

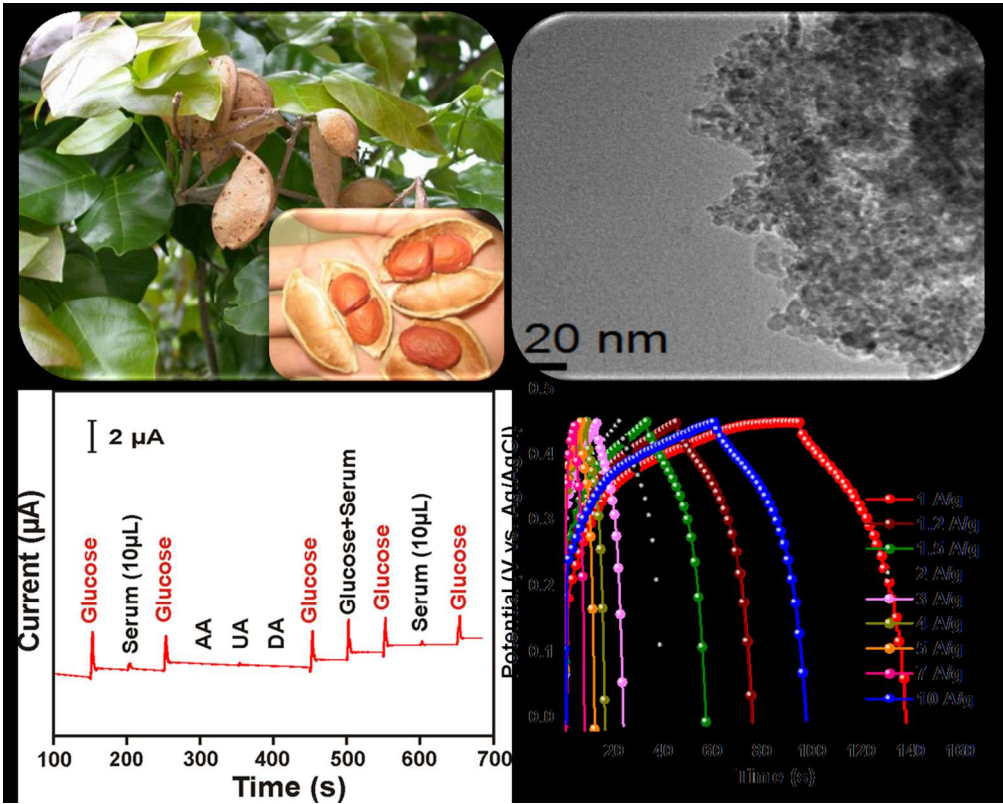
Article

Honeycomb-like Porous Carbon-Cobalt Oxide Nanocomposite for High-Performance Enzymeless Glucose Sensor and Supercapacitor ApplicationsRajesh Madhu, Vedyappan Veeramani, Shen-Ming Chen,
Arumugam Manikandan, An-Ya Lo, and Yu-Lun ChuehACS Appl. Mater. Interfaces, **Just Accepted Manuscript** • DOI: 10.1021/acsami.5b04132 • Publication Date (Web): 30 Jun 2015Downloaded from <http://pubs.acs.org> on July 1, 2015**Just Accepted**

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Synthesis of PSAC/Co₃O₄ nanocomposite towards the high-performance non-enzymatic glucose sensor and supercapacitor applications.
198x158mm (150 x 150 DPI)

Honeycomb-like Porous Carbon-Cobalt Oxide Nanocomposite for High-Performance Enzymeless Glucose Sensor and Supercapacitor Applications

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ABSTRACT Herein, we report the preparation of Pongam seed shells-derived activated carbon and cobalt oxide (~2-10 nm) nanocomposite (PSAC/Co₃O₄) by using a general and facile synthesis strategy. The as-synthesized PSAC/Co₃O₄ samples were characterized by a variety of physicochemical techniques. The PSAC/Co₃O₄-modified electrode is employed in two different applications such as high performance non-enzymatic glucose sensor and supercapacitor. Remarkably, the fabricated glucose sensor is exhibited an ultra-high sensitivity of 34.2 mA mM⁻¹ cm⁻² with a very low detection limit (21 nM) and long-term durability. The PSAC/Co₃O₄ modified stainless steel electrode possesses an appreciable specific capacitance and remarkable long-term cycling stability. The obtained results suggest the as-synthesized PSAC/Co₃O₄ is more suitable for the non-enzymatic glucose sensor and supercapacitor applications outperforming the related carbon based modified electrodes, rendering practical industrial applications.

KEYWORDS: pongam seed shells, activated carbon, cobalt oxide nanoballs, electrochemical glucose biosensor, supercapacitor.

INTRODUCTION

In recent decades, the biomass derived porous carbon materials is imperative to construct the environment-friendly energy storage devices and ultra-high performance sensors.¹⁻³ Diabetes mellitus is a group of metabolic diseases, and it is highly responsible for several major health problems. Hence, the diagnosis of glucose determination is domineering and several approaches have been devoted to construct the glucose biosensors based on the glucose oxidase enzyme. However, the enzyme immobilization on the electrode surface is one of the most challenging tasks, and it is highly affected by temperature, pH, humidity, which causes poor stability.^{4,5,6} Based on the above-mentioned problems, non-enzymatic glucose sensors have been widely explored and developed. As evidenced, graphene based metal oxides, such as NiO/Ni(OH)₂, CuO, ZnO, and Co₃O₄/Co(OH)₂, which exhibit higher sensitivity, lower detection limits for non-enzymatic glucose sensors as well as remarkable supercapacitor performance with high specific capacitance and excellent energy density which is necessary

for energy storage applications.^{3,5,7,8} In particular, cobalt oxide nanocomposites have been attracted due to their enhanced electro-catalytic behavior and good chemical stability, and have been extensively used in supercapacitors and enzyme-free glucose sensors applications.^{3,8-15} As well known, the preparation of graphene from graphite is a complicated task which may lead to a hazardous explosion while the oxidation of graphite. Besides, the procedure for the preparation of ACs is simple, eco-friendly and cost-effective, which leading to possess unique properties, such as higher surface area, modulated pore size, lower toxicity, excellent chemical stability, and presence of oxygen surface functional groups naturally.^{1,2, 15-17} Nonetheless, the availability of literature reports on biomass-derived ACs for the electrochemical applications is scarce. Moreover, pongam tree parts are widely used as efficient antibiotic, and also used to cure several types of skin and health problems. In particular, pongam seed shells are natural biomass, abundant, easily available, and there is no report available for the preparation of activated carbon using as precursor.

Fascinatingly, we report the preparation of pongam seed shells derived activated carbon (PSAC), and PSAC/Co₃O₄ nanocomposite for the first time. Owing to the outstanding physico-chemical properties of Pongam seed shells activated carbon (PSAC),

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hence, we first sought to explore the electrochemical performances of PSAC/Co₃O₄ nanocomposites in two different potential applications: sensitive enzyme-free glucose biosensor and supercapacitor applications. Herein, PSAC/Co₃O₄-modified glassy carbon electrode (GCE) is used for the enzyme-free glucose sensor, which exhibits remarkable analytical parameters, viz. favourable lower over-potential, low detection limits, ultra-high sensitivity. In addition, fabricated supercapacitor offers a good specific capacitance and long-term stability, which surpasses several carbon based materials.

RESULTS AND DISCUSSION

Figure. 1A,B displays the SEM images of the as-synthesized PSAC samples at different magnifications, which demonstrates the honeycomb structure-like highly porous morphology. Furthermore, Figure. 1C, shows the SEM image of the PSAC/Co₃O₄

nanocomposite sample, as can be seen, the Co₃O₄ are in the sphere form and they firmly covered by carbon layers with the porous nature (inset of Figure. 1C). The XRD patterns of the as-synthesized PSAC and PSAC/Co₃O₄ samples as shown in the inset of Figure. S1. As shown, the XRD pattern of PSAC demonstrates the two broad diffraction peaks at 23° and 43° which is corresponding to the (002) and (101) plane reflection, and it reveals the amorphous behavior of graphitic carbon.¹ The XRD spectrum of PSAC/Co₃O₄ represents the positions and relative intensities diffraction peaks at 2θ values denotes the pure face centered cubic (fcc) phase of the spinel Co₃O₄ structure with average crystallite size is about ~12 nm with lattice constant is 0.8018 nm, which is good agreement with the literature values (JCPDS 42-1467).^{3,18} In addition, there is no impurity peaks observed signifying the high purity behavior of as-prepared PSAC/Co₃O₄ nanocomposite.

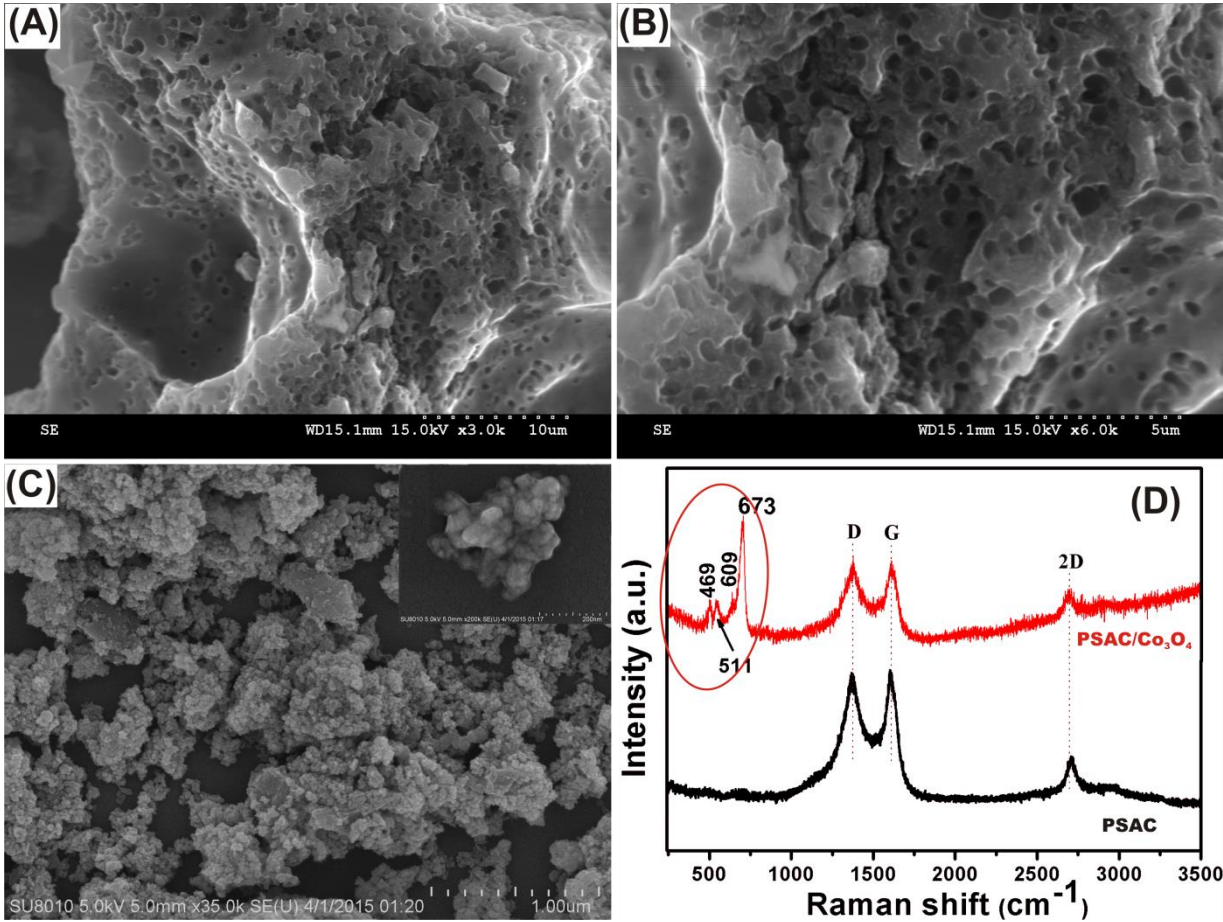


Figure. 1 (A,B) SEM images of PSAC, (C) SEM images of PSAC/Co₃O₄ nanocomposite; Inset: Higher magnification image of PSAC/Co₃O₄. (D) Raman spectra of PSAC and PSAC/Co₃O₄.

Figure. 1D depicts the Raman spectra of the PSAC and PSAC/Co₃O₄ samples, while PSAC possesses two characteristic

peaks at 1340 cm⁻¹ and 1590 cm⁻¹ corresponding to the disordered carbon of D band and E_{2g} mode phonons of the sp² graphitic carbon

G band, respectively. The nanocomposite PSAC/Co₃O₄ exhibits with four additional peaks (469, 511, 609, 673), which related to the vibrational modes of crystalline Co₃O₄ Eg, F2g¹, F2g² and Ag¹, respectively. Furthermore, the intensity ratio (I_D/I_G) of the PSAC value is 0.99 and PSAC/Co₃O₄ nanocomposites have 1.01, which is nearly same to PSAC and it may be due to the existence of Co₃O₄ nanoparticles on the PSAC surface. The above morphological and

structural characterization results evident the fruitful hybridization of PSAC/Co₃O₄ nanocomposite.

Further characterization by high resolution-transmission electron microscopy (HR-TEM) displays the honeycomb-like mesoporous structure of PSAC (Figure. 2A) and formation of PSAC/Co₃O₄ nanocomposite at different magnifications (Figure. 2B-D).

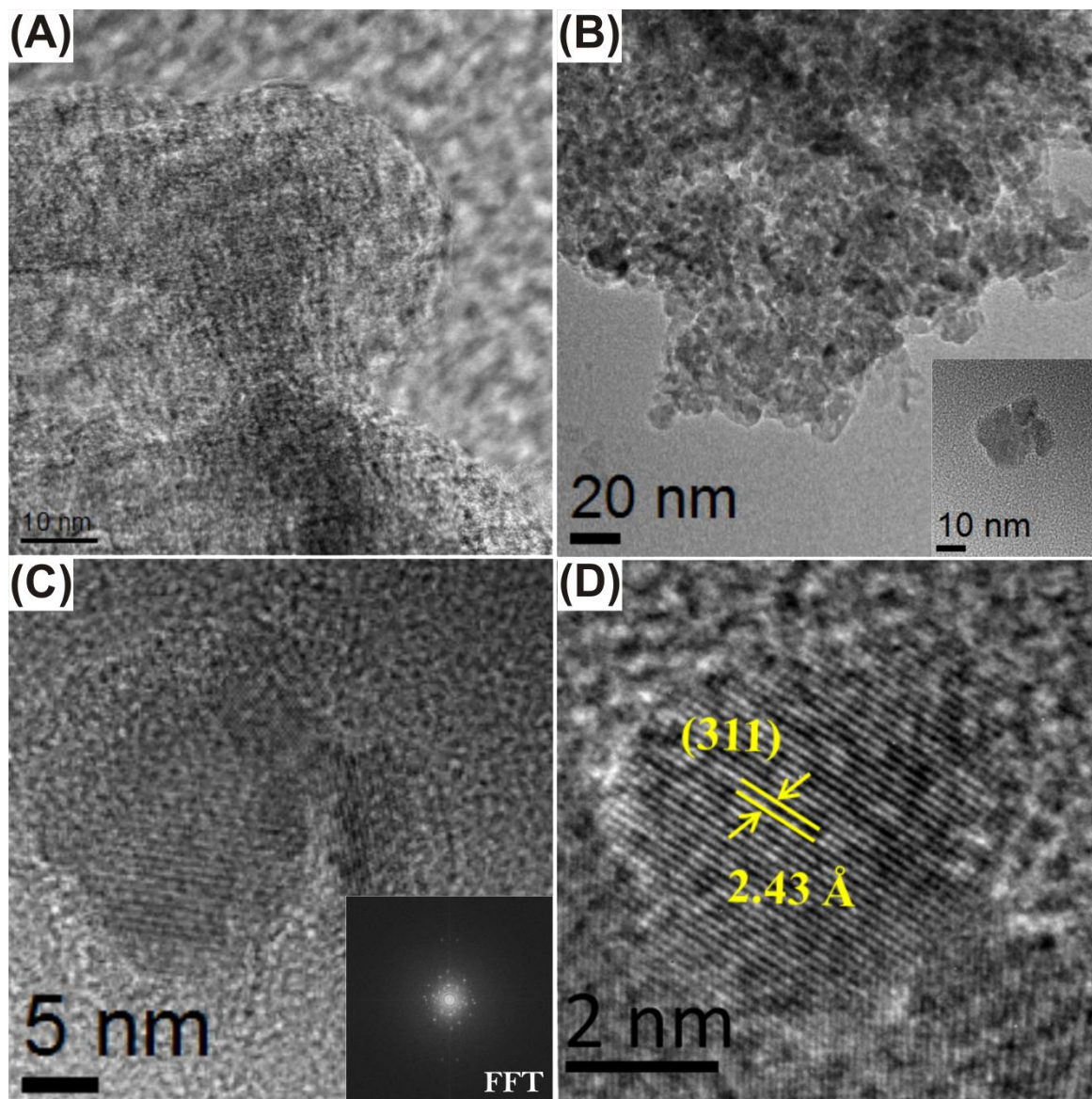


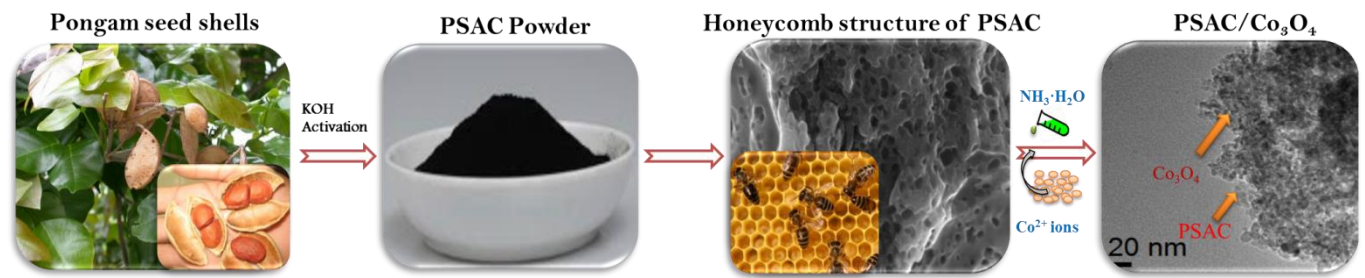
Figure. 2 (A) shows the Hr-TEM images of PSAC, and (B-D) PSAC/Co₃O₄ nanocomposite at various magnifications.

Obviously, the Co₃O₄ nanoparticles with diameter ranging from 2 to 10 nm (relatively similar with XRD results) were well-dispersed in the worm-like mesoporous structure. In addition, the HRTEM image shows the lattice distance around 2.43 Å which corresponds to (311) plane. The growth mechanism of the

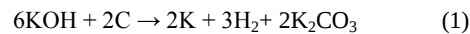
PSAC/Co₃O₄ nanocomposite is proposed based on the observed experimental results. Scheme 1 shows the schematic illustration for the formation of Co₃O₄ nanoparticles on the surfaces of PSAC. Briefly, the specific surface area (SSA) (~700 m² g⁻¹), with the highly porous nature of PSAC has prepared by using a general and

simple KOH activation method using cost-free and abundant biological waste i.e. pongam seed-shells. In agreement with our previous reports,^{1,2} during the activation process, creation of large pore volumes and oxygen containing functional groups were formed

on the PSAC surface. The proposed mechanism can be written as:^{18,19}



Scheme 1. Schematic illustration for the formation of Co₃O₄ nanoparticles on the honeycomb-like porous surface of PSAC.



The KOH activation is highly necessary to increase the surface area and create the pores, and also it helps to form the various oxygen surface functional groups as discussed in our recipe

report in detail.^{1,2} Mainly, these groups such as hydroxyl (C-OH) and carbonyl/quinone (C=O) were possibly involved in the formation of Co₃O₄ nanoparticles anchored on the negatively charged PSAC surface as shown in Figure S2, and Figure 3. The desired molar concentration of CoCl₂ was added to the black color PSAC solutions and the Co²⁺ ions which selectively bonded with epoxy and hydroxyl groups through the electrostatic interactions.

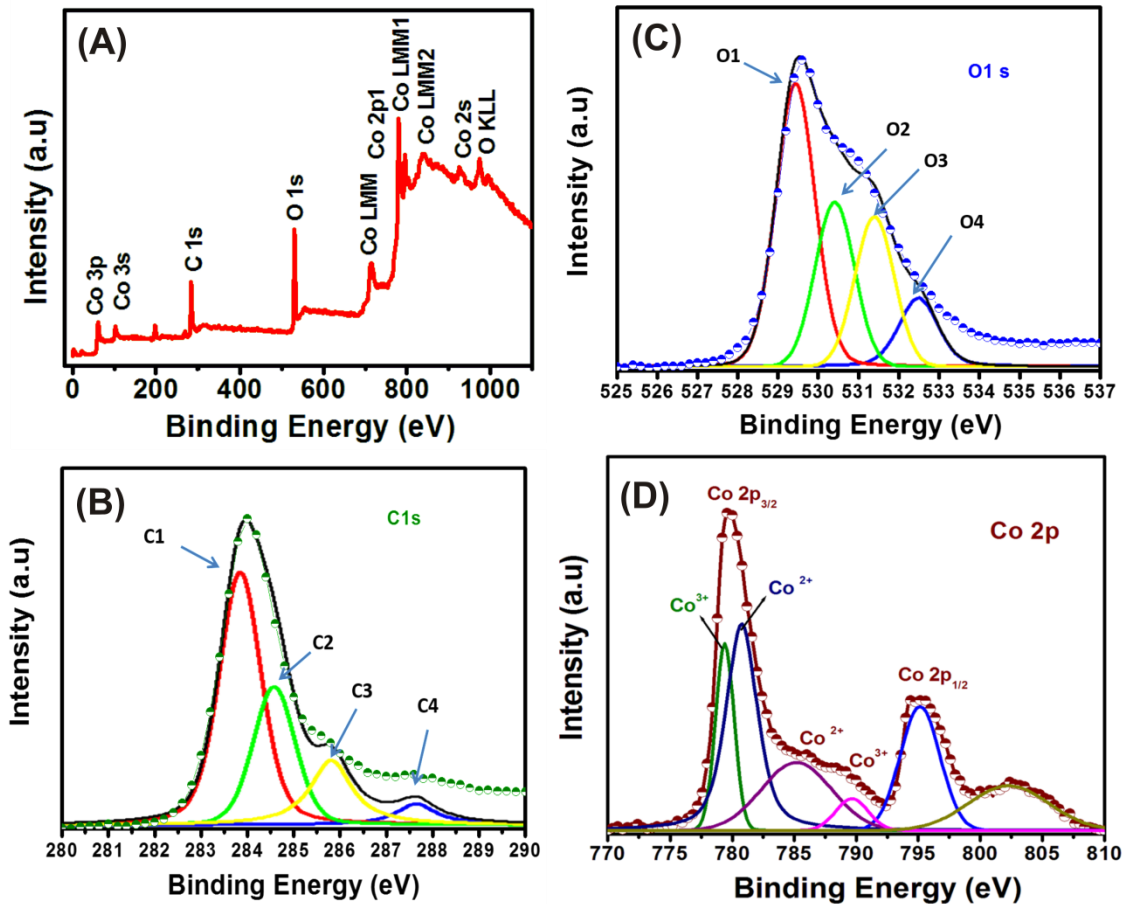


Figure. 3 (A) XPS survey spectra of PSAC/Co₃O₄ nanocomposite, (B-D) XPS core level spectra of C 1s, O 1s, and Co 2p of PSAC/Co₃O₄ nanocomposite.

The presence of the oxygen functional groups on the negatively charged PSAC has a higher capability to absorb the positively charged metal ions (i.e. Co²⁺) via electrostatic interactions, and act as nucleation sites. The Co²⁺ ions were easily diffuse and prefer the Co(OH)₂ on the PSAC surface. After hydrothermal treatment Co(OH)₂ was turned into Co₃O₄ nanoparticles and started to the crystallization growth along with the surface of PSAC. As a result, the Co₃O₄ nanoparticles was formed on the surface of PSAC support during the hydrothermal reaction. Hence, the larger amount of functional groups acting as an electron-donating source to provide a larger amount with smaller size of nanoparticles on the PSAC surface.

Furthermore, XPS survey spectra (Figure. 3A) of the PSAC/Co₃O₄ sample displays the corresponding peaks of carbon, oxygen, and Co₃O₄. Figure. 3B-D shows the enlarged view of the

XPS spectra of C1s, O1s, and Co 2P_{3/2} and 2P_{1/2} orbit of PSAC/Co₃O₄ nanocomposite at the binding energies of 284, 529.6, 780.24 eV and 795.52 eV, respectively.^{2, 20} In the enlarged view of C1s region, we observed 4 peaks at 283.82, 284.5, 285.6, 287.4 eV, which corresponding to the C-H, C-C, C-OH, and carbonyl/quinone groups (C=O), respectively. Similarly, O1s exhibits 4 peaks at 529.5, 530.3, components are assigned to carbonyl/quinone (C=O), and 531.4, 532.6, which corresponds to carbonate structures on the surface, and hydroxyl, ethers (C-OH, C-O-C), respectively. Figure. 3D represents the core level spectra for Co2p_{3/2} orbits fitted four peaks (Fig. 3D). The binding energies (BEs) of 779.5 and 785.6 eV represents the Co³⁺ and its satellite line and the BEs of 781.1 and 789.7 eV relates to Co²⁺ and its satellite line.²¹

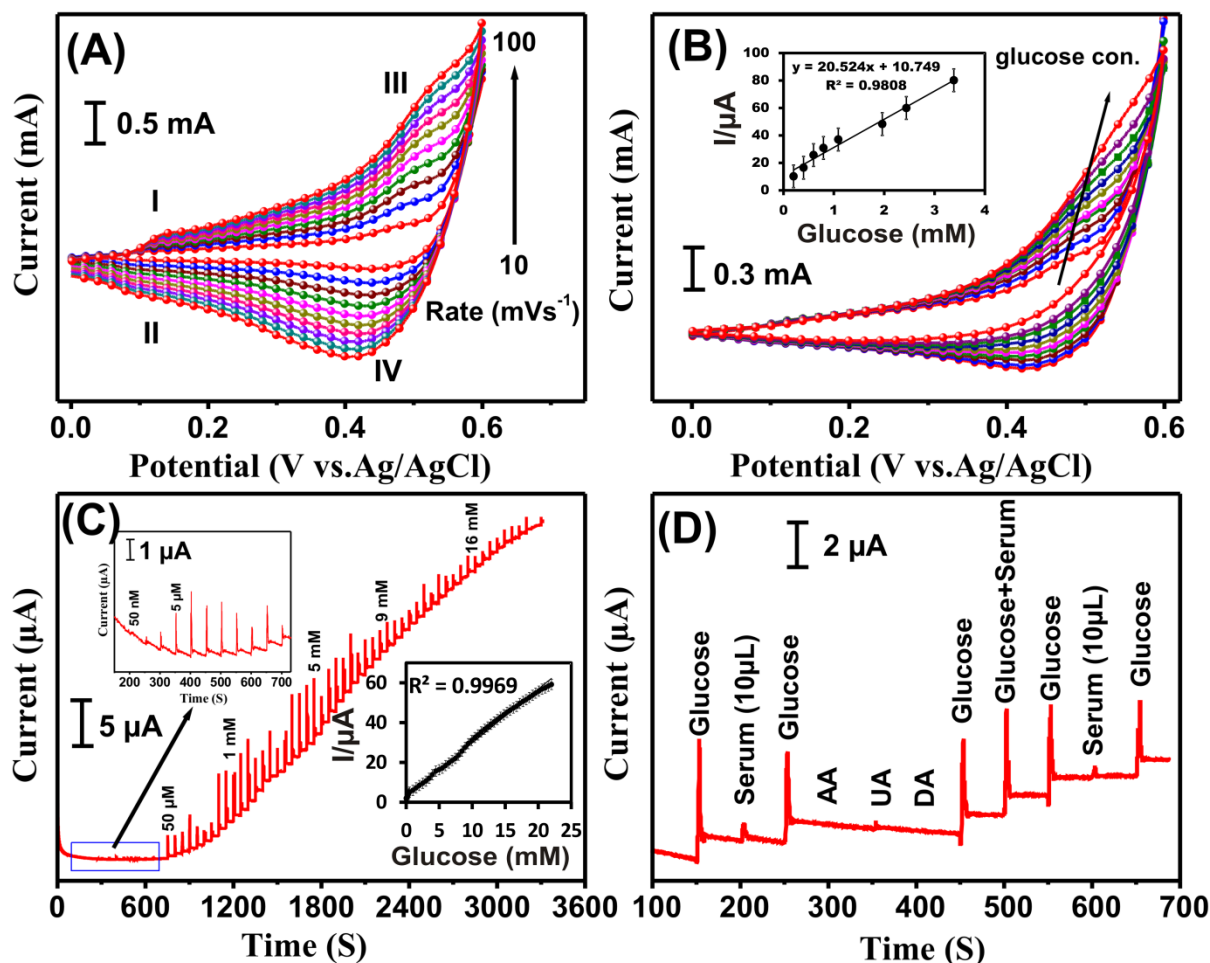


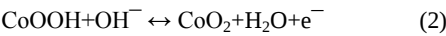
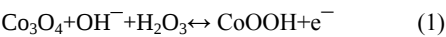
Figure. 4 (a) CVs obtained at PSAC/Co₃O₄-modified GCE at different scan rates in the presence of 1 mM glucose. (b) The CV curves at various concentrations of glucose (0-3.3 mM) at the scan rate of 50 mVs⁻¹. (c) The typical amperometric response of PSAC/Co₃O₄-modified GCE at different

concentration of glucose (0.05 to 22004 μM) and corresponding calibration plot (insert). (d) Influence of electroactive interferences (0.5 mM AA, 0.1 mM UA, 0.1 mM DA, and serum (10 μL) with and without glucose) on the response of 0.1 mM glucose. Supporting electrolyte: 0.1 M NaOH saturated with nitrogen gas; *app.* potential: +0.55 V.

To investigate the surface area and pore size distribution of the as-prepared PSAC and PSAC/Co₃O₄ samples, we analyzed Brunauer-Emmett-Teller (BET) N₂ adsorption-desorption isotherms at 77 K, and the obtained results are shown in Figure. S3. In agreement with previous reports, the observed typical Type-IV curves are denoting the existence of mesopores was revealed at P/P_0 in the range of 0.45–0.9. Compared to the pure-PSAC (700 m² g^{−1}, 0.47 cm³ g^{−1}), the PSAC/Co₃O₄ (164 m² g^{−1}, 0.34 cm³ g^{−1}) nanocomposite exhibits a lower surface area and pore volume, as expected because of the integration of Co₃O₄.

As shown in Figure. S5, Co₃O₄-free PSAC-modified GCE, does not show any corresponding glucose redox peaks, and only Co₃O₄ modified GCE shows a small response with featureless redox peak. Besides, while the introduction of Co₃O₄ on PSAC surface, we observed a two pair of redox peaks with enhanced peak currents. In addition, once the 1 mM glucose added into the 0.1 M NaOH, the corresponding oxidation peak currents of glucose were obviously increased.^[Eqn 1&2] Figure. 4A demonstrates the CVs of the PSAC/Co₃O₄ at different scan rates in the range of 10-100 mV/s in presence of 1 mM glucose in 0.1 M NaOH. Interestingly, we observed the two pairs of redox peaks of (I/II) corresponding to the reversible transition between Co₃O₄ and CoOOH, and (III/IV)

reveals the transition between CoOOH and CoO₂. Notably, we achieved a very lower oxidation potentials at 0.1 V (I) and 0.47 V (III), surpassing the several carbon based-cobalt nanocomposite electrodes.^{3,7,8,22-25} The possible reaction mechanism can be expressed in eqns. (1) and (2)



The redox peak currents were significantly increased with increasing scan rates in Figure S6 (A and B), which suggesting a surface-controlled electrochemical process. As shown in Figure. 4B, the glucose oxidation peak currents (III) were increased linearly with the increasing glucose concentration up to 3.3 mM ($R^2=0.9808$), and the peak currents were remained constant. The obtained result is more consistent with Ding’s report,²³ revealing the electrooxidation of glucose is mainly mediated by CoOOH/CoO₂ rather than Co₃O₄/CoOOH in alkaline solution.^{3,23} The superior results may be attributed to the synergistic effect between the as-synthesized PSAC and Co₃O₄ nanospheres, which includes the high surface area and modulated pore sizes with high-energy adsorption sites (BET) for the glucose oxidation provided by fast electron transfer channel offered by a simple eco-friendly fabrication method.

Table 1: Comparison of analytical parameters for detection of glucose over various modified electrodes

Electrodes	Linear range (mM)	Detection limit (μM)	Sensitivity ($\mu\text{A mM}^{-1} \text{cm}^{-2}$)	Ref.
graphene–Co ₃ O ₄ needle electrode	0.050 to 300	<10	-	3
3D Graphene-Cobalt Oxide	up to 0.080	<25 nM	3390	8
Cobalt oxide nanoparticles	0.7–60	0.15	2515.35	10
Electrospun Co ₃ O ₄ nanofibers/ Nafion/GCE	up to 2.04	0.97	36.25	23
Ni-Co NSs/RGO/GCE	0.010 to 2.65	3.79	1773.61	24
layered cobalt carbonate hydroxide (LCCH) / FTO	up to 3.5	0.058	571.8	25
Mesoporous Ni _{0.3} Co _{2.7} O ₄	0.001 to 2.55	1	0.206	26
Cobalt oxide/MWCNTs	0.15–5	70	-	27
PSAC/Co ₃ O ₄	0.05 μM -22 mM	0.02	34.23	This work

To attain the higher sensitivity, we have performed the amperometric i-t experiments at PSAC/Co₃O₄ nanocomposite modified electrode with the successive addition of glucose at different concentrations in 0.1 M NaOH solution (*app.* potential +

0.55 V) as shown in Figure. 4c. Fascinatingly, we obtained a wide linear range from 0.05 to 22004 μM , the linear equation can be expressed as $I (\mu\text{A}) = 2.7038x (\pm 0.063) + 2.7081$, ($R^2=0.9969$) (inset to Figure. 4c). The calculated detection limit and sensitivity is

21 nM and $34.2 \mu\text{A mM}^{-1} \text{cm}^{-2}$, respectively. Remarkably, we achieved a very low detection limit and ultra-high sensitivity towards detection of glucose, which surpassing the several carbon-based nanocomposite electrodes (Table 1).

To study the stability performance of non-enzymatic glucose sensor, 50 consecutive CV cycles were recorded in the absence and presence of 1 mM glucose in 0.1 M NaOH. Fascinatingly, there is no obvious peak current changes were observed. Furthermore, we have performed the long-time stability of the reported glucose sensor in 1 mM glucose in 0.1 M NaOH, and its oxidation peak current response was monitored periodically. The sensor retains about 97.2 % of its initial oxidation peak current response after 2 weeks in air at room temperature, reveals the good stability of the modified electrode for glucose determination. In order to assess the reproducibility, 6 independent modified electrodes were prepared for the determination of 1 mM glucose in 0.1 M NaOH, which exhibited a satisfactory reproducibility with a relative standard deviation (RSD) of 2.7%. In addition, the repeatability for 5 successive measurements with the RSD of 3.2%

for the determination of 1 mM glucose in 0.1 M NaOH reveals a good repeatability of the proposed sensor. Furthermore, the real-time application and specificity are the most important parameters for glucose sensor. Figure. 4d demonstrate the selectivity behavior of PSAC/ Co_3O_4 nanocomposite for glucose sensor, and it is inactive towards ascorbic acid, uric acid, dopamine (common contaminants in the blood) and $10 \mu\text{L}$ of glucose-free serum sample (contains a large variety of proteins and other molecules). Besides, we observed an enhanced peak current response for the addition of serum ($10 \mu\text{L}$) mixed glucose. Hence, the obtained results indicate that the PSAC/ Co_3O_4 modified electrode is vastly useful for glucose detection in real samples in the presence of foreign species. The PSAC/ Co_3O_4 modified GCE was further tested for the detection of glucose in human blood serum and urine sample (collected from a healthy man) by standard addition method with same experimental condition for three time measurements of each sample. In both the cases, good response is observed, as summarized in Table S2. The superior recoveries of enzyme-free glucose sensor in real samples designate that PSAC/ Co_3O_4 modified electrode is indeed reliable even in practical applications.

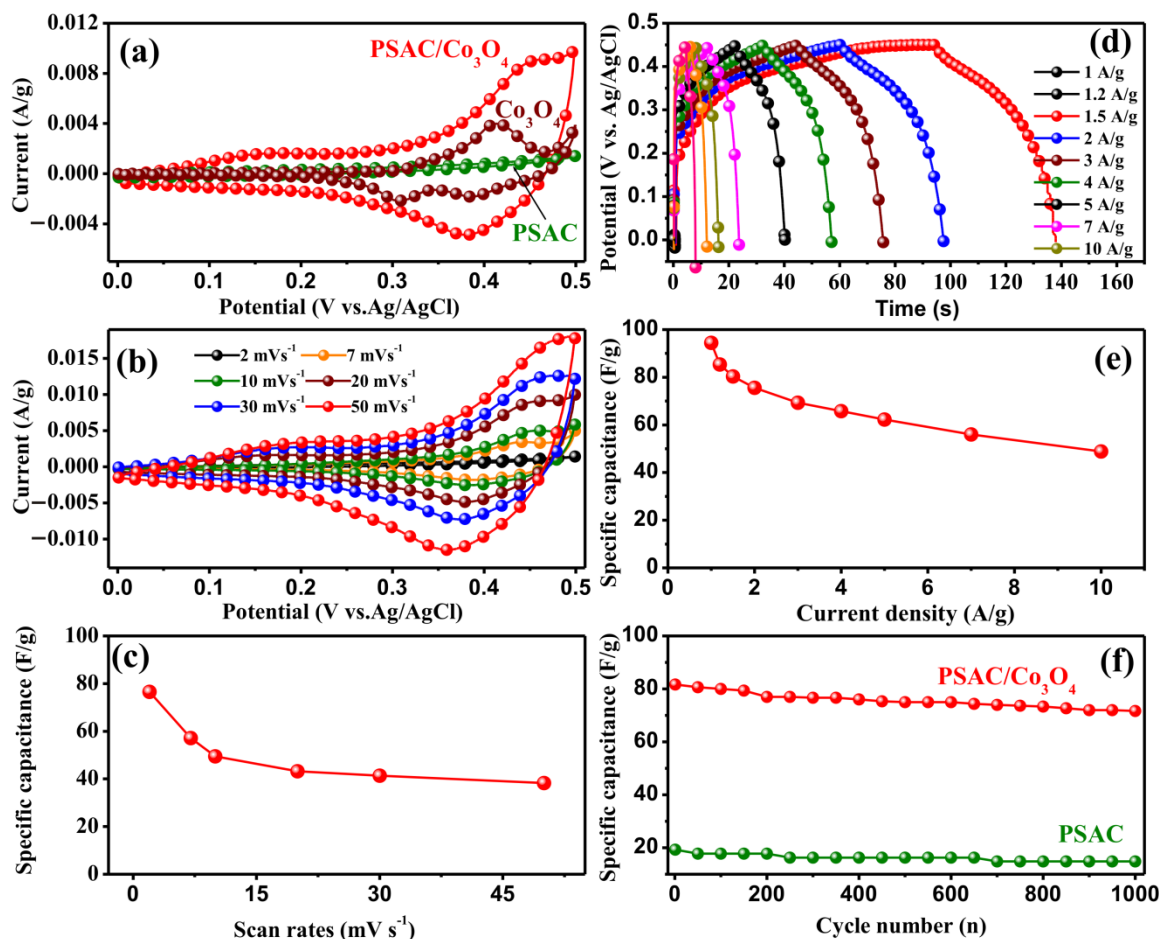


Figure 5 Electrochemical properties of PSAC/Co₃O₄-modified electrodes measured using a three-electrode system in 1.0 M KOH aqueous electrolyte. (a) The comparative CV curves for the PSAC, Co₃O₄ and PSAC/Co₃O₄ electrodes at 20 mVs⁻¹. (b) CV curves obtained at PSAC/Co₃O₄ electrode at different scan rates. (c) Calibration plot between different scan rates and specific capacitance. (d) Galvanostatic charge-discharge curves of PSAC/Co₃O₄ electrode at current density of 1, 1.2, 1.5, 2, 3, 4, 5, 7, and 10 Ag⁻¹. (e) Calibration plot between current density and specific capacitance (f) Cycle performance of PSAC/Co₃O₄ electrode measured at a current density of 1.5 Ag⁻¹.

Additionally, we employed the as-synthesized PSAC/Co₃O₄ nanocomposite for the supercapacitor application. Figure 5a shows the CVs of PSAC, Co₃O₄ and PSAC/Co₃O₄ electrodes over a voltage range of 0 to 0.5 V at the scan rate of 20 mV s⁻¹ in 1 M KOH electrolyte. For PSAC modified electrode, we observed a small oxidation peak around 0.3 V with rectangular behavior, which corresponding to the presence of oxygen functional groups carbonyl (C=O), hydroxyl (C-OH), and ethers (C-OH, C-O-C), in agreement with XPS results.^{25,26} Fascinatingly, we observed a superior redox peak current area of the CV curve with the introduction of Co₃O₄ on PSAC surface. The *ca.* specific capacitance values are 20, 47 and 80 F g⁻¹ for PSAC, Co₃O₄ and PSAC/Co₃O₄, respectively, which reveals that the specific capacitance was increased by pseudocapacitance of PSAC/Co₃O₄ nanocomposite. Moreover, the redox peaks of the CVs (Figure. 5b,c) at PSAC/Co₃O₄ fabricated electrode were maintained even at higher scan rates (2 to 50 mV s⁻¹) using 1.0 M KOH, which further revealed the good electrochemical reversibility, and the redox peak currents are proportional to scan rates, screening the process is surface controlled. This can be attributed to the pseudocapacitive contribution from the oxygen surface functional groups,^{2,26} which can improve the wettability and maximize the electroactive surface area.²⁵ Figure. 5d shows the galvanostatic charge-discharge (GCD) behavior of PSAC/Co₃O₄ electrode under varied current densities. The *ca.* specific capacitances of the PSAC/Co₃O₄ electrode are to be 48, 56, 62, 65, 69, 76, 80, 85 and 94 F/g at the current densities of 10, 7, 5, 4, 3, 2, 1.5, 1.2 and 1 A/g, respectively (Figure. 5e). The obtained specific capacitance of our PSAC/Co₃O₄ electrode is quite comparable several reported carbon based electrodes as shown in Table S2.^{8,12-15,28-30}

Figure. 5f displays the cyclic stability of the PSAC/Co₃O₄ electrode at a current density of 1.5 A g⁻¹ up to 1000 cycles. Interestingly, the specific capacitance of the PSAC/Co₃O₄ electrode only decreases up to 12% of its initial capacitance even after 1000 charge discharge cycles and stays stable afterward. For comparison, we have performed the charge-discharge experiments at PSAC, PSAC/Co₃O₄ electrodes as shown in Figure. S8. For PSAC electrode, we observed a good capacitive retention, which is more than graphene electrode,³¹ while the introduction of Co₃O₄ on the

PSAC surface, the GCD profiles were observed with a small potential plateau with higher capacitance and even after a long term cycles. For the practical applications, capacitive retention is more important in several portable electronic power tools and vehicles. In general, when the discharge current density is higher for the electrode with a lower specific capacitance is due to the occurrence of Faradaic reaction resistance and ion diffusion resistance. Hence, the obtained results establish that our PSAC/Co₃O₄ electrode has provided a good specific capacitance and cycle stability, outstanding the several carbon based cobalt oxide nanocomposite electrodes.

In addition, we have performed the electrochemical spectroscopy (EIS) experiments to analyze the electrochemical kinetics at the electrode/electrolyte interface. The Nyquist plots obtained from PSAC and PSAC/Co₃O₄ samples (Figure 6) show the well-defined semicircle in high-medium frequency region accompanied by a vertical line in the low frequency region, which is more consistent with several previous reports.³² As known, the semicircle at higher frequency region denotes the equivalent series resistance (ESR), and it is a combination of solution resistance, and intrinsic resistance of active material, which occurs by the ionic conductivity of the electrolyte ions, and it consists of a parallel combination of constant phase element (CPE) and a series of interfacial resistance and current collector.

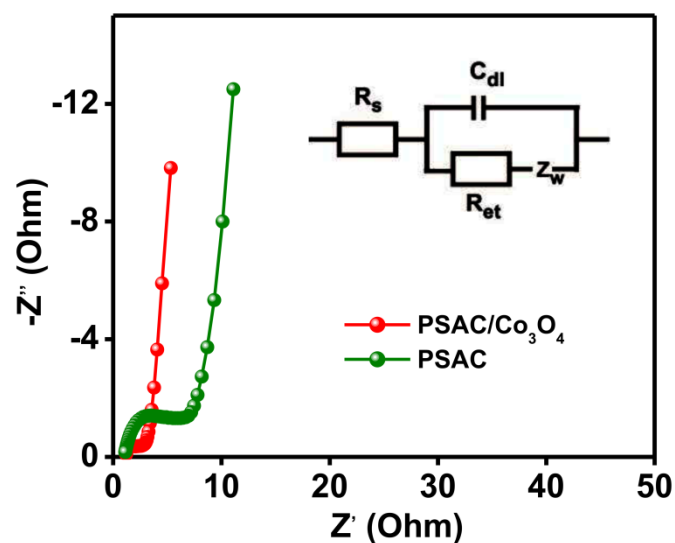


Figure 6. Nyquist plots observed for the PSAC and PSAC/Co₃O₄ samples.

Moreover, the diameter of the semicircle signifies the charge transfer resistance at electrode/electrolyte interfaces. Compared to the PSAC, the PSAC/Co₃O₄ material displayed a lower charge transfer resistance, because of the synergistic effect between the PSAC and Co₃O₄ components.³² Hence, the obtained analytical parameters suggest that the PSAC/Co₃O₄ nanocomposite is a promising electrode material for non-enzymatic glucose sensors and supercapacitors, rendering practical industrial applications.

CONCLUSIONS

In summary, PSAC/Co₃O₄ nanocomposite is synthesized by using a simple and eco-friendly method. Notably, the as-synthesized nanocomposite offers a remarkable performance towards the reported enzyme-free glucose sensor and supercapacitor application. The achieved electrochemical performance may be ascribed to the catalytic activity of the Co₃O₄ with a strong substrate of high surface area PSAC materials, which can be used for the practical industrial applications.

EXPERIMENTAL METHODS

Materials. Potassium hydroxide, Sodium hydroxide, Cobalt chloride hexahydrate (CoCl₂·6H₂O) and ethanol were purchased from Sigma Aldrich. Ammonium hydroxide water (NH₃·H₂O) and HCl purchased from J.T. BAKER chemicals. All the chemicals were used without further purification. The experiments were carried out by using doubly distilled water.

Synthesis of PSAC/Co₃O₄ nanocomposite. The high-surface-area (~700 m² g⁻¹), with the highly porous nature of PSAC has prepared by using a general and simple KOH activation method using no cost and abundant biological waste i.e. pongam seed-shells (*Pongamia pinnata*). Briefly, the sun-dried pongam seed-shells were washed and then dried in an oven at 100°C. The pulverized seed-shells were pre-carbonized at 350°C for 2 h in a N₂ flow tubular furnace; the collected pre-carbonized black solid was added to 10% of KOH solution under N₂ flow while stirring at 80°C. The activated slurry was placed in an open atmosphere for 24 h, subsequently transferred into a quartz crucible at 900°C (optimum) for 2 h in a N₂ atmosphere at a heating rate of 5°C min⁻¹ in a tube furnace. The carbonized samples were washed with HCl (5%) and double-distilled water until the pH became neutral, and dried at 100°C for 24 h to remove the moistures and thoroughly ground to yield the finest carbon powder.

The PSAC/Co₃O₄ nanocomposite was synthesized by using a simple and eco-friendly method. The PSAC/Co₃O₄ nanocomposites were prepared by hydrothermal method. In this process, 30 mg of PSAC powder was suspended in 30 ml of deionized water by ultrasonic dispersion for 30 min to form a black colour solution and the supernatant was collected for further process. Further, 30 mM of CoCl₂ was dropped into 30 ml of supernatant PSAC solutions and stirred for 50 min. Subsequently, the aqueous solution of ammonium hydroxide water (NH₃·H₂O) was added dropwise into the above suspension until the pH of the solution reached 10. Finally, the mixed solution was transferred to the Teflon-lined stainless steel

autoclave. The autoclave was put in an oven at 200°C for 12 h and slowly cooled down to room temperature naturally. The black colour precipitate was obtained and washed with several times ethanol and followed by water. The as-prepared PSAC/Co₃O₄ nanocomposites were obtained after being dried at 70°C under a vacuum. Finally, the resulting samples were calcined at 250 °C for 5 h under an argon atmosphere.

Materials Characterization. The surface morphology of the film was studied using JEOL field-emission scanning electron microscopy, and the nanostructures were examined by field emission transmission electron microscopy (FE-TEM, JEM-3000F, JEOL operated at 300 kV for high resolution-transmission electron microscope (HR-TEM). X-ray diffraction (XRD) was performed on a Rigaku, MiniFlex II instrument. The Raman spectra were recorded at ambient temperature using a WITeCK CRM200 confocal microscopy Raman system with a 488 NM laser. N₂ adsorption-desorption isotherms and the pore size distribution were studied using “Micromeritics ASAP 2020.” For the X-ray Photoelectron Spectroscopy, “XPS ULVAC-PHI PHI 5000 VersaProbe”.

Preparation of Modified Electrodes. For the electrochemical non-enzymatic glucose sensor, the as-synthesized PSAC/Co₃O₄ sample was first dispersed in ethanol under sonication treatment for 2 h. Meanwhile, the surface of the GCE was carefully polished; ca. 6 µL of dispersed PSAC (with optimal concentration, see Figure. S7) was drop casted on the GCE, followed by drying in an air oven at 30°C.

In order to prepare the supercapacitor electrode, the as-synthesized PSAC/Co₃O₄ active electrode material, 80 wt% of PSAC/Co₃O₄, 15 wt% of graphite is mixed with 0.4 ml of N-methylpyrrolidinone (NMP) to form homogeneous slurry. Further 15 µL of slurry is coated on the 1x1 cm² area of stainless steel electrode by drop casting method and dried overnight at 60°C. The mass loading of PSAC/Co₃O₄ on the substrate is 0.8 mg cm⁻². The cell was soaked in electrolyte for a few hours before undergoing the electrochemical study.

Electrochemical Measurements. For non-enzymatic glucose sensor applications, CV and Amperometry i-t profiles were recorded by using an electrochemical analyzer (CH instruments, CHI900) at room temperature using a conventional three-electrode cell consisting of a GCE as the working electrode, Ag/AgCl (saturated KCl) as the reference electrode, and a platinum wire as the counter electrode.

For supercapacitor applications, PSAC and PSAC/Co₃O₄ electrodes were assessed by three electrode system within the potential range of 0 to 0.5 V in 1.0 M KOH electrolyte solution. All the cyclic voltammetry (CV) and galvanostatic charge-discharge (GCD) profiles were performed by using the CHI627 electrochemical analyzer. The specific capacitances of the sample electrodes were calculated by using the following equations:³³

In the case of CV measurements:

$$C = Q/m\Delta V \quad (3)$$

where, C signifies the specific capacitance (unit of F g⁻¹), Q is the average charge during charge-discharge process (unit of C), m - mass of the active material (unit of g), and ΔV - working potential (unit V).

For GCD measurements:

$$C = I\Delta t/m\Delta V \quad (4)$$

where, I - current density (unit A), Δt - charge/discharge duration (unit s).

Conflict of Interest: The authors declare no competing financial interest.

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Supporting Information Available.: BET surface area analysis, BJH pore size distribution, and FTIR spectra CV results. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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