jeong et al.¹¹⁷ fabricated N-doped CNFs for sensing of cortisol, a hormone regulating glucose level, blood pressure, and carbohydrate metabolism in humans. The synthesized biosensor had a quick response toward cortisol with outstanding sensitivity, reusability, and selectivity with a detection limit as low as 100 aM and linear range of 100 aM to 10 nM.¹¹⁷ Isoaho et al.¹¹⁸ developed a promising biosensor for glutamate detection by

Table 2. Sensing Properties of Different Molecules from CNF-Based Composites

Sensor	Molecule sensed	Linear range	Limit of detection	Ref.
CNFs	dopamine	0.2–700000 μM	0.08 μM	119
CNFs	xanthine	0.03–21.19 μM	20 nM	120
CNFs	tryptophan	0.1–119 μM	0.1 μM	123
	tyrosine	0.2–107 μM		
	cysteine	0.15–64 μM		
CNFs	catechol	1–200 μM	0.2 μM	121
	hydroquinone		0.4 μM	
ZnO/CNFs	dimethyl methylphosphonate	0.1–1000 ppb	0.1 ppb	122
Polydopamine–laccase–nickel NP–CNFs	catechol	1–9100 μM	0.69 μM	124
Co ₃ O ₄ –CNFs	H ₂ O ₂	1–2580 μM	0.5 μM	128
CNFs	H ₂ O ₂	1–10 μM	1.3 μM	126
Ag–Pt/CNFs	dopamine	10–500 μM	0.11 μM	110
PdNPs/CNFs–CPE	dopamine	0.5–160 μM	0.2 μM	127
	uric acid	2–200 μM	0.7 μM	
	ascorbic acid	0.05–4 mM	15 μM	
MCNFs	bisphenol A	10 ⁰ –10 ⁴ fM	1 fM	129
P–CNFs	carbendazim	0.1–35 $\mu\text{mol L}^{-1}$	0.038 $\mu\text{mol L}^{-1}$	130
MoS ₂ –CNF	vanillin	0.3–135 μM	0.15 μM	131
MoS ₂ NSA/CNFs	levodopa	1–60 μM	1 μM	132
	uric acid			
Modified CNFs	Cd (II)	2–100 ppb	0.11 ppb	133
FeCo–CNFs	L–cysteine	1–20 μM	0.15 μM	134
ZnO–CNFs	H ₂ S	102–50 ppm		135
CoMnZIF@CNFs	α -synuclein	52.6 fM to 0.1 nM	45.7 fM	136
M–CNFs	I ₂	5–700 μM	0.59 μM (in PBS) 1.41 μM (in synthetic urine)	79

growing CNFs onto a Pt catalyst; detection was done in O₂ containing PBS at -0.15 and 0.6 V vs Ag/AgCl potentials. There were two linear ranges for glutamate when measured amperometrically at -0.15 V vs Ag/AgCl. The first one is between 1 μM and 100 μM with sensitivity of 0.266 $\mu\text{A } \mu\text{M}^{-1} \text{ cm}^{-2}$ and the second is between 100 μM and 1000 μM with a sensitivity of 0.0208 $\mu\text{A } \mu\text{M}^{-1} \text{ cm}^{-2}$. Mondal et al.⁴⁰ synthesized a biosensor for triglyceride detection (Figure 8) using electrospinning followed by carbonization and impregnation of Ag NPs in CNFs. The sensitivity of this biosensor (1.232 $\mu\text{A (mg/dL)}^{-1} \text{ cm}^{-2}$) was four-times higher than that of CNFs alone (0.33 $\mu\text{A (mg/dL)}^{-1} \text{ cm}^{-2}$). Fast response in 10 s along with good selectivity and excellent reproducibility were observed.

3.3. Sensing of Small Molecules. Various studies have used CNF-based materials for small molecule detection. For example, Mao et al.¹¹⁹ utilized electrospun CNFs for the detection of dopamine, which showed outstanding selectivity, sensitivity, and stability; its LOD was 0.08 μM (S/N ratio 3). Tang et al.¹²⁰ used sensitive nonenzymatic CNF-based sensors, which were synthesized via electrospinning and successive thermal treatment, for the detection of xanthine (Xa). A dynamic linear range for xanthine of 0.03 – 21.9 μM ($R = 0.9992$) with LOD of 20 nM was found. Guo et al.¹²¹ used electrospun CNF modified carbon paste electrodes for sensing catechol (CC) and hydroquinone (HQ) using cyclic voltammetry and differential pulse voltammetry in real samples of lake water. The results showed that CC and HQ could be detected sensitively and selectively in the presence of 50 μM isomers with a detection limit of 0.2 and 0.4 μM , respectively, and a linear range of 1 – 200 μM . Lee et al.¹²² synthesized hybrid metal oxide composite CNFs by an electrospinning method as a sensor material at room temperature for the

detection of dimethyl methyl phosphonate (DMMP). By attachment of a water suspension of CNFs from electrospun polyacrylonitrile (PAN) onto the surface of a carbon paste electrode (CNF–CPE), a CNF–CPE sensor was established by Tang et al.¹²³ Using constant potential amperometry and cyclic voltammetry, the CNF–CPE electrode could detect the amino acids L-tryptophan (Trp), L-tyrosine (Tyr), and L-cysteine (Cys) without any further treatment. The CNF–CPE showed brilliant electrocatalytic activity and analytical efficiency for amino acid oxidation. Trp, Tyr, and Cys displayed linear ranges of 0.1 – 119 , 0.2 – 107 , and 0.15 – 64 μM , respectively, with LOD of 0.1 μM (S/N = 3) for all the analytes. In addition, the sensor demonstrated good reproducibility, stability, and selectivity.

Li et al.¹²⁴ synthesized a phenolic biosensor by fabricating a composite of polydopamine, laccase, and nickel nanoparticles loaded onto CNFs as shown in Figure 9. NiCNFs are a type of 1-D magnetic material having a large specific surface area and several Ni particles, which could offer ample active sites for laccase immobilization. The fabricated composite was used for sensing of catechol in real water samples with a sensitivity of 25 $\mu\text{A } \mu\text{M}^{-1} \text{ cm}^{-2}$. It also exhibited good selectivity and stability having a lower LOD of 0.69 μM (S/N = 3). Table 2 summarizes the properties of CNF-based composites for the detection of several molecules.

Ni et al.¹²⁵ synthesized an amperometric nonenzymatic sensor for H₂O₂ detection by doping Co₃O₄ NPs onto CNFs; the resulting sensor was highly selective in the presence of electroactive interferents. Oxidation current was linear over the range of 1 to 2580 μM H₂O₂ concentration, but best was calculated at 0.2 V (vs SCE), which was used to determine H₂O₂ in spiked milk samples. The sensing of H₂O₂ was accomplished by Mao et al.¹²⁶ using electrospun CNF webs.

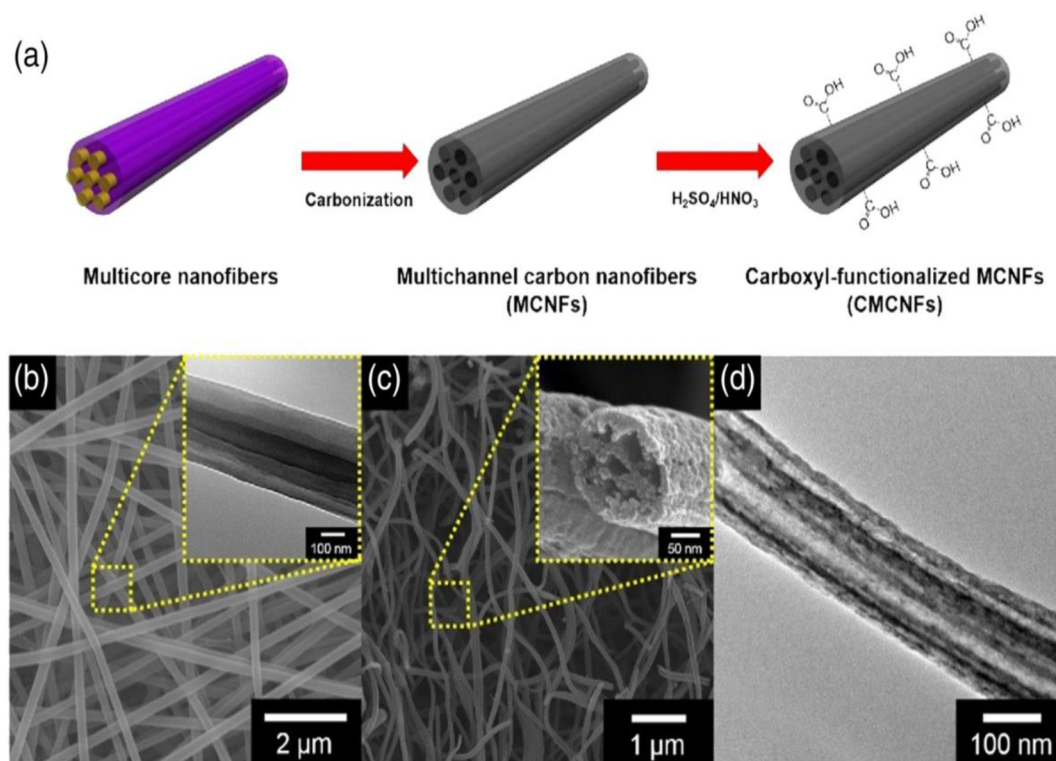


Figure 10. (a) Synthesis scheme for carboxyl-functionalized multichannel CNFs. (b) Multicore NFs are visible through FE-SEM and TEM micrographs. (c) FE-SEM micrographs showing multichannel CNFs. (d) TEM of carboxyl-functionalized multichannel CNFs. Reprinted with permission from ref 129. Copyright 2016 American Chemical Society.

They were made by manipulating the concentration of nanosized graphite domains by changing the carbonization conditions. The response to H_2O_2 concentration was linear in the range of 1–10 μM with sensitivity of 31200 $\mu\text{A mM}^{-1}$. Wang et al.¹²⁷ synthesized Pd nanoparticle loaded CNFs by electrospinning PAN with palladium acetate and then thermally treating to coat a carbon paste electrode (CPE). The PdNP/CNF-CPE demonstrated outstanding electrochemical catalytic activities against dopamine (DA), uric acid (UA), and ascorbic acid (AA) with LOD of DA, AA, and UA of 0.2, 0.7, and 15 μM , respectively. Electrospinning of polyacrylonitrile (PAN) with rhodium acetate accompanied by carbonization yielded RhNP-loaded CNFs (RhNP/CNFs), which were dispersed in DMF using ultrasonic agitation before casting onto the surface of pyrolytic graphite electrodes that were pretreated. The sensor showed excellent electrocatalytic activity against hydrazine oxidation and was used for hydrazine sensing, since it was highly sensitive and selective.

3.4. Detection of Food Preservatives. Bisphenol A is utilized commonly in applications for food packaging, since its structure resembles that of estrogen hormone and it has an endocrine-disrupting impact. Kim et al.¹²⁹ developed a transistor sensor, created on aptamer modified multichannel-CNFs, with carboxyl groups on the surface of the CNFs, immobilized on an amine-functionalized electrode (Figure 10). The synthesized sensor showed excellent selectivity and sensitivity toward bisphenol A at a comparatively low concentration (1 fM). The prepared sensor was stable and was reused for repetitive assays.

Cui et al.¹³⁰ prepared a 3-D hierarchical helical structure of phosphorus-doped CNFs fabricated by a CVD method and high-pressure annealing, which exhibited a rapid electron

transfer. This sensor had high sensitivity and electrocatalytic activity, making it a suitable choice for carbendazim detection in food samples; the LOD was 0.038 $\mu\text{mol L}^{-1}$ (S/N = 3). The sensors' storage stability was also investigated, and only a slight current decrease ($\sim 2.8\%$) was observed after the electrode was stored for 15 days at room temperature. N-doped CNFs fabricated by nozzle-less electrospinning were used for electrochemical sensing of carbendazim.¹³⁷ This sensor has good stability and high sensitivity for the detection of carbendazim in 0.1 mol/L PBS. The studies showed the linear relation with regression equation $I_p = 0.891C + 3.37$ and a detection limit of 0.053 (S/N = 3). Sundaresan et al. prepared chitosan–CNF composite supported Cu nanoparticles using a sonication approach and used them for an electrochemical sensing platform for carbendazim. The prepared sensor exhibited excellent selectivity with linear range of 0.8–277.0 μM and detection limit of 0.028 μM . They also tested this sensor for analysis of carbendazim in real water washed soil samples.¹³⁸

3.5. Detection of Disease and Toxicity Biomarkers. Periyakaruppan et al. detected ricin, a toxic glycoprotein extracted from castor bean seeds, using aptamer and antibody probes on a nanoelectrode array containing vertically aligned CNFs.¹⁴⁰ The authors also developed a label-free biosensor using a CNF nanoelectrode to detect cardiac troponin-I (cTnI) in the initial evaluation of myocardial infarction.¹³⁹ The developed immunosensor showed excellent sensitivity in response to the human-cTnI analyte, detecting it at low concentration (~ 0.2 ng/mL), which was 25-times lower than what the traditional methods could detect. It displayed a wide linear range of 0.25–1 and 5–100 mg/mL.¹³⁹ Gupta and co-workers¹⁴¹ designed a CNF-based sensor for the detection of

C-reactive protein, which acts as a biomarker for cardiac disease; the sensor had a detection limit of ~ 90 pM.

Phenolic compounds are toxic and can cause cancer and loss of immunity. Li et al.¹⁴² fabricated Ni–Cu nanoparticle-doped CNFs through electrospinning and carbonization. The synthesized composite was mixed with laccase and Nafion to develop a novel sensor. It showed promising electrocatalytic effects for the detection of hydroquinone at $1.5 \mu\text{A } \mu\text{M}^{-1}$ with a LOD of 90 nM ($S/N = 3$). The sensor achieved hydroquinone detection in lake water.¹⁴² Yin et al.¹⁴³ synthesized a novel phenolic sensor for hydroquinone (HQ), catechol (CC), and resorcinol (RS) detection using Co–Fe selenides doped in a porous CNF modified glassy carbon electrode. The synthesized composite showed 3-D nanostructure, and the electrode showed wide linear ranges and good LOD values of $0.5\text{--}200 \mu\text{M}$ and $0.13 \mu\text{M}$, $0.5\text{--}190 \mu\text{M}$ and $0.15 \mu\text{M}$, $5\text{--}350 \mu\text{M}$ and $1.36 \mu\text{M}$ for HQ, CC, and RS, respectively. The synthesized phenolic sensor had excellent stability and selectivity.¹⁴³ The ability of the sensor to detect HQ, CC, and RS was also demonstrated by increasing the concentration of the three analytes at the same time. The linearity for HQ, CC, and RS was $0.5\text{--}180 \mu\text{M}$, $1\text{--}160 \mu\text{M}$, and $10\text{--}120 \mu\text{M}$, respectively, with the LOD values of 0.53 ($R^2 = 0.998$), 0.84 ($R^2 = 0.998$), and 2.62 ($R^2 = 0.995$).¹⁴³

3.6. Gas Sensors. Since CNFs are considered to be sensitive to gas exposure, they could be used to make a variety of gas sensors for NO_2 ,^{144,145} NH_3 ,¹⁴⁵ H_2S ,¹³⁵ H_2 ,¹⁴⁶ and $\text{C}_2\text{H}_5\text{OH}$.¹⁴⁷ The construction of a NO_2 sensor utilizing 2-D WS_2 functionalized multichannel CNFs was proposed by Cha et al.¹⁴⁴ as depicted in Figure 11a. A NO_2 sensor with a 15% response, 4–0.1 ppm concentration range, and gas detection limit of 1 ppm was successfully developed by combining WS_2 , a promising gas sensor material, with CNFs. Further inves-

tigation of the sensing mechanism using CNFs and WS_2 -doped multichannel CNFs revealed that several aspects, including elongated pore channels, high open porosity, and ample gas spots on the edge sites of WS_2 , have synergistic effects on enhancing the sensing efficiency (Figure 11b).

H_2S is a flammable and neurotoxic typical atmospheric contaminant for which Zhang and co-workers¹³⁵ fabricated a stable sensor with ZnO–CNFs. When compared to pure ZnO nanofibers, the synthesized sensor had an excellent selectivity and response, particularly at 102–50 ppm of H_2S . Furthermore, over 60 days, these sensors showed nearly constant response of $\sim 40\text{--}20$ ppm of H_2S . The superior performance of the sensors was credited to the carbon protection, which ensured higher ZnO stability, as well as oxygen vacancies, which enhanced its response and selectivity to H_2S .

Wang and co-workers¹⁴⁶ fabricated hierarchical p–n junction nanostructures of SnO_2 nanosheets (n-type) doped onto CNFs (p-type) by the combination of electrospinning and hydrothermal methods. The efficacy for gas sensing of the hierarchical nanostructured material with hydrogen as a target molecule was investigated. Due to the synergistic effect between SnO_2 nanosheets, CNFs, and the sensor, the predicted sensing outputs (large response, low operating temperature, and quick response recovery behavior) were achieved; in 100 ppm of H_2 , the gas sensor had a ~ 4 s response time and less than 16 s recovery time.¹⁴⁶

Yan and Wu¹⁴⁷ synthesized Sn– SnO_2 /carbon heterostructure NFs using electrospinning and heat treatment and used them for selective sensing of $\text{C}_2\text{H}_5\text{OH}$; the heterostructured fibers showed enhanced sensing efficacy compared to pure SnO_2 , along with a linear dependence of response on the $\text{C}_2\text{H}_5\text{OH}$ concentration (10–100 ppm) with brilliant selectivity and recovery due to high surface area and pore structures offered by the 1-D CNFs as well as the unique sensing mechanism by the hybrid Sn– SnO_2 /CNFs.¹⁴⁷ Lee et al.¹⁴⁸ fabricated WO_3 nanonodule–CNFs employing single nozzle co-electrospinning for the detection of NO_2 . The composite had a larger sensing surface area ($147\text{--}276 \text{ m}^2 \text{ g}^{-1}$), and WO_3 on the material's surface combined with O_2 of NO_2 , thereby allowing NO_2 detection at room temperature (rt) within a detection limit of 1 ppm. Monereo et al.¹⁴⁵ studied the application of self-heating effects in larger arrays of randomly focused CNFs for gas sensors for NO_2 and NH_3 . Similarly, Claramunt et al.¹⁴⁹ used CNFs decorated with metal NPs on Kapton for NH_3 detection.

3.7. Human Locomotion Monitoring. Persistent human motion tracking with the help of CNFs can help physicians and therapists build tailored treatment programs for individuals suffering from disorders like multiple sclerosis, Parkinson's disease, and strokes. Sengupta et al.¹⁵⁰ prepared a stitchable, wearable carbon nanofiber–polydimethylsiloxane (CNFs–PDMS) based piezoelectric sensor. The developed sensor exhibited force sensitivity of $\sim 1.82 \text{ kN}^{-1}$ with a linear response measured through electromechanical tactile sensing for a normal force up to 20 N. The functionality of the as-prepared sensor was tested through gesture monitoring and human motion detection via different experiments of wrist movement, walking, and jogging. Lu et al.¹⁵¹ prepared a PAN-based CNF and carbon black sensor for the detection of human motion. These sensors exhibited a linear range of 0.1–20%, could detect small strain of 0.1%, and showed a strain sensing maximum of 100%. Furthermore, quick response time (67 ms) and exceptional long-term durability (continuous stretching

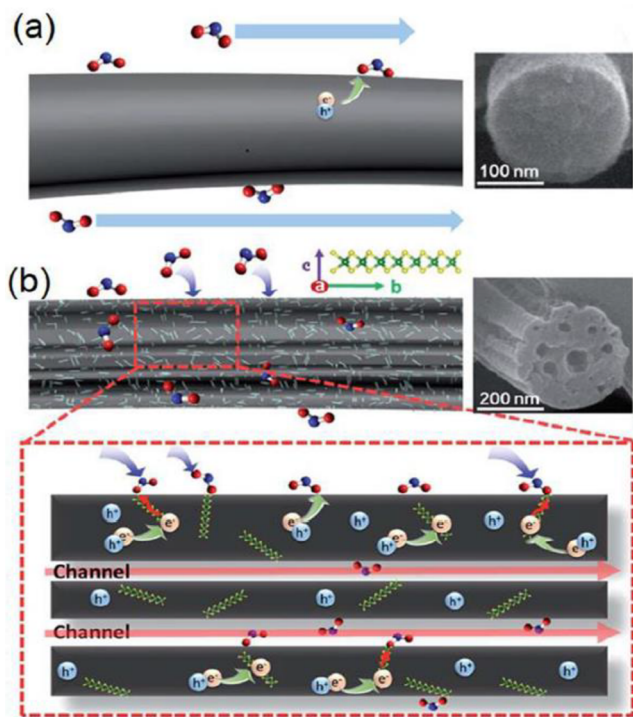


Figure 11. Representation of the sensing process for (a) CNFs and (b) WS_2 @MTCNFs for sensing of NO_2 . Reprinted with permission from ref 144. Copyright 2017 Royal Society of Chemistry.

time of 7500 s) showed that the film sensor was capable of detecting human motion in real time. Cheng and co-workers¹⁵² fabricated a compressible and stretchable hydrogel strain sensor using CNF powder and poly(vinyl alcohol) (PVA) by freezing–thawing cycles. The sensor exhibited low plastic deformation (10%) and little energy loss efficiency of 5.62% and 12.13% for stretching and compressing, respectively. The authors used it for detecting multiple stretching behaviors of humans like joint bending, breathing, or swallowing.

3.8. Energy Applications. Advanced energy conversion and storage systems such as supercapacitors and lithium-ion batteries are urgently needed to meet energy needs and to support new applications like electric vehicles and portable electronics. CNFs can be used in supercapacitors owing to their high specific surface area. For supercapacitor applications, a new porous electrode material (graphene-doped CNFs) was utilized by Zhou and Wu.¹⁵³ They prepared a supercapacitor by electrospinning and carbonization method. In 6 M KOH, the developed material showed specific capacitance of 263.7 F/g and cycling stability for energy storage with 86.9% retention ratio. Kim et al.¹⁵⁴ used a mesoporous CNF web for high-performance supercapacitor applications. The developed carbonized fiber showed a high surface area of $\sim 500 \text{ m}^2 \text{ g}^{-1}$. The observed specific capacitance was 128 F/g along with energy density of 16.0–21.4 Wh kg^{-1} and 75.0–58.2 Wh kg^{-1} in aqueous and organic electrolyte, respectively. Jung et al.¹⁵⁵ used CNFs for supercapacitors prepared from poly-(acrylonitrile-co-vinylimidazole) via electrospinning and thermal treatment. The capacitor showed specific capacitance ranging between 122 F/g and 86 F/g; power density and maximum energy were calculated from the Ragone plot and were 7.2 kW/kg and 47.4 Wh/kg, respectively. Lithium-ion batteries are effective power sources for electronic devices. CNFs are excellent anode and cathode materials to use in lithium-ion batteries. Zeng et al.¹⁵⁶ prepared N-doped CNF–Si/C composites by carbonization and electrospinning technique. The developed composite increased the electrochemical properties of lithium-ion battery anodes. At 0.2 A/g, it showed reversible capacity of 945.5 mAh g^{-1} with 64% capacity retention for 150 cycles, and at 0.5 A/g, it showed reversible capacity of 538.6 mAh g^{-1} for 500 cycles. $\text{LiMn}_2\text{O}_4/\text{C}/\text{CNFs}$ were used as a cathode material in lithium ion batteries with electrochemical performance of 126 mAh g^{-1} at 0.2 C at a capacity of 92 mAh g^{-1} after 1000 cycles¹⁵⁷ at 1 C. Other researchers have also used CNFs for applications in lithium-ion batteries.^{20,158,159}

4. CHALLENGES AND OBSTACLES FOR CARBON NANOFIBERS

Despite the positive outcomes derived from using CNFs as a sensor, future risks must be addressed. CNFs are indeed promising in biomedical applications as biosensors. Still, limited information on CNF toxicity, cytocompatibility, and dispersibility has been the foremost hurdle preventing them from being used as effective biomaterials or for clinical applications. Major challenges with CNFs are the dispersion of the composite in the matrix material, orientation and alignment, interface, and densification of the composite processing.¹⁶⁰ Inhalation toxicity was the focus of the majority of toxicity studies. They have the unique capacity for inducing proliferation of mesenchymal cells, granuloma formation, and fibrogenesis.¹⁶¹ However, surface area, chemical structure, purity, agglomeration propensity, and functionalization are

some of the factors that can influence and regulate CNF toxicity. Furthermore, the concentration of CNFs utilized in toxicity studies is commonly specified in $\mu\text{g}/\text{mL}$, which is considerably higher than the actual concentrations used in clinical settings. Thereby toxicity levels get unreasonably high.

Studies have shown that CNFs with nonfunctionalized hydrophobic composition and heavy metal impurities are more cytotoxic,¹⁶² while many researchers have a different opinion concerning the functionalization of the CNFs with carbonyl, hydroxyl, or carboxyl groups that can result in higher toxicity than the unfunctionalized ones.¹⁶³ Larger physical sizes (length, diameter) have led to increased cytotoxicity compared to thinner and smaller materials.¹⁶⁴ To obtain unswerving toxicity results of CNFs, in-depth investigation and protocols are urgently needed to integrate research efforts on safety, administration evaluations, and exposure.

5. CONCLUSION AND FUTURE PERSPECTIVE

Based on the facts discussed here on the CNF-based materials, it can be visualized that they play a vital part in fabricating different types of sensors for various applications. The different synthesis routes along with challenges and obstacles of CNFs have been emphasized. Among all, electrospinning is the most prevalent route for synthesizing CNFs. Heteroatom-doping of CNFs with other materials, such as metals, nitrogen, nanoparticles, or polymers, has improved the sensing efficacy. The overall success of such studies has immense consequences for fundamental science, technological explorations, and further tailoring of CNFs. The high aspect ratio and large surface area facilitate adsorption of a vast number of target molecules. More active sites facilitate high electron transfer contributing to their extensive range of applications in chemical sensing. This work comprehensively reviewed the considerable role of CNFs in various sensing applications.

More efforts and focus should be put into uncovering and exploring the possible benefits of implementing CNFs. Finally, the lack of validated data on CNF toxicity should prompt researchers to compile the data set needed for risk assessment. For example, standard tests and chronic animal bioassays may be used to carefully mimic real-world situations for systemic, local, and occupational implementation of CNFs.

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Notes

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REFERENCES

- (1) Mishra, A.; Shetti, N. P.; Basu, S.; Raghava Reddy, K.; Aminabhavi, T. M. Carbon Cloth-based Hybrid Materials as Flexible Electrochemical Supercapacitors. *ChemElectroChem* **2019**, *6* (23), 5771–5786.
- (2) Mishra, A.; Mehta, A.; Basu, S.; Shetti, N. P.; Reddy, K. R.; Aminabhavi, T. M. Graphitic Carbon Nitride (g-C₃N₄)-Based Metal-Free Photocatalysts for Water Splitting: A Review. *Carbon N. Y.* **2019**, *149*, 693–721.
- (3) Reddy, K. R.; Reddy, C. H. V.; Nadagouda, M. N.; Shetti, N. P.; Jaesool, S.; Aminabhavi, T. M. Polymeric Graphitic Carbon Nitride (g-C₃N₄)-Based Semiconducting Nanostructured Materials: Synthesis Methods, Properties and Photocatalytic Applications. *J. Environ. Manage.* **2019**, *238*, 25–40.
- (4) Mishra, A.; Basu, S.; Shetti, N. P.; Reddy, K. R.; Aminabhavi, T. M. Photocatalysis of Graphene and Carbon Nitride-Based Functional Carbon Quantum Dots. *Nanoscale Materials in Water Purification*; Elsevier, 2019; pp 759–781.
- (5) Kundu, A.; Sharma, S.; Basu, S. Modulated BiOCl Nanoplates with Porous G-C₃N₄ Nanosheets for Photocatalytic Degradation of Color/Colorless Pollutants in Natural Sunlight. *J. Phys. Chem. Solids* **2021**, *154*, 110064.
- (6) Mehta, A.; Mishra, A.; Basu, S.; Shetti, N. P.; Reddy, K. R.; Saleh, T. A.; Aminabhavi, T. M. Band Gap Tuning and Surface Modification of Carbon Dots for Sustainable Environmental Remediation and Photocatalytic Hydrogen Production-A Review. *J. Environ. Manage.* **2019**, *250*, 109486.
- (7) Singla, S.; Sharma, S.; Basu, S.; Shetti, N. P.; Reddy, K. R. Graphene/Graphitic Carbon Nitride-Based Ternary Nanohybrids: Synthesis Methods, Properties, and Applications for Photocatalytic Hydrogen Production. *FlatChem* **2020**, *24*, 100200.
- (8) Garg, A.; Basu, S.; Shetti, N. P.; Reddy, K. R. 2D Materials and Its Heterostructured Photocatalysts: Synthesis, Properties, Functionalization and Applications in Environmental Remediation. *J. Environ. Chem. Eng.* **2021**, *9*, 106408.
- (9) Bukkittgar, S. D.; Shetti, N. P. Electrochemical Behavior of an Anticancer Drug 5-Fluorouracil at Methylene Blue Modified Carbon Paste Electrode. *Mater. Sci. Eng., C* **2016**, *65*, 262–268.
- (10) Shetti, N. P.; Nayak, D. S.; Malode, S. J.; Kakarla, R. R.; Shukla, S. S.; Aminabhavi, T. M. Sensors Based on Ruthenium-Doped TiO₂ Nanoparticles Loaded into Multi-Walled Carbon Nanotubes for the Detection of Flufenamic Acid and Mefenamic Acid. *Anal. Chim. Acta* **2019**, *1051*, 58–72.
- (11) Shetti, N. P.; Malode, S. J.; Nayak, D. S.; Bagihalli, G. B.; Reddy, K. R.; Ravindranadh, K.; Reddy, C. V. A Novel Biosensor Based on Graphene Oxide-Nanoclay Hybrid Electrode for the Detection of Theophylline for Healthcare Applications. *Microchem. J.* **2019**, *149*, 103985.
- (12) Monga, D.; Basu, S. Enhanced Photocatalytic Degradation of Industrial Dye by G-C₃N₄/TiO₂ Nanocomposite: Role of Shape of TiO₂. *Adv. Powder Technol.* **2019**, *30* (5), 1089–1098.
- (13) Monga, D.; Ilager, D.; Shetti, N. P.; Basu, S.; Aminabhavi, T. M. 2D/2d Heterojunction of MoS₂/g-C₃N₄ Nanoflowers for Enhanced Visible-Light-Driven Photocatalytic and Electrochemical Degradation of Organic Pollutants. *J. Environ. Manage.* **2020**, *274*, 111208.
- (14) Nagar, A.; Basu, S. Ternary G-C₃N₄/Ag/BiVO₄ Nanocomposite: Fabrication and Implementation to Remove Organic Pollutants. *Environ. Technol. Innov.* **2021**, *23*, 101646.
- (15) Kundu, A.; Maity, B.; Basu, S. Coal-Derived Graphene Quantum Dots with a Mn²⁺/Mn⁷⁺ Nanosensor for Selective Detection of Glutathione by a Fluorescence Switch-off-on Assay. *New J. Chem.* **2022**, *46* (16), 7545–7556.
- (16) Sohal, N.; Maity, B.; Basu, S. Carbon Dot-MnO₂ Nanosphere Composite Sensors for Selective Detection of Glutathione. *ACS Appl. Nano Mater.* **2020**, *3* (6), 5955–5964.
- (17) Shetti, N. P.; Nayak, D. S.; Malode, S. J.; Kulkarni, R. M.; Kulkarni, D. B.; Teggi, R. A.; Joshi, V. V. Electrooxidation and Determination of Flufenamic Acid at Graphene Oxide Modified Carbon Electrode. *Surfaces and Interfaces* **2017**, *9*, 107–113.
- (18) Shetti, N. P.; Mishra, A.; Basu, S.; Mascarenhas, R. J.; Kakarla, R. R.; Aminabhavi, T. M. Skin-Patchable Electrodes for Biosensor Applications: A Review. *ACS Biomater. Sci. Eng.* **2020**, *6* (4), 1823–1835.
- (19) Shetti, N. P.; Mishra, A.; Basu, S.; Aminabhavi, T. M. Versatile Fullerenes as Sensor Materials. *Mater. Today Chem.* **2021**, *20*, 100454.
- (20) Mishra, A.; Mehta, A.; Basu, S.; Malode, S. J.; Shetti, N. P.; Shukla, S. S.; Nadagouda, M. N.; Aminabhavi, T. M. Electrode Materials for Lithium-Ion Batteries. *Mater. Sci. Energy Technol.* **2018**, *1* (2), 182–187.
- (21) Mishra, A.; Shetti, N. P.; Basu, S.; Reddy, K. R.; Aminabhavi, T. M. Recent Developments in Ionic Liquid-Based Electrolytes for Energy Storage Supercapacitors and Rechargeable Batteries. *Green Sustainable Process for Chemical and Environmental Engineering and Science: Ionic Liquids as Green Solvents*; Elsevier, 2019; pp 199–221, DOI: 10.1016/B978-0-12-817386-2.00007-X.
- (22) Moon, S. C.; Farris, R. J. Strong Electrospun Nanometer-Diameter Polyacrylonitrile Carbon Fiber Yarns. *Carbon N. Y.* **2009**, *47* (12), 2829–2839.
- (23) Ruiz-Cornejo, J. C.; Sebastián, D.; Lázaro, M. J. Synthesis and Applications of Carbon Nanofibers: A Review. *Rev. Chem. Eng.* **2020**, *36* (4), 493–511.
- (24) Mohamed, A. Synthesis, Characterization, and Applications Carbon Nanofibers. In *Carbon-based nanofillers and their rubber nanocomposites*; Elsevier, 2019; pp 243–257.
- (25) Din, I. U.; Shaharun, M. S.; Naem, A.; Alotaibi, M. A.; Alharthi, A. I.; Bakht, M. A.; Nasir, Q. Carbon Nanofibers as Potential Materials for Catalysts Support, a Mini-Review on Recent Advances and Future Perspective. *Ceram. Int.* **2020**, *46* (11), 18446–18452.
- (26) Chen, I. H.; Wang, C. C.; Chen, C. Y. Fabrication and Structural Characterization of Polyacrylonitrile and Carbon Nanofibers Containing Plasma-Modified Carbon Nanotubes by Electrospinning. *J. Phys. Chem. C* **2010**, *114* (32), 13532–13539.

- (27) De Jong, K. P.; Geus, J. W. Carbon Nanofibers: Catalytic Synthesis and Applications. *Catal. Rev. - Sci. Eng.* **2000**, *42* (4), 481–510.
- (28) Rauti, R.; Musto, M.; Bosi, S.; Prato, M.; Ballerini, L. Properties and Behavior of Carbon Nanomaterials When Interfacing Neuronal Cells: How Far Have We Come? *Carbon N. Y.* **2019**, *143*, 430–446.
- (29) Wang, Z.; Wu, S.; Wang, J.; Yu, A.; Wei, G. Carbon Nanofiber-Based Functional Nanomaterials for Sensor Applications. *Nanomaterials* **2019**, *9* (7), 1045.
- (30) Chen, L. F.; Lu, Y.; Yu, L.; Lou, X. W. Designed Formation of Hollow Particle-Based Nitrogen-Doped Carbon Nanofibers for High-Performance Supercapacitors. *Energy Environ. Sci.* **2017**, *10* (8), 1777–1783.
- (31) Li, Y.; Zhang, M.; Zhang, X.; Xie, G.; Su, Z.; Wei, G. Nanoporous Carbon Nanofibers Decorated with Platinum Nanoparticles for Non-Enzymatic Electrochemical Sensing of H₂O₂. *Nanomaterials* **2015**, *5* (4), 1891–1905.
- (32) Chen, L. F.; Feng, Y.; Liang, H. W.; Wu, Z. Y.; Yu, S. H. Macroscopic-Scale Three-Dimensional Carbon Nanofiber Architectures for Electrochemical Energy Storage Devices. *Adv. Energy Mater.* **2017**, *7* (23), 1700826.
- (33) Mukkabl, R.; Kuldeep; Deepa, M. Poly (Carbazole)-Coated Selenium@ Conical Carbon Nanofibers Hybrid for Lithium-Selenium Batteries with Enhanced Lifespan. *ACS Appl. Energy Mater.* **2018**, *1* (12), 6964–6976.
- (34) Liu, Y.; Zhou, J.; Chen, L.; Zhang, P.; Fu, W.; Zhao, H.; Ma, Y.; Pan, X.; Zhang, Z.; Han, W.; Xie, E. Highly Flexible Freestanding Porous Carbon Nanofibers for Electrodes Materials of High-Performance All-Carbon Supercapacitors. *ACS Appl. Mater. Interfaces* **2015**, *7* (42), 23515–23520.
- (35) Sharma, S.; Basu, S.; Shetti, N. P.; Mondal, K.; Sharma, A.; Aminabhavi, T. M. Versatile Graphitized Carbon Nanofibers in Energy Applications. *ACS Sustain. Chem. Eng.* **2022**, *10* (4), 1334–1360.
- (36) Cai, T.; Huang, Y.; Huang, M.; Xi, Y.; Pang, D.; Zhang, W. Enhancing Oxygen Reduction Reaction of Supercapacitor Microbial Fuel Cells with Electrospun Carbon Nanofibers Composite Cathode. *Chem. Eng. J.* **2019**, *371*, 544–553.
- (37) Massaglia, G.; Margaria, V.; Sacco, A.; Castellino, M.; Chiodoni, A.; Pirri, F. C.; Quaglio, M. N-Doped Carbon Nanofibers as Catalyst Layer at Cathode in Single Chamber Microbial Fuel Cells. *Int. J. Hydrogen Energy* **2019**, *44* (9), 4442–4449.
- (38) Zhao, Y.; Liang, J.; Wang, C.; Ma, J.; Wallace, G. G. Tunable and Efficient Tin Modified Nitrogen-doped Carbon Nanofibers for Electrochemical Reduction of Aqueous Carbon Dioxide. *Adv. Energy Mater.* **2018**, *8* (10), 1702524.
- (39) Ali, M. A.; Mondal, K.; Jiao, Y.; Oren, S.; Xu, Z.; Sharma, A.; Dong, L. Microfluidic Immuno-Biochip for Detection of Breast Cancer Biomarkers Using Hierarchical Composite of Porous Graphene and Titanium Dioxide Nanofibers. *ACS Appl. Mater. Interfaces* **2016**, *8* (32), 20570–20582.
- (40) Mondal, K.; Ali, M. A.; Singh, C.; Sumana, G.; Malhotra, B. D.; Sharma, A. Highly Sensitive Porous Carbon and Metal/Carbon Conducting Nanofiber Based Enzymatic Biosensors for Triglyceride Detection. *Sensors Actuators, B Chem.* **2017**, *246*, 202–214.
- (41) Inagaki, M.; Yang, Y.; Kang, F. Carbon Nanofibers Prepared via Electrospinning. *Adv. Mater.* **2012**, *24* (19), 2547–2566.
- (42) Mondal, K.; Bhattacharyya, S.; Sharma, A. Photocatalytic Degradation of Naphthalene by Electrospun Mesoporous Carbon-Doped Anatase TiO₂ Nanofiber Mats. *Ind. Eng. Chem. Res.* **2014**, *53* (49), 18900–18909.
- (43) Mondal, K.; Sharma, A. Recent Advances in Electrospun Metal-Oxide Nanofiber Based Interfaces for Electrochemical Biosensing. *RSC Adv.* **2016**, *6* (97), 94595–94616.
- (44) Zhang, L.; Aboagye, A.; Kelkar, A.; Lai, C.; Fong, H. A Review: Carbon Nanofibers from Electrospun Polyacrylonitrile and Their Applications. *J. Mater. Sci.* **2014**, *49* (2), 463–480.
- (45) Su, Z.; Ding, J.; Wei, G. Electrospinning: A Facile Technique for Fabricating Polymeric Nanofibers Doped with Carbon Nanotubes and Metallic Nanoparticles for Sensor Applications. *Rsc Adv.* **2014**, *4* (94), 52598–52610.
- (46) Maitra, T.; Sharma, S.; Srivastava, A.; Cho, Y. K.; Madou, M.; Sharma, A. Improved Graphitization and Electrical Conductivity of Suspended Carbon Nanofibers Derived from Carbon Nanotube/ Polyacrylonitrile Composites by Directed Electrospinning. *Carbon N. Y.* **2012**, *50* (5), 1753–1761.
- (47) Jadhav, S. A.; Dhavale, S. B.; Patil, A. H.; Patil, P. S. Brief Overview of Electrospun Polyacrylonitrile Carbon Nanofibers: Preparation Process with Applications and Recent Trends. *Mater. Design Process. Commun.* **2019**, *1* (5), No. e83.
- (48) Ruiz-Rosas, R.; Bedia, J.; Lallave, M.; Loscertales, I. G.; Barrero, A.; Rodríguez-Mirasol, J.; Cordero, T. The Production of Submicron Diameter Carbon Fibers by the Electrospinning of Lignin. *Carbon N. Y.* **2010**, *48* (3), 696–705.
- (49) Singh, P.; Mondal, K.; Sharma, A. Reusable Electrospun Mesoporous ZnO Nanofiber Mats for Photocatalytic Degradation of Polycyclic Aromatic Hydrocarbon Dyes in Wastewater. *J. Colloid Interface Sci.* **2013**, *394* (1), 208–215.
- (50) Feng, L.; Xie, N.; Zhong, J. Carbon Nanofibers and Their Composites: A Review of Synthesizing, Properties and Applications. *Materials (Basel)*. **2014**, *7* (5), 3919–3945.
- (51) Zahid, M. U.; Pervaiz, E.; Hussain, A.; Shahzad, M. I.; Niazi, M. B. K. Synthesis of Carbon Nanomaterials from Different Pyrolysis Techniques: A Review. *Mater. Res. Express* **2018**, *5* (5), 052002.
- (52) Hulicova-Jurcakova, D.; Li, X.; Zhu, Z.; de Marco, R.; Lu, G. Q. Graphitic Carbon Nanofibers Synthesized by the Chemical Vapor Deposition (CVD) Method and Their Electrochemical Performances in Supercapacitors. *Energy Fuels* **2008**, *22* (6), 4139–4145.
- (53) VM, S.; Mohamed, A. R.; Abdullah, A. Z.; Chai, S.-P. Role of Reaction and Factors of Carbon Nanotubes Growth in Chemical Vapour Decomposition Process Using Methane—a Highlight. *J. Nanomater.* **2010**, *2010*, 395191.
- (54) Guan, H.; Zhang, J.; Liu, Y.; Zhao, Y.; Zhang, B. Rapid Quantitative Determination of Hydrogen Peroxide Using an Electrochemical Sensor Based on PtNi Alloy/CeO₂ Plates Embedded in N-Doped Carbon Nanofibers. *Electrochim. Acta* **2019**, *295*, 997–1005.
- (55) Bao, J.; Kishi, N.; Khatri, I.; Soga, T.; Jimbo, T. Catalyst-Free Synthesis of Carbon Nanofibers by Ultrasonic Spray Pyrolysis of Ethanol. *Mater. Lett.* **2012**, *68*, 240–242.
- (56) Lim, S.; Shimizu, A.; Yoon, S. H.; Korai, Y.; Mochida, I. High Yield Preparation of Tubular Carbon Nanofibers over Supported Co-Mo Catalysts. *Carbon N. Y.* **2004**, *42* (7), 1279–1283.
- (57) García-Bordejé, E.; Kvande, I.; Chen, D.; Rønning, M. Synthesis of Composite Materials of Carbon Nanofibers and Ceramic Monoliths with Uniform and Tuneable Nanofibre Layer Thickness. *Carbon N. Y.* **2007**, *45* (9), 1828–1838.
- (58) Bauman, Y. I.; Mishakov, I. V.; Rudneva, Y. V.; Plyusnin, P. E.; Shubin, Y. V.; Korneev, D. V.; Vedyagin, A. A. Formation of Active Sites of Carbon Nanofibers Growth in Self-Organizing Ni-Pd Catalyst during Hydrogen-Assisted Decomposition of 1, 2-Dichloroethane. *Ind. Eng. Chem. Res.* **2019**, *58* (2), 685–694.
- (59) Saidin, M. A. R.; Ismail, A. F.; Sanip, S. M.; Goh, P. S.; Aziz, M.; Tanemura, M. Controlled Growth of Carbon Nanofibers Using Plasma Enhanced Chemical Vapor Deposition: Effect of Catalyst Thickness and Gas Ratio. *Thin Solid Films* **2012**, *520* (7), 2575–2581.
- (60) Shoukat, R.; Khan, M. I. Synthesis of Vertically Aligned Carbon Nanofibers Using Inductively Coupled Plasma-Enhanced Chemical Vapor Deposition. *Electr. Eng.* **2018**, *100* (2), 997–1002.
- (61) Gupta, R.; Sharma, S. C. Modelling the Effects of Nitrogen Doping on the Carbon Nanofiber Growth via Catalytic Plasma-enhanced Chemical Vapour Deposition Process. *Contrib. to Plasma Phys.* **2019**, *59* (1), 72–85.
- (62) Gupta, K.; Jain, N. K.; Laubscher, R. Advances in Gear Manufacturing. *Adv. Gear Manuf. Finish.* **2017**, 67–125.
- (63) Wang, Y.; Zheng, M.; Lu, H.; Feng, S.; Ji, G.; Cao, J. Template Synthesis of Carbon Nanofibers Containing Linear Mesocage Arrays. *Nanoscale Res. Lett.* **2010**, *5* (6), 913–916.

- (64) Xing, Y.; Fang, B.; Bonakdarpour, A.; Zhang, S.; Wilkinson, D. Facile Fabrication of Mesoporous Carbon Nanofibers with Unique Hierarchical Nanoarchitecture for Electrochemical Hydrogen Storage. *Int. J. Hydrogen Energy* **2014**, *39* (15), 7859–7867.
- (65) Fan, Z.; Zhang, H. Template Synthesis of Noble Metal Nanocrystals with Unusual Crystal Structures and Their Catalytic Applications. *Acc. Chem. Res.* **2016**, *49* (12), 2841–2850.
- (66) Zheng, M.; Ji, G.; Wang, Y.; Cao, J.; Feng, S.; Liao, L.; Du, Q.; Zhang, L.; Ling, Z.; Liu, J.; et al. A New Restriction Effect of Hard Templates for the Shrinkage of Mesoporous Polymer during Carbonization. *Chem. Commun.* **2009**, No. 33, 5033–5035.
- (67) Chen, C. S.; Lin, J. H.; You, J. H.; Yang, K. H. Effects of Potassium on Ni-K/Al₂O₃ Catalysts in the Synthesis of Carbon Nanofibers by Catalytic Hydrogenation of CO₂. *J. Phys. Chem. A* **2010**, *114* (11), 3773–3781.
- (68) Ren, J.; Li, F. F.; Lau, J.; González-Urbina, L.; Licht, S. One-Pot Synthesis of Carbon Nanofibers from CO₂. *Nano Lett.* **2015**, *15* (9), 6142–6148.
- (69) Gupta, V. K.; Agarwal, S.; Tyagi, I.; Sohrabi, M.; Fakhri, A.; Rashidi, S.; Sadeghi, N. Microwave-Assisted Hydrothermal Synthesis and Adsorption Properties of Carbon Nanofibers for Methamphetamine Removal from Aqueous Solution Using a Response Surface Methodology. *J. Ind. Eng. Chem.* **2016**, *41*, 158–164.
- (70) Klein, K. L.; Melechko, A. V.; McKnight, T. E.; Retterer, S. T.; Rack, P. D.; Fowlkes, J. D.; Joy, D. C.; Simpson, M. L. Surface Characterization and Functionalization of Carbon Nanofibers. *J. Appl. Phys.* **2008**, *103* (6), 061301.
- (71) Melechko, A. V.; Klein, K. L.; Fowlkes, J. D.; Hensley, D. K.; Merkulov, I. A.; McKnight, T. E.; Rack, P. D.; Horton, J. A.; Simpson, M. L. Control of Carbon Nanostructure: From Nanofiber toward Nanotube and Back. *J. Appl. Phys.* **2007**, *102* (7), 074314.
- (72) Baker, S. E.; Tse, K.-Y.; Lee, C.-S.; Hamers, R. J. Fabrication and Characterization of Vertically Aligned Carbon Nanofiber Electrodes for Biosensing Applications. *Diam. Relat. Mater.* **2006**, *15* (2–3), 433–439.
- (73) Zhou, J.-H.; Sui, Z.-J.; Zhu, J.; Li, P.; Chen, D.; Dai, Y.-C.; Yuan, W.-K. Characterization of Surface Oxygen Complexes on Carbon Nanofibers by TPD, XPS and FT-IR. *Carbon N. Y.* **2007**, *45* (4), 785–796.
- (74) Moon, J. S.; Alegaonkar, P. S.; Han, J. H.; Lee, T. Y.; Yoo, J. B.; Kim, J. M. Enhanced Field Emission Properties of Thin-Multiwalled Carbon Nanotubes: Role of Si O_x Coating. *J. Appl. Phys.* **2006**, *100* (10), 104303.
- (75) He, P.; Shi, D.; Lian, J.; Wang, L. M.; Ewing, R. C.; van Ooij, W.; Li, W. Z.; Ren, Z. F. Plasma Deposition of Thin Carbonfluorine Films on Aligned Carbon Nanotube. *Appl. Phys. Lett.* **2005**, *86* (4), 043107.
- (76) Dhindsa, M. S.; Smith, N. R.; Heikenfeld, J.; Rack, P. D.; Fowlkes, J. D.; Doktycz, M. J.; Melechko, A. V.; Simpson, M. L. Reversible Electrowetting of Vertically Aligned Superhydrophobic Carbon Nanofibers. *Langmuir* **2006**, *22* (21), 9030–9034.
- (77) Guillorn, M. A.; McKnight, T. E.; Melechko, A.; Merkulov, V. I.; Britt, P. F.; Austin, D. W.; Lowndes, D. H.; Simpson, M. L. Individually Addressable Vertically Aligned Carbon Nanofiber-Based Electrochemical Probes. *J. Appl. Phys.* **2002**, *91* (6), 3824–3828.
- (78) Naguib, N. N.; Mueller, Y. M.; Bojczuk, P. M.; Rossi, M. P.; Katsikis, P. D.; Gogotsi, Y. Effect of Carbon Nanofiber Structure on the Binding of Antibodies. *Nanotechnology* **2005**, *16* (4), 567.
- (79) Cho, E.; Perebikovskiy, A.; Benice, O.; Holmberg, S.; Madou, M.; Ghazinejad, M. Rapid Iodine Sensing on Mechanically Treated Carbon Nanofibers. *Sensors* **2018**, *18* (5), 1486.
- (80) Adabi, M.; Saber, R.; Faridi-Majidi, R.; Faridbod, F. Performance of Electrodes Synthesized with Polyacrylonitrile-Based Carbon Nanofibers for Application in Electrochemical Sensors and Biosensors. *Mater. Sci. Eng., C* **2015**, *48*, 673–678.
- (81) Galao, O.; Baeza, F. J.; Zornoza, E.; Garcés, P. Strain and Damage Sensing Properties on Multifunctional Cement Composites with CNF Admixture. *Cem. Concr. Compos.* **2014**, *46*, 90–98.
- (82) Wu, Z.; Li, C.; Liang, H.; Chen, J.; Yu, S. Ultralight, Flexible, and Fire-resistant Carbon Nanofiber Aerogels from Bacterial Cellulose. *Angew. Chem.* **2013**, *125* (10), 2997–3001.
- (83) Yan, T.; Wang, Z.; Wang, Y. Q.; Pan, Z. J. Carbon/Graphene Composite Nanofiber Yarns for Highly Sensitive Strain Sensors. *Mater. Des.* **2018**, *143*, 214–223.
- (84) Mordkovich, V. Z. Carbon Nanofibers: A New Ultrahigh-Strength Material for Chemical Technology. *Theor. Found. Chem. Eng.* **2003**, *37* (5), 429–438.
- (85) Coleman, J. N.; Khan, U.; Gun'ko, Y. K. Mechanical Reinforcement of Polymers Using Carbon Nanotubes. *Adv. Mater.* **2006**, *18* (6), 689–706.
- (86) Zhu, J.; Wei, S.; Ryu, J.; Guo, Z. Strain-Sensing Elastomer/Carbon Nanofiber “Metacomposites”. *J. Phys. Chem. C* **2011**, *115* (27), 13215–13222.
- (87) Hu, N.; Itoi, T.; Akagi, T.; Kojima, T.; Xue, J.; Yan, C.; Atobe, S.; Fukunaga, H.; Yuan, W.; Ning, H.; Surina; Liu, Y.; Alamusi. Ultrasensitive Strain Sensors Made from Metal-Coated Carbon Nanofiller/Epoxy Composites. *Carbon N. Y.* **2013**, *51* (1), 202–212.
- (88) Azhari, F.; Banthia, N. Cement-Based Sensors with Carbon Fibers and Carbon Nanotubes for Piezoresistive Sensing. *Cem. Concr. Compos.* **2012**, *34* (7), 866–873.
- (89) Baeza, F. J.; Galao, O.; Zornoza, E.; Garcés, P. Multifunctional Cement Composites Strain and Damage Sensors Applied on Reinforced Concrete (RC) Structural Elements. *Materials (Basel)*. **2013**, *6* (3), 841–855.
- (90) Wang, Y.; Wang, Y.; Han, B.; Wan, B.; Cai, G.; Li, Z. Strain Monitoring of Concrete Components Using Embedded Carbon Nanofibers/Epoxy Sensors. *Constr. Build. Mater.* **2018**, *186*, 367–378.
- (91) Wu, S.; Zhang, J.; Ladani, R. B.; Ravindran, A. R.; Mouritz, A. P.; Kinloch, A. J.; Wang, C. H. Novel Electrically Conductive Porous PDMS/Carbon Nanofiber Composites for Deformable Strain Sensors and Conductors. *ACS Appl. Mater. Interfaces* **2017**, *9* (16), 14207–14215.
- (92) Luo, W.; Charara, M.; Saha, M. C.; Liu, Y. Fabrication and Characterization of Porous CNF/PDMS Nanocomposites for Sensing Applications. *Appl. Nanosci.* **2019**, *9* (6), 1309–1317.
- (93) Chowdhury, S. A.; Saha, M. C.; Patterson, S.; Robison, T.; Liu, Y. Highly Conductive Polydimethylsiloxane/Carbon Nanofiber Composites for Flexible Sensor Applications. *Adv. Mater. Technol.* **2019**, *4* (1), 1800398.
- (94) Charara, M.; Luo, W.; Saha, M. C.; Liu, Y. Investigation of Lightweight and Flexible Carbon Nanofiber/Poly Dimethylsiloxane Nanocomposite Sponge for Piezoresistive Sensor Application. *Adv. Eng. Mater.* **2019**, *21* (5), 1801068.
- (95) Naveen, M. H.; Gurudatt, N. G.; Shim, Y.-B. Applications of Conducting Polymer Composites to Electrochemical Sensors: A Review. *Appl. Mater. today* **2017**, *9*, 419–433.
- (96) Wang, J.; Liu, Y.; Cheng, L.; Chen, R.; Ni, H. Quasi-Aligned Nanorod Arrays Composed of Nickel-Cobalt Nanoparticles Anchored on TiO₂/C Nanofiber Arrays as Free Standing Electrode for Enzymeless Glucose Sensors. *J. Alloys Compd.* **2020**, *821*, 153510.
- (97) Li, M.; Liu, L.; Xiong, Y.; Liu, X.; Nsabimana, A.; Bo, X.; Guo, L. Bimetallic MCo (M = Cu, Fe, Ni, and Mn) Nanoparticles Doped-Carbon Nanofibers Synthesized by Electrosynthesis for Nonenzymatic Glucose Detection. *Sensors Actuators B Chem.* **2015**, *207*, 614–622.
- (98) Keerthi, M.; Mutharani, B.; Chen, S.-M.; Ranganathan, P. Carbon Fibers Coated with Urchin-like Copper Sulfide for Non-enzymatic Voltammetric Sensing of Glucose. *Microchim. Acta* **2019**, *186* (12), 807.
- (99) Wang, L.; Ye, Y.; Zhu, H.; Song, Y.; He, S.; Xu, F.; Hou, H. Controllable Growth of Prussian Blue Nanostructures on Carboxylic Group-Functionalized Carbon Nanofibers and Its Application for Glucose Biosensing. *Nanotechnology* **2012**, *23* (45), 455502.
- (100) Guo, Q.; Liu, L.; Wu, T.; Wang, Q.; Wang, H.; Liang, J.; Chen, S. Flexible and Conductive Titanium Carbide-Carbon Nanofibers for High-Performance Glucose Biosensing. *Electrochim. Acta* **2018**, *281*, 517–524.

- (101) Liu, L.; Wang, Z.; Yang, J.; Liu, G.; Li, J.; Guo, L.; Chen, S.; Guo, Q. NiCo₂O₄ Nanoneedle-Decorated Electrospun Carbon Nanofiber Nanohybrids for Sensitive Non-Enzymatic Glucose Sensors. *Sensors Actuators, B Chem.* **2018**, 258, 920–928.
- (102) Ding, Y.; Sun, H.; Ren, C.; Zhang, M.; Sun, K. A Nonenzymatic Glucose Sensor Platform Based on Specific Recognition and Conductive Polymer-Decorated CuCO₂O₄ Carbon Nanofibers. *Materials (Basel)*. **2020**, 13 (12), 2874.
- (103) Weinstain, R.; Savariar, E. N.; Felsen, C. N.; Tsien, R. Y. In Vivo Targeting of Hydrogen Peroxide by Activatable Cell-Penetrating Peptides. *J. Am. Chem. Soc.* **2014**, 136 (3), 874–877.
- (104) Rahman, A.; Pallichankandy, S.; Thayyullathil, F.; Galadari, S. Critical Role of H₂O₂ in Mediating Sanguinarine-Induced Apoptosis in Prostate Cancer Cells via Facilitating Ceramide Generation, ERK1/2 Phosphorylation, and Par-4 Cleavage. *Free Radic. Biol. Med.* **2019**, 134, 527–544.
- (105) Zhang, X.; Liu, D.; Yu, B.; You, T. A Novel Nonenzymatic Hydrogen Peroxide Sensor Based on Electrospun Nitrogen-Doped Carbon Nanoparticles-Embedded Carbon Nanofibers Film. *Sensors Actuators B Chem.* **2016**, 224, 103–109.
- (106) Riaz, M. A.; Yuan, Z.; Mahmood, A.; Liu, F.; Sui, X.; Chen, J.; Huang, Q.; Liao, X.; Wei, L.; Chen, Y. Hierarchically Porous Carbon Nanofibers Embedded with Cobalt Nanoparticles for Efficient H₂O₂ Detection on Multiple Sensor Platforms. *Sensors Actuators, B Chem.* **2020**, 319, 128243.
- (107) Laurila, T.; Sainio, S.; Jiang, H.; Isoaho, N.; Koehne, J. E.; Etula, J.; Koskinen, J.; Meyyappan, M. Application-Specific Catalyst Layers: Pt-Containing Carbon Nanofibers for Hydrogen Peroxide Detection. *ACS omega* **2017**, 2 (2), 496–507.
- (108) Cui, K.; Song, Y.; Guo, Q.; Xu, F.; Zhang, Y.; Shi, Y.; Wang, L.; Hou, H.; Li, Z. Architecture of Electrospun Carbon Nanofibers-Hydroxyapatite Composite and Its Application Act as a Platform in Biosensing. *Sensors Actuators B Chem.* **2011**, 160 (1), 435–440.
- (109) Habibi, B.; Jahanbakhshi, M.; Pournaghi-Azar, M. H. Simultaneous Determination of Acetaminophen and Dopamine Using SWCNT Modified Carbon-Ceramic Electrode by Differential Pulse Voltammetry. *Electrochim. Acta* **2011**, 56 (7), 2888–2894.
- (110) Huang, Y.; Miao, Y.-E.; Ji, S.; Tjiu, W. W.; Liu, T. Electrospun Carbon Nanofibers Decorated with Ag-Pt Bimetallic Nanoparticles for Selective Detection of Dopamine. *ACS Appl. Mater. Interfaces* **2014**, 6 (15), 12449–12456.
- (111) Yue, Y.; Hu, G.; Zheng, M.; Guo, Y.; Cao, J.; Shao, S. A Mesoporous Carbon Nanofiber-Modified Pyrolytic Graphite Electrode Used for the Simultaneous Determination of Dopamine, Uric Acid, and Ascorbic Acid. *Carbon N. Y.* **2012**, 50 (1), 107–114.
- (112) Yue, H. Y.; Wu, P. F.; Huang, S.; Wang, Z. Z.; Gao, X.; Song, S. S.; Wang, W. Q.; Zhang, H. J.; Guo, X. R. Golf Ball-like MoS₂ Nanosheet Arrays Anchored onto Carbon Nanofibers for Electrochemical Detection of Dopamine. *Microchim. Acta* **2019**, 186 (6), 378.
- (113) Afzali, M.; Mostafavi, A.; Nekooie, R.; Jahromi, Z. A Novel Voltammetric Sensor Based on Palladium Nanoparticles/Carbon Nanofibers/Ionic Liquid Modified Carbon Paste Electrode for Sensitive Determination of Anti-Cancer Drug Pemetrexed. *J. Mol. Liq.* **2019**, 282, 456–465.
- (114) Rajendran, K.; Kokulnathan, T.; Chen, S.-M.; Allen, J. A.; Viswanathan, C.; Therese, H. A. Nitrogen Doped Carbon Nanofibers Loaded with Hierarchical Vanadium Tetrasulfide for the Voltammetric Detection of the Non-Steroidal Anti-Prostate Cancer Drug Nilutamide. *Microchim. Acta* **2019**, 186 (3), 141.
- (115) Kokulnathan, T.; Karthik, R.; Chen, S.-M.; Kumar, J. V.; Sakthiathan, S. A Cerium Vanadate Interconnected with a Carbon Nanofiber Heterostructure for Electrochemical Determination of the Prostate Cancer Drug Nilutamide. *Microchim. Acta* **2019**, 186 (8), 579.
- (116) Loaiza, O. A.; Lamas-Ardisana, P. J.; Añorga, L.; Jubete, E.; Ruiz, V.; Borghei, M.; Cabañero, G.; Grande, H. J. Graphitized Carbon Nanofiber-Pt Nanoparticle Hybrids as Sensitive Tool for Preparation of Screen Printing Biosensors. Detection of Lactate in Wines and Ciders. *Bioelectrochemistry* **2015**, 101, 58–65.
- (117) Jeong, G.; Oh, J.; Jang, J. Fabrication of N-Doped Multidimensional Carbon Nanofibers for High-Performance Cortisol Biosensors. *Bioelectron.* **2019**, 131, 30–36.
- (118) Isoaho, N.; Peltola, E.; Sainio, S.; Koskinen, J.; Laurila, T. Pt-Grown Carbon Nanofibers for Enzymatic Glutamate Biosensors and Assessment of Their Biocompatibility. *RSC Adv.* **2018**, 8 (62), 35802–35812.
- (119) Mao, X.; Yang, X.; Rutledge, G. C.; Alan Hatton, T. Ultra-Wide-Range Electrochemical Sensing Using Continuous Electrospun Carbon Nanofibers with High Densities of States. *ACS Appl. Mater. Interfaces* **2014**, 6 (5), 3394–3405.
- (120) Tang, X.; Liu, Y.; Hou, H.; You, T. A Nonenzymatic Sensor for Xanthine Based on Electrospun Carbon Nanofibers Modified Electrode. *Talanta* **2011**, 83 (5), 1410–1414.
- (121) Guo, Q.; Huang, J.; Chen, P.; Liu, Y.; Hou, H.; You, T. Simultaneous Determination of Catechol and Hydroquinone Using Electrospun Carbon Nanofibers Modified Electrode. *Sensors Actuators B Chem.* **2012**, 163 (1), 179–185.
- (122) Lee, J. S.; Kwon, O. S.; Park, S. J.; Park, E. Y.; You, S. A.; Yoon, H.; Jang, J. Fabrication of Ultrafine Metal-Oxide-Decorated Carbon Nanofibers for DMMP Sensor Application. *ACS Nano* **2011**, 5 (10), 7992–8001.
- (123) Tang, X.; Liu, Y.; Hou, H.; You, T. Electrochemical Determination of L-Tryptophan, L-Tyrosine and L-Cysteine Using Electrospun Carbon Nanofibers Modified Electrode. *Talanta* **2010**, 80 (5), 2182–2186.
- (124) Li, D.; Luo, L.; Pang, Z.; Ding, L.; Wang, Q.; Ke, H.; Huang, F.; Wei, Q. Novel Phenolic Biosensor Based on a Magnetic Polydopamine-Laccase-Nickel Nanoparticle Loaded Carbon Nanofiber Composite. *ACS Appl. Mater. Interfaces* **2014**, 6 (7), 5144–5151.
- (125) Ni, Y.; Liao, Y.; Zheng, M.; Shao, S. In-Situ Growth of Co₃O₄ Nanoparticles on Mesoporous Carbon Nanofibers: A New Nanocomposite for Nonenzymatic Amperometric Sensing of H₂O₂. *Microchim. Acta* **2017**, 184 (10), 3689–3695.
- (126) Mao, X.; Simeon, F.; Rutledge, G. C.; Hatton, T. A. Electrospun Carbon Nanofiber Webs with Controlled Density of States for Sensor Applications. *Adv. Mater.* **2013**, 25 (9), 1309–1314.
- (127) Wang, X.; Wu, M.; Tang, W.; Zhu, Y.; Wang, L.; Wang, Q.; He, P.; Fang, Y. Simultaneous Electrochemical Determination of Ascorbic Acid, Dopamine and Uric Acid Using a Palladium Nanoparticle/Graphene/Chitosan Modified Electrode. *J. Electroanal. Chem.* **2013**, 695 (4), 10–16.
- (128) Ni, Y.; Liao, Y.; Zheng, M.; Shao, S. In-Situ Growth of Co₃O₄ Nanoparticles on Mesoporous Carbon Nanofibers: A New Nanocomposite for Nonenzymatic Amperometric Sensing of H₂O₂. *Microchim. Acta* **2017**, 184 (10), 3689–3695.
- (129) Kim, S. G.; Lee, J. S.; Jun, J.; Shin, D. H.; Jang, J. Ultrasensitive Bisphenol A Field-Effect Transistor Sensor Using an Aptamer-Modified Multichannel Carbon Nanofiber Transducer. *ACS Appl. Mater. Interfaces* **2016**, 8 (10), 6602–6610.
- (130) Cui, R.; Xu, D.; Xie, X.; Yi, Y.; Quan, Y.; Zhou, M.; Gong, J.; Han, Z.; Zhang, G. Phosphorus-Doped Helical Carbon Nanofibers as Enhanced Sensing Platform for Electrochemical Detection of Carbendazim. *Food Chem.* **2017**, 221, 457–463.
- (131) Qianwen, M.; Yaping, D.; Li, L.; Anqing, W.; Dingding, D.; Yijun, Z. Electrospun MoS₂ Composite Carbon Nanofibers for Determination of Vanillin. *J. Electroanal. Chem.* **2019**, 833, 297–303.
- (132) Wang, W. Q.; Yue, H. Y.; Yu, Z. M.; Huang, S.; Song, S. S.; Gao, X.; Guan, E. H.; Zhang, H. J.; Wang, Z. Synthesis and Application of MoS₂ Nanosheet Arrays/Carbon Nanofibers for Simultaneous Electrochemical Determination of Levodopa and Uric Acid. *IEEE Sens. J.* **2019**, 19 (15), 5988–5994.
- (133) Fakude, C. T.; Arotiba, O. A.; Arduini, F.; Mabuba, N. Flexible Polyester Screen-printed Electrode Modified with Carbon Nanofibers for the Electrochemical Aptasensing of Cadmium (II). *Electroanalysis* **2020**, 32 (12), 2650–2658.
- (134) Yang, Z.; Zhu, Y.; Nie, G.; Li, M.; Wang, C.; Lu, X. FeCo Nanoparticles-Embedded Carbon Nanofibers as Robust Peroxidase

Mimics for Sensitive Colorimetric Detection of L-Cysteine. *Dalt. Trans.* **2017**, 46 (28), 8942–8949.

(135) Zhang, J.; Zhu, Z.; Chen, C.; Chen, Z.; Cai, M.; Qu, B.; Wang, T.; Zhang, M. ZnO-Carbon Nanofibers for Stable, High Response, and Selective H₂S Sensors. *Nanotechnology* **2018**, 29 (27), 275501.

(136) Guo, C.; Hu, M.; Li, Z.; Duan, F.; He, L.; Zhang, Z.; Marchetti, F.; Du, M. Structural Hybridization of Bimetallic Zeolitic Imidazolate Framework (ZIF) Nanosheets and Carbon Nanofibers for Efficiently Sensing α -Synuclein Oligomers. *Sensors Actuators B Chem.* **2020**, 309, 127821.

(137) Yang, P.; Pan, X.; Wang, J.; Yang, R.; Chu, J.; Chen, H.; Nan, H.; Yang, L.; Zhao, X. Nozzle-less Electrospun Nitrogen-doped Hollow Carbon Nanofibers as Enhanced Sensing Platform for Carbendazim Electrochemical Detection. *ChemistrySelect* **2019**, 4 (7), 2059–2063.

(138) Sundaresan, P.; Fu, C.-C.; Liu, S.-H.; Juang, R.-S. Facile Synthesis of Chitosan-Carbon Nanofiber Composite Supported Copper Nanoparticles for Electrochemical Sensing of Carbendazim. *Colloids Surfaces A Physicochem. Eng. Asp.* **2021**, 625, 126934.

(139) Periyakaruppan, A.; Gandhiraman, R. P.; Meyyappan, M.; Koehne, J. E. Label-Free Detection of Cardiac Troponin-I Using Carbon Nanofiber Based Nanoelectrode Arrays. *Anal. Chem.* **2013**, 85 (8), 3858–3863.

(140) Periyakaruppan, A.; Arumugam, P. U.; Meyyappan, M.; Koehne, J. E. Detection of Ricin Using a Carbon Nanofiber Based Biosensor. *Biosens. Bioelectron.* **2011**, 28 (1), 428–433.

(141) Gupta, R. K.; Periyakaruppan, A.; Meyyappan, M.; Koehne, J. E. Label-Free Detection of C-Reactive Protein Using a Carbon Nanofiber Based Biosensor. *Biosens. Bioelectron.* **2014**, 59, 112–119.

(142) Li, D.; Lv, P.; Zhu, J.; Lu, Y.; Chen, C.; Zhang, X.; Wei, Q. NiCu Alloy Nanoparticle-Loaded Carbon Nanofibers for Phenolic Biosensor Applications. *Sensors* **2015**, 15 (11), 29419–29433.

(143) Yin, D.; Liu, J.; Bo, X.; Guo, L. Cobalt-Iron Selenides Embedded in Porous Carbon Nanofibers for Simultaneous Electrochemical Detection of Trace of Hydroquinone, Catechol and Resorcinol. *Anal. Chim. Acta* **2020**, 1093, 35–42.

(144) Cha, J.-H.; Choi, S.-J.; Yu, S.; Kim, I.-D. 2D WS₂-Edge Functionalized Multi-Channel Carbon Nanofibers: Effect of WS₂ Edge-Abundant Structure on Room Temperature NO₂ Sensing. *J. Mater. Chem. A* **2017**, 5 (18), 8725–8732.

(145) Monereo, O.; Prades, J. D.; Cirera, A. Self-Heating Effects in Large Arrangements of Randomly Oriented Carbon Nanofibers: Application to Gas Sensors. *Sensors Actuators, B Chem.* **2015**, 211, 489–497.

(146) Wang, Z.; Liu, S.; Jiang, T.; Xu, X.; Zhang, J.; An, C.; Wang, C. N-Type SnO₂ Nanosheets Standing on p-Type Carbon Nanofibers: A Novel Hierarchical Nanostructures Based Hydrogen Sensor. *RSC Adv.* **2015**, 5 (79), 64582–64587.

(147) Yan, S.; Wu, Q. Micropored Sn-SnO₂/Carbon Heterostructure Nanofibers and Their Highly Sensitive and Selective C₂H₅OH Gas Sensing Performance. *Sensors Actuators B Chem.* **2014**, 205, 329–337.

(148) Lee, J. S.; Kwon, O. S.; Shin, D. H.; Jang, J. WO₃ Nanonodule-Decorated Hybrid Carbon Nanofibers for NO₂ Gas Sensor Application. *J. Mater. Chem. A* **2013**, 1 (32), 9099–9106.

(149) Claramunt, S.; Monereo, O.; Boix, M.; Leghrib, R.; Prades, J. D.; Cornet, A.; Merino, P.; Merino, C.; Cirera, A. Flexible Gas Sensor Array with an Embedded Heater Based on Metal Decorated Carbon Nanofibers. *Sensors Actuators, B Chem.* **2013**, 187, 401–406.

(150) Sengupta, D.; Romano, J.; Kottapalli, A. G. P. Electrospun Bundled Carbon Nanofibers for Skin-Inspired Tactile Sensing, Proprioception and Gesture Tracking Applications. *npj Flex. Electron.* **2021**, 5 (1), 29.

(151) Lu, X.; Qin, Y.; Chen, X.; Peng, C.; Yang, Y.; Zeng, Y. An Ultra-Wide Sensing Range Film Strain Sensor Based on a Branch-Shaped PAN-Based Carbon Nanofiber and Carbon Black Synergistic Conductive Network for Human Motion Detection and Human-Machine Interfaces. *J. Mater. Chem. C* **2022**, 10 (16), 6296–6305.

(152) Cheng, B.; Chang, S.; Li, H.; Li, Y.; Shen, W.; Shang, Y.; Cao, A. Highly Stretchable and Compressible Carbon Nanofiber-Polymer Hydrogel Strain Sensor for Human Motion Detection. *Macromol. Mater. Eng.* **2020**, 305 (3), 1900813.

(153) Zhou, Z.; Wu, X. F. Graphene-Beaded Carbon Nanofibers for Use in Supercapacitor Electrodes: Synthesis and Electrochemical Characterization. *J. Power Sources* **2013**, 222, 410–416.

(154) Kim, B.-H.; Yang, K. S.; Ferraris, J. P. Highly Conductive, Mesoporous Carbon Nanofiber Web as Electrode Material for High-Performance Supercapacitors. *Electrochim. Acta* **2012**, 75, 325–331.

(155) Jung, K.-H.; Deng, W.; Smith, D. W., Jr.; Ferraris, J. P. Carbon Nanofiber Electrodes for Supercapacitors Derived from New Precursor Polymer: Poly (Acrylonitrile-Co-Vinylimidazole). *Electrochem. commun.* **2012**, 23, 149–152.

(156) Zeng, Y.; Huang, Y.; Liu, N.; Wang, X.; Zhang, Y.; Guo, Y.; Wu, H.-H.; Chen, H.; Tang, X.; Zhang, Q. N-Doped Porous Carbon Nanofibers Sheathed Pumpkin-like Si/C Composites as Free-Standing Anodes for Lithium-Ion Batteries. *J. Energy Chem.* **2021**, 54, 727–735.

(157) Yu, X.; Deng, J.; Yang, X.; Li, J.; Huang, Z. H.; Li, B.; Kang, F. A Dual-Carbon-Anchoring Strategy to Fabricate Flexible LiMn₂O₄ Cathode for Advanced Lithium-Ion Batteries with High Areal Capacity. *Nano Energy* **2020**, 67, 104256.

(158) Zhao, X.; Jia, W.; Wu, X.; Lv, Y.; Qiu, J.; Guo, J.; Wang, X.; Jia, D.; Yan, J.; Wu, D. Ultrafine MoO₃ Anchored in Coal-Based Carbon Nanofibers as Anode for Advanced Lithium-Ion Batteries. *Carbon N. Y.* **2020**, 156, 445–452.

(159) Gao, M.; Liu, B.; Zhang, X.; Zhang, Y.; Li, X.; Han, G. Ultrathin MoS₂ Nanosheets Anchored on Carbon Nanofibers as Free-Standing Flexible Anode with Stable Lithium Storage Performance. *J. Alloys Compd.* **2022**, 894, 162550.

(160) Neubauer, E.; Kitzmantel, M.; Hulman, M.; Angerer, P. Potential and Challenges of Metal-Matrix-Composites Reinforced with Carbon Nanofibers and Carbon Nanotubes. *Compos. Sci. Technol.* **2010**, 70 (16), 2228–2236.

(161) Donaldson, K.; Aitken, R.; Tran, L.; Stone, V.; Duffin, R.; Forrest, G.; Alexander, A. Carbon Nanotubes: A Review of Their Properties in Relation to Pulmonary Toxicology and Workplace Safety. *Toxicol. Sci.* **2006**, 92 (1), 5–22.

(162) Pikula, K.; Chaika, V.; Zakharenko, A.; Markina, Z.; Vedyagin, A.; Kuznetsov, V.; Gusev, A.; Park, S.; Golokhvast, K. Comparison of the Level and Mechanisms of Toxicity of Carbon Nanotubes, Carbon Nanofibers, and Silicon Nanotubes in Bioassay with Four Marine Microalgae. *Nanomaterials* **2020**, 10 (3), 485.

(163) Magrez, A.; Kasas, S.; Salicio, V.; Pasquier, N.; Seo, J. W.; Celio, M.; Catsicas, S.; Schwaller, B.; Forró, L. Cellular Toxicity of Carbon-Based Nanomaterials. *Nano Lett.* **2006**, 6 (6), 1121–1125.

(164) Nagai, H.; Okazaki, Y.; Chew, S. H.; Misawa, N.; Yamashita, Y.; Akatsuka, S.; Ishihara, T.; Yamashita, K.; Yoshikawa, Y.; Yasui, H.; et al. Diameter and Rigidity of Multiwalled Carbon Nanotubes Are Critical Factors in Mesothelial Injury and Carcinogenesis. *Proc. Natl. Acad. Sci. U. S. A.* **2011**, 108 (49), E1330–E1338.